



## MOUSE TNF $\alpha$ KITS

Part # 62MTNFAPEG & 62MTNFAPEH

Test Size#: 500 tests (62MTNFAPEG), 10,000 tests (62MTNFAPEH)

Assay volume: 20  $\mu$ L

Revision: #10 of April 2024 Store at:  $\leq -16^{\circ}\text{C}$

This product is intended for research purposes only. It is not intended to be used for therapeutic or diagnostic purposes.

### ASSAY PRINCIPLE

Revvity's mouse TNF $\alpha$  assay is only intended for the quantitative measurement of TNF $\alpha$  in supernatant using HTRF™ technology. The assay is compatible with mouse samples, and is highly specific for TNF $\alpha$ .

TNF $\alpha$  is detected in a sandwich assay format using 2 different specific antibodies, one labeled with Europium Cryptate (donor) and the second with d2 (acceptor).

The detection principle is based on HTRF™ technology. When the labelled antibodies bind to the same antigen, the excitation of the donor with a light source (laser or flash lamp) triggers a Fluorescence Resonance Energy Transfer (FRET) towards the acceptor, which in turn fluoresces at a specific wavelength (665 nm). The two antibodies bind to the TNF $\alpha$  present in the sample, thereby generating FRET. Signal intensity is proportional to the number of antigen-antibody complexes formed and therefore to the TNF $\alpha$  concentration. (Fig. 1).

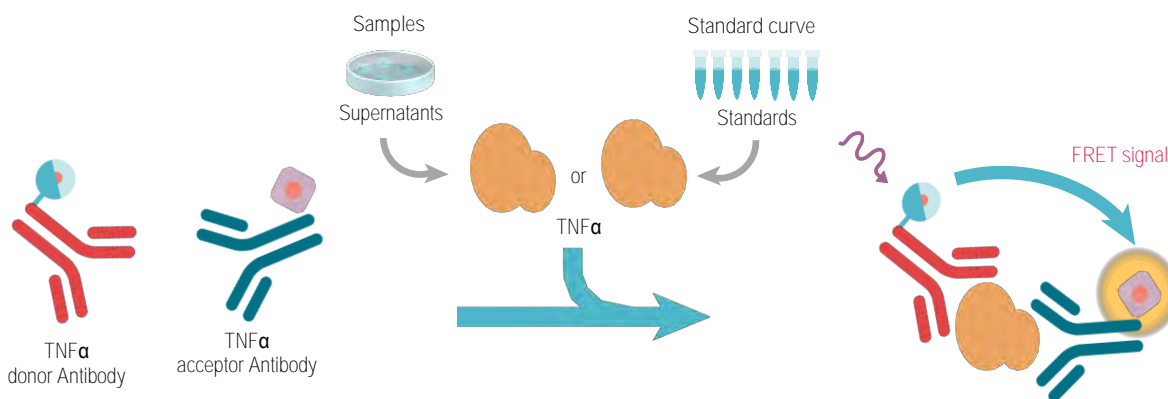
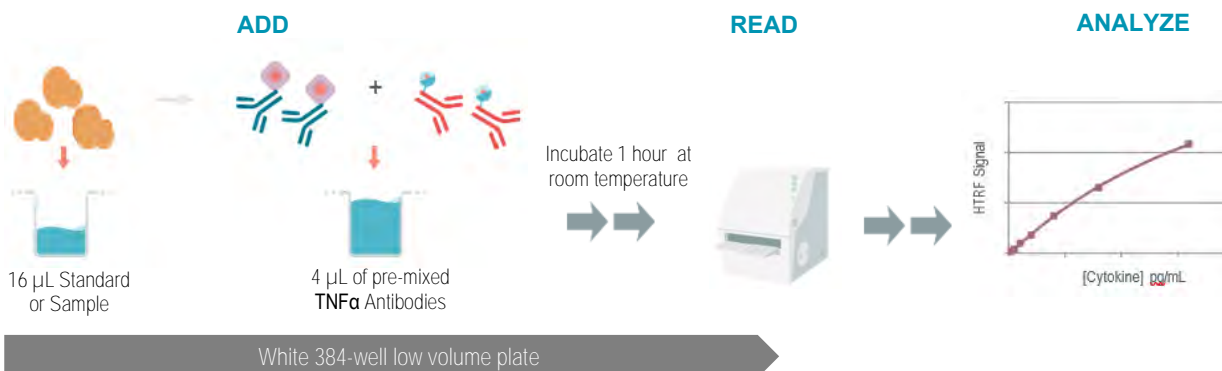


Figure 1. Principle of HTRF TNF $\alpha$  sandwich assay

### MANUAL AT A GLANCE



Make sure to use the set-up for Eu<sup>3+</sup> Cryptate.

