



## GLUCAGON GIPR LABELED CELLS

**Glucagon GIPR labeled cells for:** 200 tests

**Part#:** C1TT1GIPR

**Rev:** #02 of September 2023

**Store at:** -80°C or below, see expiration date on package label.

**For research use only. Not for use in diagnostic procedures.**

### ASSAY PRINCIPLE

The Tag-lite Glucagon GIPR cells transiently expressing the Glucagon GIPR receptor are labeled with Terbium for conducting receptor binding studies on the aforementioned receptor.

The Tag-lite® Glucagon GIPR Receptor Ligand Binding Assay is a homogeneous alternative to radio ligand binding assays for HTS and compound profiling.

It is suitable for both saturation binding assays ( $K_d$ ) and competitive binding assays ( $K_i$ ). At equilibrium, the fraction of labeled ligand bound to the receptor is proportional to the FRET signal recorded. From this resulting signal, binding affinities can be calculated.

### MATERIALS & EQUIPMENT

#### MATERIALS PROVIDED:

- Tag-lite Glucagon GIPR labeled Cells, ready-to-use (transformed & labeled), 200 tests\* (Part# C1TT1GIPR)

\*Sufficient for 200 tests tests using a 96 or 384-well small volume white plate (20  $\mu$ L). Purchase additional labeled cells for larger runs.

Notes:

1. Differences in  $K_d$  values may be observed between batches of labeled cells. Variability between  $K_d$  values reported in this package insert and values calculated during your experiment may also occur.
2. To ensure optimal reproducibility and consistency, single lot-bulk batches are available as a custom service. Our technical support team can help you set up this assay.

#### FOR $K_D$ AND $K_I$ DETERMINATION, PURCHASE SEPARATELY:

- Glucagon GIPR Receptor red agonist Fluorescent Ligand (Revvity Part# L0018RED)
- Tag-lite Buffer (5X concentrate), 100 mL (Revvity Part# LABMED)
- Unlabeled ligand to measure non-specific signal: GIP - (recommended)
- Microplates - For HTRF microplate recommendations, please visit [www.revvity.com](http://www.revvity.com)
- HTRF®-Certified Reader - For a list of HTRF-compatible readers and setup recommendations, please visit [www.revvity.com](http://www.revvity.com)

Use of an inappropriate set-up may seriously impair results. Check that you are using the set-up for Tb donor and red ligand. HTRF-approved readers using a monochromator for detection are not compatible with Tag-lite binding assays.

### STORAGE AND HANDLING

Cells must be stored at -80°C or in liquid nitrogen until thawing. For storage > 1 month, store the frozen cells in liquid nitrogen.

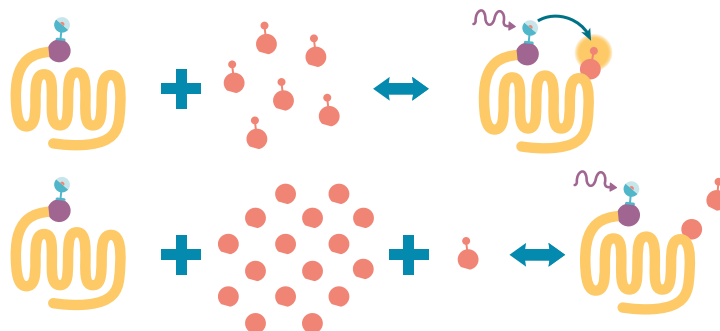
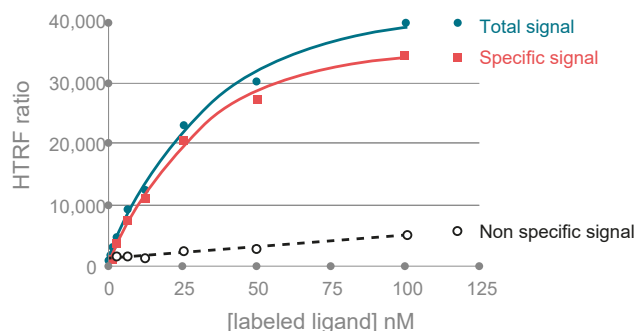
Keep the cells frozen until all the other reagents are ready.

