

# ALDOSTERONE KITS

## Part # 64ALDPEG & 64ALDPEH

Test size#: 500 tests (64ALDPEG) and 10,000 tests (64ALDPEH) - assay volume: 20 µL

Revision: #05 of September 2023

Store at: -60°C or below (64ALDPEG); -60°C or below (64ALDPEH) For research use only. Not for use in diagnostic procedures.

#### **ASSAY PRINCIPLE**

This kit is intended for the simple and rapid quantification of Aldosterone and offers a fast alternative to ELISA.

The detection principle of this kit is based on HTRF® technology (Homogeneous Time-Resolved Fluorescence). As shown in Figure 1, Aldosterone is detected in a competitive assay by using anti Aldosterone antibody labeled with Europium cryptate (donor), and Aldosterone labeled with XL665 (acceptor).

When the dyes are in close proximity, the excitation of the donor with a light source (laser or flash lamp) triggers a Fluorescence Resonance Energy Transfer (FRET) towards the acceptor, which in turn fluoresces at a specific wavelength (665 nm). The Aldosterone present in the sample competes with the binding between the two HTRF detection solutions and thereby prevents FRET from occurring. The specific signal is inversely proportional to the Aldosterone concentration.

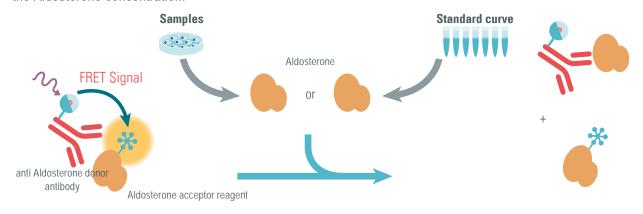
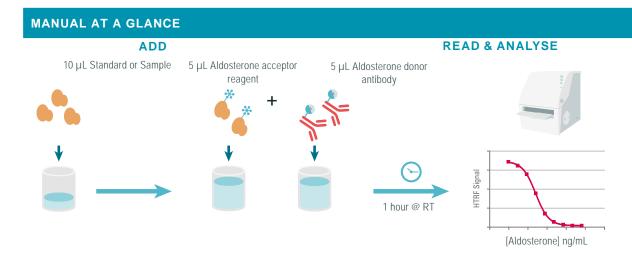


Figure 1: Principle of HTRF Aldosterone competitive assay.



Do not pre-mix the XL665 and Cryptate solutions prior to dispensing. Make sure to use the set-up for Eu Cryptate.

#### **MATERIALS PROVIDED:**

| KIT COMPONENTS                                 | 500 TESTS *<br>CAT # 64ALDPEG      | 10,000 TESTS *<br>CAT # 64ALDPEH   |
|--|------------------------------------|------------------------------------|
| Aldosterone Standard<br>Frozen                 | 1 vial - 50 μL<br>100 μg/mL - DMSO | 1 vial - 50 μL<br>100 μg/mL - DMSO |
| anti Aldosterone antibody Eu Cryptate antibody | 1 vial - 50 μL<br>Frozen - 50X     | 1 vial - 1 mL<br>Frozen - 50X      |
| Aldosterone XL665 reagent                      | 1 vial - 50 μL<br>Frozen - 50X     | 1 vial - 1 mL<br>Frozen - 50X      |
| Diluent ** ready-to-use                        | 1 vial<br>20 mL                    | 1 vial<br>20 mL                    |
| Detection buffer *** ready to use              | 1 vial<br>7 mL                     | 1 vial<br>105 mL                   |

<sup>\*</sup> When used as advised, the two available kit sizes will provide sufficient reagents for 500 tests and 10,000 tests respectively in 20 μL final volume.. Assay volumes can be adjusted proportionally to run the assay in 96 or 1536 well microplates.

#### **PURCHASE SEPARATELY:**

HTRF®-Certified Reader. Make sure the setup for Eu Cryptate is used.

For a list of HTRF-compatible readers and set-up recommendations, please visit www.revvity.com

Small volume (SV) detection microplates - .

For more information about microplate recommendations, please visit our website at: www.revvity.com

## STORAGE AND STABILITY

Store the kit at -60°C or below.