For more information about set-up and compatible HTRF™ readers, please visit our website at: [www.revvity.com](http://www.revvity.com)

## MATERIALS

KIT COMPONENTS	500 TESTS CAT#62HIL12P40PEG	10,000 TESTS CAT#62HIL12P40PEH
TNF $\alpha$ Standard Lyophilized	2 vials	2 vials
TNF $\alpha$ Eu Cryptate Antibody Frozen 20X	1 vial 50 $\mu$ L	1 vial 1 mL
TNF $\alpha$ d2 Antibody Frozen 20X	1 vial 50 $\mu$ L	1 vial 1 mL
Diluent* #5 5X	1 vial 2 mL	1 vial 10 mL
Detection Buffer** #3 Ready-to-use	2 vials 1.5 mL	1 vial 50 mL

\* To prepare working standard solutions, culture medium can be an alternative the diluent.

\*\* The Detection Buffer is used to prepare working solutions of acceptor and donor reagents.

### For reading, an HTRF™ - certified reader is needed

For a list of HTRF-compatible readers and set-up recommendations, please visit [www.revvy.com](http://www.revvy.com)

### Purchase separately

96-well or 384-well small volume (SV) detection microplates - For more information about microplate recommendations, please visit our website at: [www.revvy.com](http://www.revvy.com)

## STORAGE AND STABILITY

### KIT:

- Store the kit at  $\leq -16^{\circ}\text{C}$ . Under appropriate storage conditions, reagents are stable until the expiry date indicated on the label

### REAGENTS:

- Once reconstituted, standard stock solution may be frozen, and can be thawed only once.
- Once thawed, antibody solutions can be frozen once.
- To avoid freeze/thaw cycles, it is recommended to dispense remaining stock solutions into disposable plastic vials for storage at  $\leq -60^{\circ}\text{C}$ .
- Volume of standard and antibody aliquots should not be under 10  $\mu$ L.
- Thawed diluent and detection buffer can be stored at 2-8 $^{\circ}\text{C}$  on your premises.




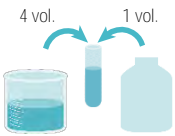
## REAGENT PREPARATION

### BEFORE YOU BEGIN

- It is very important to prepare reagents in the specified buffers. The use of an incorrect diluent may affect reagent stability and assay results.
- Thaw the frozen reagents at room temperature.
- Before use, allow all kit's reagents to warm up at room temperature then
  - homogenise buffer and diluent with a vortex
  - centrifuge (NEVER vortex) the antibodies to gather all liquid at the bottom of the vial
- It is recommended to filter buffers before use.
- Antibody solutions must be prepared in individual vials and can be mixed prior to dispensing.

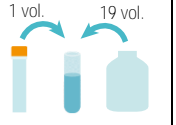
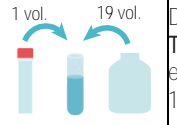
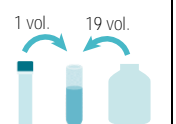