A saturation binding assay measures total and non-specific binding of increasing concentrations of ligand under conditions of equilibrium.

To perform the assay, the fluorescent ligand is titrated into a solution containing a fixed amount of labeled cells and incubated to equilibrium. The HTRF ratio obtained from this titration is the total binding.

A negative control using unlabeled ligand is included to account for the non-specific binding of the labeled ligand to the receptor, non-receptor molecules, and the microplate. The fluorescent ligand is titrated into a solution containing a fixed amount of labeled cells and a 100-fold molar excess of unlabeled ligand. The HTRF ratio obtained from this titration is the non-specific binding.

The specific binding is calculated by subtracting the non-specific binding from the total binding at each fluorescent ligand concentration.



## REAGENT PREPARATION

### Step 1: prepare working Tag-lite buffer (1X TLB).

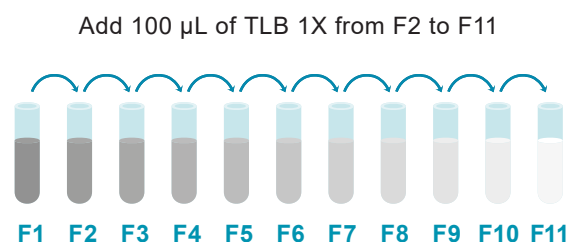
1. Determine the amount of 1 X TLB needed for the assay.
2. Thaw the 100 mL vial of Tag-lite buffer 5X (5X TLB).
3. Dilute 5-fold the 5X TLB in distilled water to prepare 1X TLB.
4. Mix gently.

### Step 2: prepare fluorescent ligand.

The concentration of fluorescent ligand provided (Glucagon GIPR receptor red agonist) is indicated on the vial label.

1. Centrifuge the vial
2. Dilute labeled ligand stock solution using 1X TLB to obtain the highest concentration **F1 = 400 nM** for the saturation binding curve.  
Use the following formula  $C_1V_1 = C_2V_2$  to calculate the final volume needed to produce the **400nM** solution  
 $V_1 = (C_2 \times V_2) / C_1$ , where  $V_1$  is the final volume needed,  $C_1$  is the labeled ligand stock concentration,  $C_2$  is the labeled ligand desired concentration of 400nM, and  $V_2$  is the volume of stock labeled ligand.  
Example for a ligand concentration  $C_1 = 10\,000\text{ nM}$ . Take  $8\text{ }\mu\text{L}$  ( $V_1$ ) of fluorescent ligand stock solution and add  $192\text{ }\mu\text{L}$  of 1X TLB in order to obtain  $200\text{ }\mu\text{L}$  ( $V_2$ ) of  $400\text{ nM}$  ( $C_2$ ) solution
3. Starting with the F1 solution (400 nM), prepare 1/2 serial dilutions in 1X TLB.
4. Add  $100\text{ }\mu\text{L}$  of F1 to  $100\text{ }\mu\text{L}$  of 1X TLB, mix gently and repeat the 1/2 serial dilutions to prepare 200-0.4 nM solutions.

RECOMMENDED DILUTION PROCEDURE FOR FLUORESCENT LIGAND		FLUORESCENT LIGAND CONCENTRATION (nM)	
		INITIAL CONCENTRATIONS (WORKING SOLUTIONS)	FINAL CONCENTRATIONS (IN WELL)
F1	Made from stock solution	400	100
F2	100 $\mu\text{L}$ F1 + 100 $\mu\text{L}$ 1X TLB	200	50
F3	100 $\mu\text{L}$ F2 + 100 $\mu\text{L}$ 1X TLB	100	25
F4	100 $\mu\text{L}$ F3 + 100 $\mu\text{L}$ 1X TLB	50	12.5
F5	100 $\mu\text{L}$ F4 + 100 $\mu\text{L}$ 1X TLB	25	6.3
F6	100 $\mu\text{L}$ F5 + 100 $\mu\text{L}$ 1X TLB	12.5	3.1
F7	100 $\mu\text{L}$ F6 + 100 $\mu\text{L}$ 1X TLB	6.3	1.6
F8	100 $\mu\text{L}$ F7 + 100 $\mu\text{L}$ 1X TLB	3.1	0.8
F9	100 $\mu\text{L}$ F8 + 100 $\mu\text{L}$ 1X TLB	1.6	0.4
F10	100 $\mu\text{L}$ F9 + 100 $\mu\text{L}$ 1X TLB	0.8	0.2
F11	100 $\mu\text{L}$ F10 + 100 $\mu\text{L}$ 1X TLB	0.4	0.1