Under proper storage conditions, reagents are stable until the expiry date indicated on the label. Diluent and detection buffer are shipped frozen, but can be stored at 2-8°C in your premises.

If lyophilized, reconstituted reagents, antibodies, and standard stock solutions may be frozen and thawed only once. To avoid freeze/ thaw cycles, it is recommended to dispense remaining stock solutions into disposable plastic vials for storage at -60°C or below.

# **REAGENT PREPARATION**

### **BEFORE YOU BEGIN:**

- It is very important to prepare reagents in the specified buffers. The use of an incorrect diluent may affect reagent stability and assay results.
- Before use, allow Diluent and Detection buffer to warm up at room temperature and homogenize them with a vortex.
- Thaw all reagents at room temperature, allow them to warm up.
- Antibody solutions must be prepared in individual vials and can be mixed prior to dispensing.
- Aldosterone standards (for standard curve) must be prepared in diluent or in the same medium as the samples.

TAKE CARE TO PREPARE STOCK AND WORKING SOLUTIONS ACCORDING TO THE DIRECTIONS FOR THE KIT SIZE YOU HAVE PURCHASED.

<sup>\*\*</sup> Medium like cell culture medium can be an alternative to the diluent.

<sup>\*\*\*</sup> The Detection buffer is used to prepare working solutions of acceptor and donor reagents.

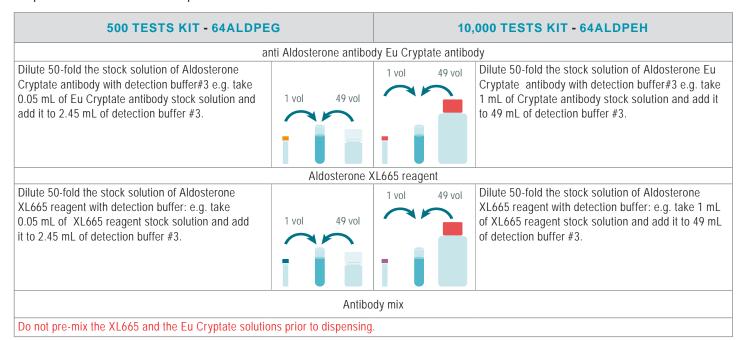
#### TO PREPARE REAGENT STOCK SOLUTIONS:

| 500 TESTS KIT - 64ALDPEG   | j                    | 10,000 TESTS           | KIT - 64ALDPEH   |  |
|--|----------------------|------------------------|--|--|
| anti   | i Aldosterone antibo | y Eu Cryptate antibody |  |  |
| Thaw the anti Aldosterone antibody Eu Cryptate antibody. Mix gently. This 50X stock solution can be frozen and stored at -60°C or below. | i                    | antibody. Mix g        | solution can be frozen and stored at                                       |  |
|  | Aldosterone 2        | <br>_665 reagent       |  |  |
| Thaw the Aldosterone XL665 reagent. Mix gently. This 50X stock solution can be frozen and stored at -60°C or below.                      | ī                    |                        | terone XL665 reagent. Mix gently.<br>solution can be frozen and stored at  |  |
|  | Aldosteror           | Standard               |  |  |
| Thaw the Aldosterone standard solution in order to obtain a 100 µg/mL (see vial label) stock solution.  Mix gently.                      |                      |                        | terone standard solution in order to y/mL (see vial label) stock solution. |  |
| Diluent  |                      |                        |  |  |
| The diluent is ready-to-use  |                      | The diluent is re      | eady-to-use  |  |
|  | Detection            | buffer                 |  |  |
| The Detection buffer is ready-to-use.  |                      | The Detection to       | ouffer is ready-to-use.  |  |

## TO PREPARE ANTIBODY WORKING SOLUTIONS:

Each well requires 5 µL anti Aldosterone antibody Eu Cryptate antibody and 5 µL Aldosterone XL665 reagent.

Prepare the two solutions in separate vials.