**TAKE CARE TO PREPARE STOCK AND WORKING SOLUTIONS ACCORDING TO THE DIRECTIONS FOR THE KIT SIZE YOU HAVE PURCHASED.**

### TO PREPARE DILUENT, STANDARD & ANTIBODY STOCK SOLUTIONS:

500 TESTS		10,000 TESTS	
<b>TNF<math>\alpha</math> Eu Cryptate antibody</b>			
Thaw the <b>TNF<math>\alpha</math></b> Eu Cryptate antibody. Centrifuge. This 20X stock solution can be frozen and stored at $\leq -60^{\circ}\text{C}$ .			Thaw the <b>TNF<math>\alpha</math></b> Eu Cryptate antibody. Centrifuge. This 20X stock solution can be frozen and stored at $\leq -60^{\circ}\text{C}$ .
<b>TNF<math>\alpha</math> d2 antibody</b>			
Thaw the <b>TNF<math>\alpha</math></b> d2 antibody. Centrifuge. This 20X stock solution can be frozen and stored at $\leq -60^{\circ}\text{C}$ .			Thaw the <b>TNF<math>\alpha</math></b> d2 antibody. Centrifuge. This 20X stock solution can be frozen and stored at $\leq -60^{\circ}\text{C}$ .
<b>TNF<math>\alpha</math> Standard</b>			
Reconstitute the <b>TNF<math>\alpha</math></b> standard with distilled water. Volume of reconstitution is indicated on the vial label. The reconstituted standard solution can be frozen and stored at $-60^{\circ}\text{C}$ or below.			Reconstitute the <b>TNF<math>\alpha</math></b> standard with distilled water. Volume of reconstitution is indicated on the vial label. The reconstituted standard solution can be frozen and stored at $-60^{\circ}\text{C}$ or below.
<b>Diluent</b>			
Dilute 5-fold the 5X diluent #5 with distilled water: Homogenize the 5X diluent #5 with a vortex and add 1 volume of stock solution in 4 volumes of distilled water e.g. 1 mL of diluent + 4 mL of distilled water Mix gently after dilution.			Dilute 5-fold the 5X diluent #5 with distilled water: Homogenize the 5X diluent #5 with a vortex and add 1 volume of stock solution in 4 volumes of distilled water e.g. 10 mL of diluent + 40 mL of distilled water Mix gently after dilution.

### TO PREPARE WORKING ANTIBODY SOLUTIONS:

Each well requires 4  $\mu\text{L}$  of pre-mixed TNF $\alpha$  antibodies. Prepare the two antibody solutions in separate vials

500 TESTS		10,000 TESTS	
<b>TNF<math>\alpha</math> Eu Cryptate antibody</b>			
Dilute 20-fold the 20 X stock solution (thawed reagent) of <b>TNF<math>\alpha</math></b> Eu Cryptate antibody with detection buffer #3: e.g. 10 $\mu\text{L}$ of thawed Eu Cryptate antibody stock solution + 190 $\mu\text{L}$ of detection buffer.			Dilute 20-fold the 20 X stock solution (thawed reagent) of <b>TNF<math>\alpha</math></b> Eu Cryptate antibody with detection buffer #3: e.g. 10 $\mu\text{L}$ of thawed Eu Cryptate antibody stock solution + 190 $\mu\text{L}$ of detection buffer).
<b>TNF<math>\alpha</math> d2 antibody</b>			
Dilute 20-fold the 20 X stock solution (thawed reagent) of <b>TNF<math>\alpha</math></b> d2 antibody with detection buffer #3: e.g. 10 $\mu\text{L}$ of thawed d2 antibody stock solution + 190 $\mu\text{L}$ of detection buffer.			Dilute 20-fold the 20 X stock solution (thawed reagent) of <b>TNF<math>\alpha</math></b> d2 antibody with detection buffer #3: e.g. 10 $\mu\text{L}$ of thawed d2 antibody stock solution + 190 $\mu\text{L}$ of detection buffer.
<b>Antibody mix</b>			
Pre-mix the two ready-to-use antibody solutions just prior to dispensing the reagents: e.g. 200 $\mu\text{L}$ of d2 antibody + 200 $\mu\text{L}$ of Eu Cryptate antibody			Pre-mix the two ready-to-use antibody solutions just prior to dispensing the reagents: e.g. 200 $\mu\text{L}$ of d2 antibody + 200 $\mu\text{L}$ of Eu Cryptate- antibody

**TO PREPARE WORKING STANDARDS SOLUTIONS:**

- Each well requires 16 µL of standard.
- Serially dilute the standard stock solution with diluent #5 or with the cell culture medium used to prepare your samples, supplemented with BSA or 10% FCS.
- **Due to the stability of the TNFα, it is mandatory to prepare the standard curve just before the assay.**
- In order to check for a potential interference effect from your own assay buffer when using the assay for the first time, we highly recommend the parallel preparation of a standard curve in your own supplemented cell culture medium and in diluent.
- In order to counteract any standard sticking, we recommend changing tips between each dilution.