### Step 3: prepare unlabeled ligand

Prepare a working solution of unlabeled ligand GIP in 1X TLB at 100-fold the concentration of F1 solution = 40  $\mu$ M.  
Please refer to literature accompanying the unlabeled ligand for stock concentration provided.

### Step 4: prepare cells

1. Prepare a conical vial containing 5 mL of cold 1X TLB.
  2. Thaw labeled frozen cells (1 vial) in a 37°C water bath - **manual shaking** - until all the ice is thawed (1-2 min).
  3. Quickly transfer the cells by pipetting into the conical vial containing 1X TLB.
  4. Centrifuge 5 min at 300 G. *The pellet may not be visible.*
  5. Gently remove supernatant by aspiration. **Do not pour out the supernatant.**
  6. Resuspend the pellet in 1 mL of 1X TLB. Mix gently by pipetting up and down several times.
  7. Add 1.7 mL of 1X TLB. Mix gently by pipetting up and down several times.
- Keep the cells at room temperature.

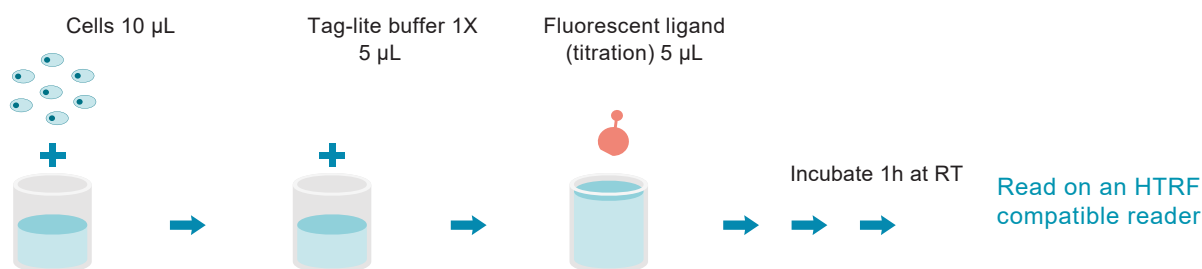
Step 1	Step 2	Step 3	Step 4	Step 5
Prepare a conical vial (A) containing 5 mL of cold 1X TLB	Thaw labeled frozen cells (1vial) at 37°C (water bath, manual shaking) until all the ice is thawed (1-2 min) and transfer them quickly by pipetting into the vial prepared in Step 1.	Centrifuge 5 min at 300 G.  Be careful the pellet may not be visible.	Gently remove supernatant by aspiration do not pour out supernatant. 	Resuspend the pellet in 1 mL of 1X TLB. Mix gently by pipetting up and down several times. Add 1.7 mL of 1X TLB. Mix gently by pipetting up and down several times. Keep the cells at R.T. 