# TO PREPARE STANDARD WORKING SOLUTIONS:

- Each well requires 10 μL of standard.
- Dilute the standard stock solution serially with diluent or in the medium used for the preparation of the samples.
- In order to check for a potential interference effect from your own assay buffer when using the assay for the first time, we highly recommend the parallel preparation of a standard curve in your own supplemented cell culture medium and in diluent.
- · In order to counteract any standard sticking, we recommend changing tips between each dilution.

#### A recommended standard dilution procedure is listed and illustrated below:

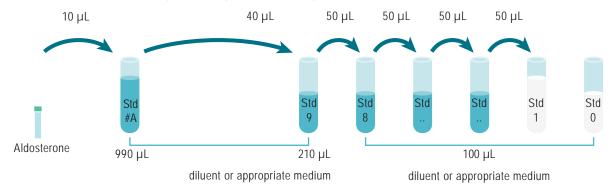
Dilute the standard stock solution 100-fold with diluent; this yields the Intermediate Standard Solution # A (1000 ng/mL). e.g. take  $10 \mu L$  of standard stock solution and add it to 990  $\mu L$  of diluent. Mix gently.

Dilute the intermediate Standard Solution #A 6.25-fold with diluent to prepare high standard (Std 9): e.g. take 40  $\mu$ L of intermediate Standard Solution #A and add it to 210  $\mu$ L of diluent. Mix gently.

Use the high standard (Std 9) to prepare the standard curve using 1/3 serial dilutions as follows:

- Dispense 100 µL of diluent in each vial from Std 8 to Std 0.
- Add 50  $\mu$ L of standard to 100  $\mu$ L of diluent, mix gently and repeat the 1/3 serial dilution to make standard solutions: std8, std7, std6, std5, std4, std3, std2, std1.

This will create 9 standards for the analyte. Std 0 (Positive control) is diluent or appropriate culture medium alone.

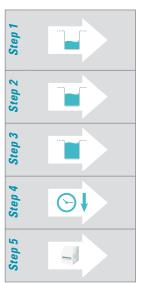


| STANDARD                          | SERIAL DILUTIONS  | ALDOSTERONE WORKING<br>SOLUTION (ng/mL) |
|-----------------------------------|---|---|
| Standard Stock solution           | Thawed stock solution                                   | 100,000                                 |
| Intermediate standard solution #A | 10 μL Standard stock solution + 990 μL Diluent          | 1,000                                   |
| Standard 9                        | 40μL Intermediate Standard Solution #A + 210 μL Diluent | 160                                     |
| Standard 8                        | 50 μL standard 9 + 100 μL Diluent                       | 53.3                                    |
| Standard 7                        | 50 μL standard 8 + 100 μL Diluent                       | 17.7                                    |
| Standard 6                        | 50 μL standard 7 + 100 μL Diluent                       | 5.9                                     |
| Standard 5                        | 50 μL standard 6 + 100 μL Diluent                       | 2                                       |
| Standard 4                        | 50 μL standard 5 + 100 μL Diluent                       | 0.65                                    |
| Standard 3                        | 50 μL standard 4 + 100 μL Diluent                       | 0.22                                    |
| Standard 2                        | 50 μL standard 3 + 100 μL Diluent                       | 0.07                                    |
| Standard 1                        | 50 μL standard 2 + 100 μL Diluent                       | 0.02                                    |
| Standard 0                        | 100 µL Diluent  | 0                                       |

## TO PREPARE SAMPLES:

- Each well requires 10 μL of sample.
- Just after their collection, put the samples at 4°C and test them immediately. For later use, samples should be dispensed into disposable plastic vials and stored at -60°C or below. Avoid multiple freeze/thaw cycles.
- Samples with a concentration above the highest standard (Std 9) must be diluted diluent or in your appropriate sample medium.