A recommended standard dilution procedure is listed and illustrated below:

1. Reconstitute the standard vial with the volume indicated on the vial label using distilled water.
2. Prepare the following dilutions:

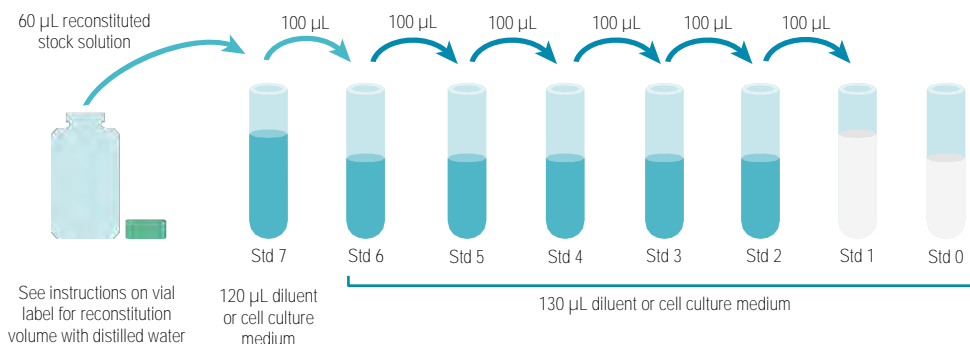
- Dilute the reconstituted standard stock solution 3-fold with diluent or with cell culture medium.

In practice: take 60 µL of stock solution and add it to 120 µL of diluent or cell culture medium. Mix gently. This yields the high standard (Std 7: 6000 pg/mL) for the top of the curve.

- Use the high standard (Std 7) to prepare the standard curve using serial dilutions as follows:

- Dispense 120 µL of diluent or cell culture medium into each vial from Std 6 to Std 0
- Add 100 µL of standard to 120 µL of diluent or cell culture medium, mix gently and repeat the serial dilution to make standard solutions: std6, std5, std4, std3, std2, std1

This will create 7 standards for the analyte. Std 0 (Negative control) is diluent or appropriate culture medium alone.



STANDARD	SERIAL DILUTIONS	WORKING SOLUTIONS
Standard Stock solution	Reconstitute the vial following the indications given on the vial label	18 ng/mL
Standard 7	60 µL reconstituted standard stock solution + 120 µL diluent	6000 pg/mL
Standard 6	100 µL Standard 7 + 130 µL diluent	2608.7 pg/mL
Standard 5	100 µL Standard 6 + 130 µL diluent	1134.2 pg/mL
Standard 4	100 µL Standard 5 + 130 µL diluent	493.1 pg/mL
Standard 3	100 µL Standard 4 + 130 µL diluent	214.4 pg/mL
Standard 2	100 µL Standard 3 + 130 µL diluent	93.2 pg/mL
Standard 1	100 µL Standard 2 + 130 µL diluent	40.5 pg/mL
Standard 0	130 µL diluent	0



## DATA REDUCTION & INTERPRETATION

1. Calculate the ratio of the acceptor and donor emission signals for each individual well.

$$\text{Ratio} = \frac{\text{Signal 665 nm}}{\text{Signal 620 nm}} \times 10^4$$

2. Calculate the delta ratio of the acceptor and donor emission signals for each individual well. The Standard 0 (Negative control) plays the role of an internal assay control.

$$\text{delta Ratio} = \text{Ratio Standard or sample} - \text{Ratio Standard 0}$$

3. Calculate the % CVs. The mean and standard deviation can then be worked out from ratio replicates.

$$\text{CV(\%)} = \frac{\text{Standard deviation}}{\text{Mean Ratio}} \times 100$$

For more information about data reduction, please visit [www.revivity.com](http://www.revivity.com)

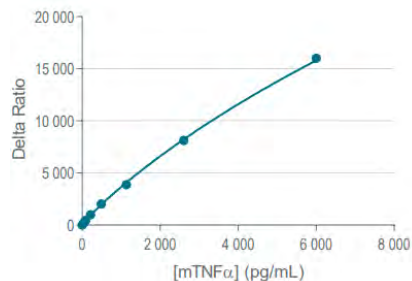
## RESULTS

This data must not be substituted for the data obtained in the laboratory, and should be considered only as an example.