## SATURATION BINDING ASSAY MANUAL

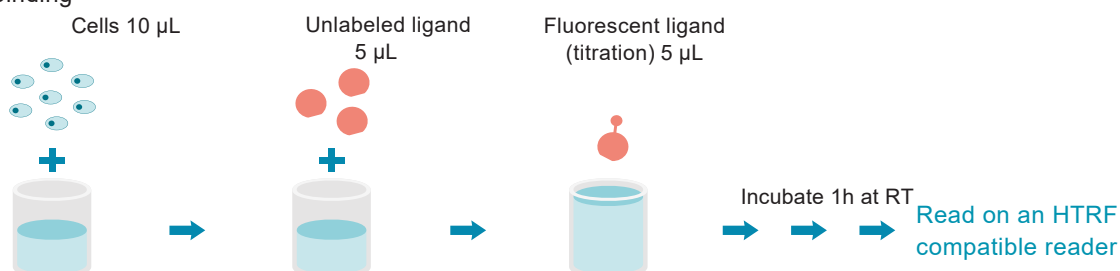
Run all assay points in triplicate. An example of plate map is indicated on page 4.

1. Dispense 10  $\mu$ L labeled cells into each well for both total and nonspecific binding.
2. Dispense 5  $\mu$ L 1X TLB into total binding wells.
3. Dispense 5  $\mu$ L unlabeled ligand (40  $\mu$ M) into nonspecific binding wells.
4. Dispense 5  $\mu$ L labeled ligand dilutions into each appropriate well.
5. Incubate 1h at room temperature.
6. Read on an HTRF-compatible reader - **HTRF-approved readers using a monochromator for detection are not compatible with Tag-lite binding assays.**

### Total binding



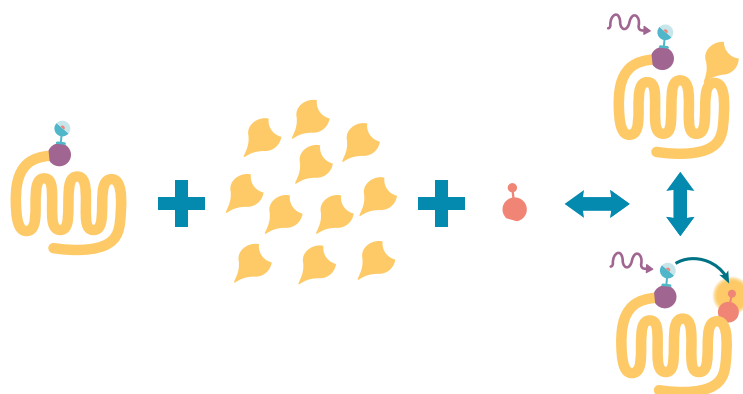
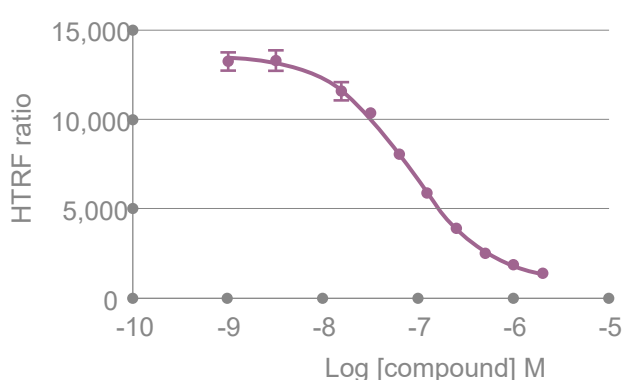
### Non specific binding



A large grid of 26 columns and 14 rows, labeled A through P vertically and 1 through 26 horizontally. A red rectangle highlights the first 6 columns and the first 14 rows (A through P).

## COMPETITION BINDING (K<sub>i</sub> DETERMINATION)

Competitive binding assay is performed to measure the dissociation constant, K<sub>i</sub>. To perform the assay, the compound is titrated into a solution containing a fixed concentration of fluorescent ligand and a fixed amount of cells.



### REAGENT PREPARATION

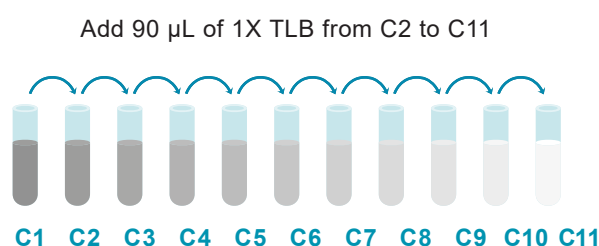
#### Step 1: prepare working tag-lite buffer (1X TLB).