# **ASSAY MANUAL**



| Negative control or Cryptate control                              | Standard (Std 0 - Std 9)   | Samples |
|---|--|---------|
| Dispense 10 µL of diluent into each negative control well         | Dispense 10 µL of each Aldosterone standard (Std 0 - Std 9) into each sample well  Add 5 µL Aldosterone acceptor reagent working solution to all wells |         |
| Add 5 µL of detection buffer to all negative control wells        |  |         |
| Add 5 µL Aldosterone donor antibody working solution to all wells |  |         |
| Seal the plate and incubate 1 hour @ RT                           |  |         |
| Remove the plate sealer and read on an HTRF® compatible reader    |  |         |

|    | 1  | 2              | 3              | 4  | 5                            | 6                          |  |
|----|--|----------------|----------------|--|------------------------------|----------------------------|--|
|    | 10 µL diluent (Negative control)                                     |                |                | 10 μL Sample 1   |                              |                            |  |
| Δ. | 5 μL Detection Buffer# 3<br>5 μL Aldosterone donor antibody          | Repeat Well A1 | Repeat Well A1 | 5 μL Aldosterone acceptor reagent<br>5 μL Aldosterone donor antibody | Repeat Well A4               | Repeat Well A4             |  |
|    | 10 μL Std 0 (Positive control)                                       |                |                | 10 μL Sample 2   |                              |                            |  |
| В  | 5 μL Aldosterone acceptor reagent<br>5 μL Aldosterone donor antibody | Repeat Well B1 | Repeat Well B1 | 5 μL Aldosterone acceptor reagent<br>5 μL Aldosterone donor antibody | Repeat Well B4               | Repeat Well B4             |  |
|    | 10 μL Std 1  |                |                | 10 μL Sample 3   |                              |                            |  |
| С  | 5 μL Aldosterone acceptor reagent<br>5 μL Aldosterone donor antibody | Repeat Well C1 | Repeat Well C1 | 5 μL Aldosterone acceptor reagent<br>5 μL Aldosterone donor antibody | Repeat Well C4               | Repeat Well C4             |  |
|    | 10 µL Std 2  |                |                | 10 μL Sample   |                              |                            |  |
| D  | 5 μL Aldosterone acceptor reagent<br>5 μL Aldosterone donor antibody | Repeat Well D1 | Repeat Well D1 | 5 μL Aldosterone acceptor reagent<br>5 μL Aldosterone donor antibody | Repeat Well D4               | Repeat Well D4             |  |
|    | 10 μLStd   |                |                | 10 μL Sample   |                              |                            |  |
| E  | 5 μL Aldosterone acceptor reagent<br>5 μL Aldosterone donor antibody | Repeat Well E1 | Repeat Well E1 | 5 μL Aldosterone acceptor reagent<br>5 μL Aldosterone donor antibody | Repeat Well E4               | Repeat Well E4             |  |
|    | 10 μL Std  |                |                | 10 μL Sample   |                              |                            |  |
| F  | 5 μL Aldosterone acceptor reagent<br>5 μL Aldosterone donor antibody | Repeat Well F1 | Repeat Well F1 | 5 μL Aldosterone acceptor reagent<br>5 μL Aldosterone donor antibody | Repeat Well F4               | Repeat Well F4             |  |
|    | 10 μL Std  |                |                | 10 μL Sample   |                              |                            |  |
| 3  | 5 μL Aldosterone acceptor reagent<br>5 μL Aldosterone donor antibody | Repeat Well G1 | Repeat Well G1 | 5 μL Aldosterone acceptor reagent<br>5 μL Aldostero                  | Repeat Well G4 Repeat Well G |                            |  |
|    | 10 μL Std  |                |                | 10 µL Sample   |                              |                            |  |
| 4  | 5 μL Aldosterone acceptor reagent<br>5 μL Aldosterone donor antibody | Repeat Well H1 | Repeat Well H1 | 1 2 3 4 V 6 7 8 9 10 11 5 $\mu$ 8 5 $\mu$ 8 9 10 11                  | 12 13 14 15 16 1             | 7   18   19   20   21   22 |  |

# DATA REDUCTION

1. Calculate the ratio of the acceptor and donor emission signals for each individual well.

Ratio = 
$$\frac{\text{Signal 665 nm}}{\text{Signal 620 nm}} \times 10^4$$

2. Calculate the % CVs. The mean and standard deviation can then be worked out from ratio replicates.

3. Calculate the % delta F which reflects the signal to background of the assay. The negative control plays the role of an internal assay control. Delta F is used for the comparison of day to day runs of the same assay.