Results may vary from one HTRF™ compatible reader to another.

Standard curve fitting with the 4 Parameter Logistic (4PL 1/y2) model (For more information about curve fitting please visit [www.revivity.com](http://www.revivity.com))

		Ratio (1)	delta R (2)	CV% (3)
Standard 0	Negative control	552	0	0%
Standard 1	40.5 pg/mL	755	203	3%
Standard 2	93.2 pg/mL	989	437	2%
Standard 3	214.4 pg/mL	1539	987	2%
Standard 4	493.1 pg/mL	2572	2020	0%
Standard 5	1134.2 pg/mL	4425	3873	4%
Standard 6	2608.7 pg/mL	8670	8118	1%
Standard 7	6000 pg/mL	16554	16002	0%

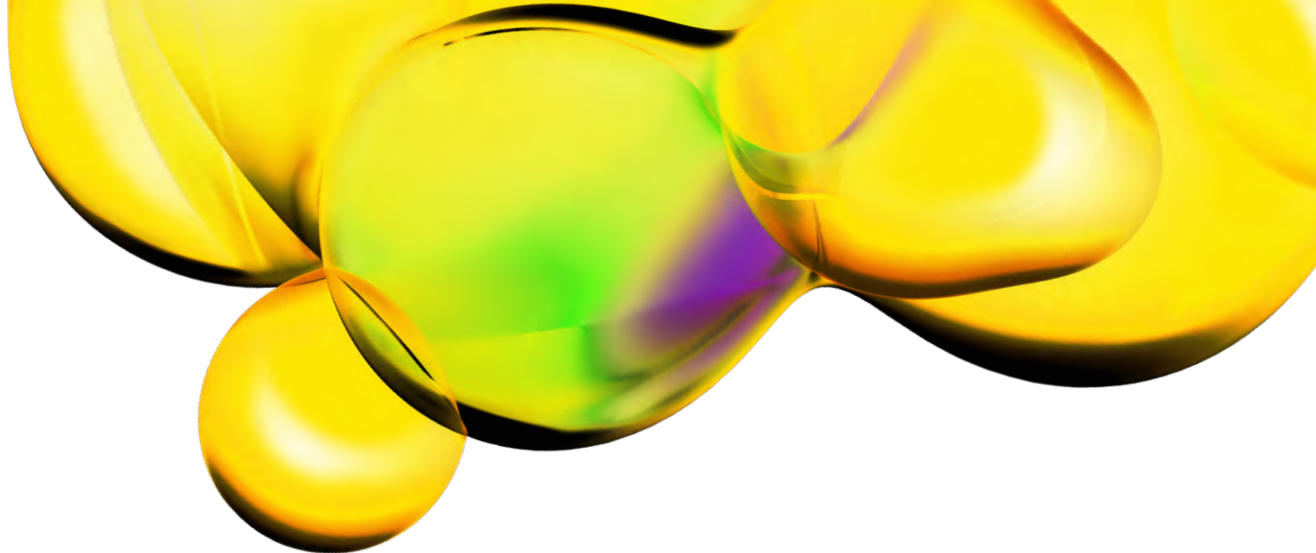


## ANALYTICAL ASSAY PERFORMANCE

	Diluent	DMEM	RPMI
Assay range (pg/mL**)	20 pg/mL to 6000 pg/mL		
Limit of detection (LoD*) = Std 0 mean + 2 SD	3 pg/mL	11 pg/mL	8 pg/mL
Limit of quantification (LoQ*)	20 pg/mL		
Incubation time	1 hour at room temperature		

\* The analytical sensitivity was calculated from data obtained with an HTRF compatible reader after 1 hour incubation, this may vary from one HTRF compatible reader to another.

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