1. Determine the amount of 1X TLB needed for the assay. (Approximately 10 mL is required to assay one compound + 1 mL for each additional compound.)
2. Thaw the 100 mL vial of Tag-lite buffer 5X (5X TLB).
3. Dilute 5-fold the 5X TLB in distilled water to prepare 1X TLB. (E.g. 10 mL of 5X TLB + 40 mL distilled water.)
4. Mix gently.

#### Step 2: prepare compounds

1. Dilute compounds with 1X TLB to an initial concentration of 4.E-04 M (C1).
2. Starting with the C1 solution (4.E-04 M), prepare 1/10 serial dilutions in 1X TLB.
3. Add 10  $\mu$ L C1 to 90  $\mu$ L of 1X TLB, mix gently and repeat the 1/10 serial dilutions to prepare C2, C3, C4, C5, C6, C7, C8, C9, C10, C11 solutions.

RECOMMENDED DILUTION PROCEDURE FOR COMPOUNDS		COMPOUND CONCENTRATIONS (M)	
		INITIAL CONCENTRATIONS (WORKING SOLUTIONS)	FINAL CONCENTRATIONS (IN WELL)
C1	Made from stock compounds	4.E-04	1.E-04
C2	10 $\mu$ L C1 + 90 $\mu$ L 1X TLB	4.E-05	1.E-05
C3	10 $\mu$ L C2 + 90 $\mu$ L 1X TLB	4.E-06	1.E-06
C4	10 $\mu$ L C3 + 90 $\mu$ L 1X TLB	4.E-07	1.E-07
C5	10 $\mu$ L C4 + 90 $\mu$ L 1X TLB	4.E-08	1.E-08
C6	10 $\mu$ L C5 + 90 $\mu$ L 1X TLB	4.E-09	1.E-09
C7	10 $\mu$ L C6 + 90 $\mu$ L 1X TLB	4.E-10	1.E-10
C8	10 $\mu$ L C7 + 90 $\mu$ L 1X TLB	4.E-11	1.E-11
C9	10 $\mu$ L C8 + 90 $\mu$ L 1X TLB	4.E-12	1.E-12
C10	10 $\mu$ L C9 + 90 $\mu$ L 1X TLB	4.E-13	1.E-13
C11	10 $\mu$ L C10 + 90 $\mu$ L 1X TLB	4.E-14	1.E-14



#### step 3: prepare fluorescent ligand






For the competition dose-response of compounds, the optimal fluorescent ligand concentration is the one that allows 50% (K<sub>d</sub>) to 80% of receptor binding.

The concentration of fluorescent ligand Glucagon GPCR receptor red agonist is indicated on the vial label<sup>40</sup>.

Centrifuge the vial then dilute the fluorescent ligand Glucagon GPCR receptor red agonist with 1X TLB

#### step 4: prepare cells

1. Prepare a conical vial containing 5 mL of cold 1X TLB.
2. Thaw labeled frozen cells (1 vial) in a 37°C water bath (manual shaking) until all the ice is thawed (1-2 min).
3. Quickly transfer them by pipetting into the conical vial containing 1X TLB.
4. Centrifuge 5 min at 300 G. (The pellet may not be visible.)
5. Gently remove supernatant by aspiration. (Do not pour out the supernatant.)
6. Resuspend the pellet in 1 mL of 1X TLB. Mix gently by pipetting up and down several times.
7. Add 1.7 mL of 1X TLB. Mix gently by pipetting up and down several times.
8. Keep the cells at room temperature