For more information about data reduction, please visit www.revvity.com

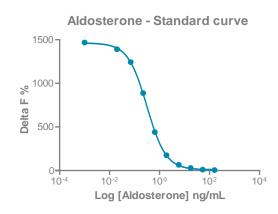
## **RESULTS**

This data must not be substituted for the data obtained in the laboratory and should be considered only as an example.

Results may vary from one HTRF® compatible reader to another.

The assay standard curve is created by plotting delta F% versus the analyte concentration.

|                          | Ratio (1) | CV (2) | Delta F% (3) |
|--------------------------|-----------|--------|--------------|
| Negative control         | 459       | 3.8%   |              |
| Std 0 – Positive control | 7,190     | 2.2%   | 1,468%       |
| Std 1 - 0.02 ng/mL       | 6,835     | 1.4%   | 1,390%       |
| Std 2 - 0.07 ng/mL       | 6,159     | 1.9%   | 1,243%       |
| Std 3 - 0.22 ng/mL       | 4,532     | 2.3%   | 888%         |
| Std 4 - 0.65 ng/mL       | 2,480     | 2.8%   | 441%         |
| Std 5 - 1.9 ng/mL        | 1,260     | 3.3%   | 175%         |
| Std 6 - 5.9 ng/mL        | 758       | 3.8%   | 65%          |
| Std 7 - 17.7 ng/mL       | 591       | 4.2%   | 29%          |
| Std 8 - 53.3 ng/mL       | 503       | 4.7%   | 10%          |
| Std 9 - 160 ng/mL        | 479       | 5.2%   | 5%           |



# ANALYTICAL CHARACTERISTICS

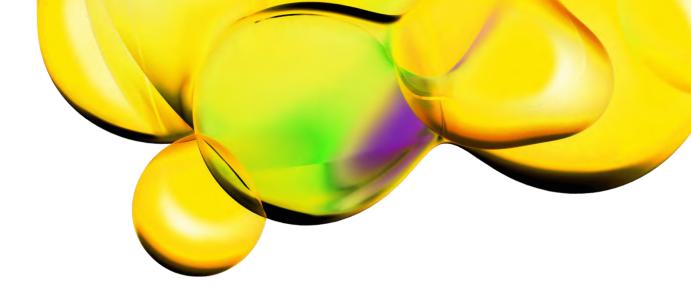
# **CROSS-REACTIVITY**

|  | Cross-reactivity (%) |                                   | Cross-reactivity (%) |                | Cross-reactivity (%) |
|--|----------------------|-----------------------------------|----------------------|----------------|----------------------|
| Aldosterone                                | 100.00%              | Cortisol                          | 0.00%                | Prednisolone   | 0.00%                |
| 3 beta, 5 alpha tetrahy-<br>droaldosterone | 7.60%                | 11-deoxycortisol                  | 0.07%                | Prednisone     | 0.00%                |
| 3 alpha, 5 beta tetrahy-<br>droaldosterone | 0.09%                | Cortisone                         | 0.01%                | Pregnenolone   | 0.00%                |
| Androsterone                               | 0.00%                | Dexamethasone                     | 0.00%                | Pregnanetriol  | 0.00%                |
| DHEA                                       | 0.00%                | Estradiol                         | 0.00%                | Testosterone   | 0.01%                |
| Androstendione                             | 0.00%                | Estriol                           | 0.00%                | Canrrenone     | 0.00%                |
| Cortexolone                                | 0.01%                | Estrone                           | 0.00%                | Prazosin-HCI   | 0.00%                |
| Corticosterone                             | 0.05%                | 9 alphafludrocortisone            | 0.00%                | Spironolactone | 0.00%                |
| 18-hydroxydeoxycorti-<br>costerone         | 0.01%                | Progesterone                      | 0.00%                |                |                      |
| 11-deoxycorticosterone                     | 0.00%                | 17 alpha-hydroxyproges-<br>terone | 0.01%                |                |                      |

# **DETECTION LIMIT & EC50 WORKING CONCENTRATIONS**

| Detection limit | 25 pg/mL  |
|-----------------|-----------|
| EC50            | 300 pg/mL |

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