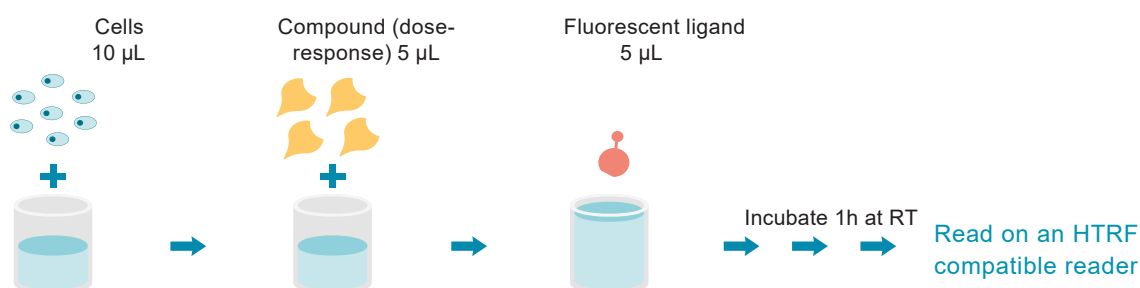
Step 1	Step 2	Step 3	Step 4	Step 5
Prepare a conical vial (A) containing 5 mL of cold 1X TLB 	Thaw labeled frozen cells (1 vial) at 37°C (water bath, manual shaking) until all the ice is thawed (1-2 min) and transfer them quickly by pipetting into the vial prepared in Step 1. 	Centrifuge 5 min at 300 G.  Be careful the pellet may not be visible.	Gently remove supernatant by aspiration do not pour out supernatant. 	Resuspend the pellet in 1 mL of 1X TLB. Mix gently by pipetting up and down several times. Add 1.7 mL of 1X TLB. Mix gently by pipetting up and down several times. Keep the cells at R.T. 

### COMPETITIVE BINDING ASSAY MANUAL

Run all assay points in triplicate. An example of plate map is indicated on page 7.

Up to ten (10) compounds can be tested in one 384-well plate for a total of 36 wells per compound.

1. Dispense 10  $\mu$ L labeled cells into each well.
2. Dispense 5  $\mu$ L 1X TLB or 5  $\mu$ L compound dilutions into each appropriate well as shown.
3. Repeat for each compound tested.
4. Dispense 5  $\mu$ L labeled ligand into each well.
5. Incubate 1h at room temperature.
6. Read on an HTRF-compatible reader - **HTRF-approved readers using a monochromator for detection are not compatible with Tag-lite binding assays.**





A 26x26 grid with columns numbered 1 to 26 and rows lettered A to Z. A red rectangle highlights the first three columns (1, 2, 3) and the first eight rows (A, B, C, D, E, F, G, H).

## DATA REDUCTION

1. Calculate the ratio of the acceptor and donor emission signals for each individual well.

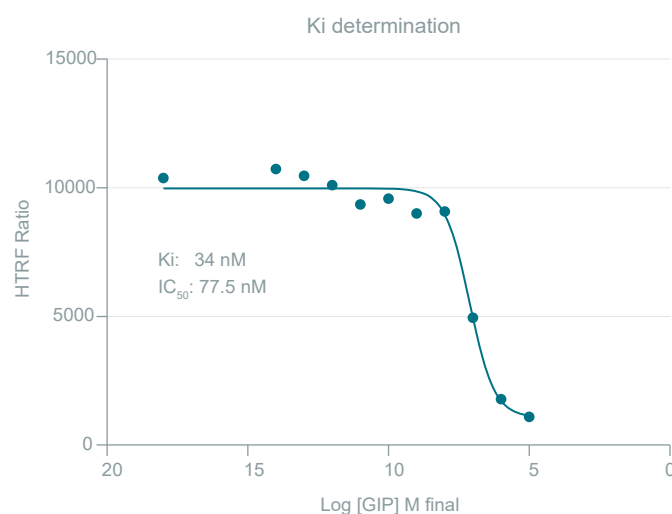
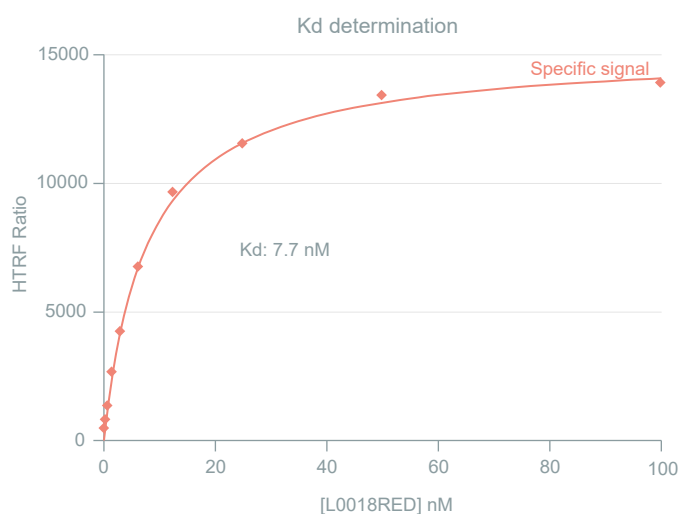
$$\text{Ratio} = \frac{\text{Signal 665 nm}}{\text{Signal 620 nm}} \times 10^4$$

2. Plot the HTRF ratio versus the [fluorescent ligand] or [compound] concentrations.

For more information about data reduction, please visit [www.revvity.com](http://www.revvity.com)

## RESULTS

Example of data obtained for Glucagon GIPR receptor with GIP as reference ligand and readings taken on a Infinite 500 with flash lamp. Results may vary from one HTRF® compatible reader to another.

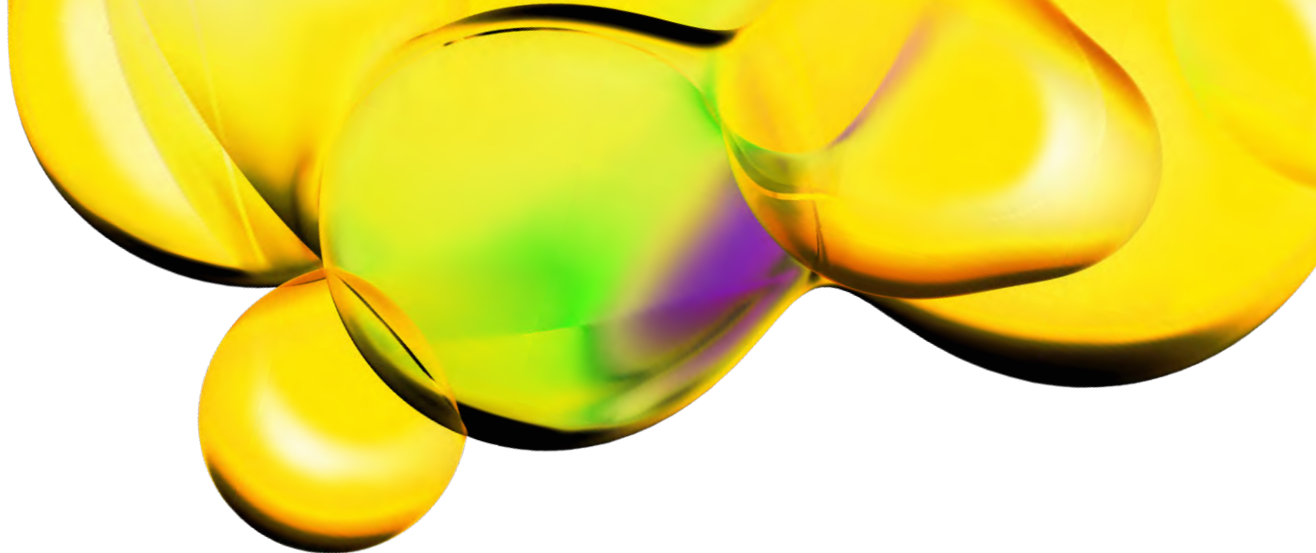


### Notes:

1. Differences in K<sub>d</sub> values may be observed between batches of labeled cells. Variability between K<sub>d</sub> values reported in this package insert and values calculated during your experiment may also occur.
2. To ensure optimal reproducibility and consistency, single lot-bulk batches are available as a custom service. Our technical support team can help you set up this assay.

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