

MANUAL

Technology: HTRF[™] Cytokine

HTRF High Performance Human IL-6 Kit

Part number:	62HIL6V2PEG	62HIL6V2PEH
Test size	500 tests	10,000 tests

Storage: ≤ 20°C

Version: 01 Date: July 2024

ASSAY PRINCIPLE

This kit is intended for the simple and rapid quantification of human IL-6 in supernatant and offers a fast alternative to ELISA.

The detection principle of this kit is based on $\mathsf{HTRF}^\mathsf{TM}$ technology (Homogeneous Time-Resolved Fluorescence). As shown in Figure 1, IL-6 is detected in a sandwich assay by using anti-IL-6 antibody labeled with Europium cryptate (donor), and anti-IL-6 antibody labeled with d2 (acceptor).

When the dyes are in close proximity, the excitation of the donor with a light source (laser or flash lamp) triggers a Fluorescence Resonance Energy Transfer (FRET) towards the acceptor, which in turn fluoresces at a specific wavelength (665 nm). Signal intensity is proportional to the number of antigen-antibody complexes formed and therefore to the IL-6 concentration.

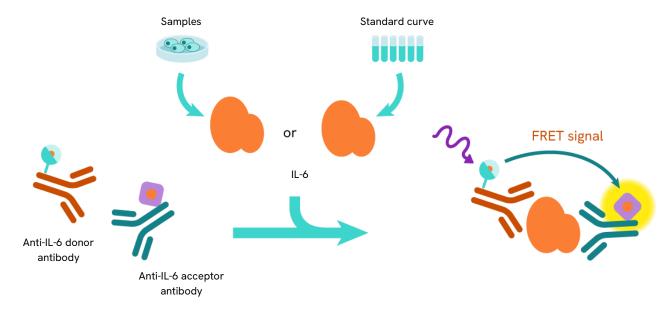
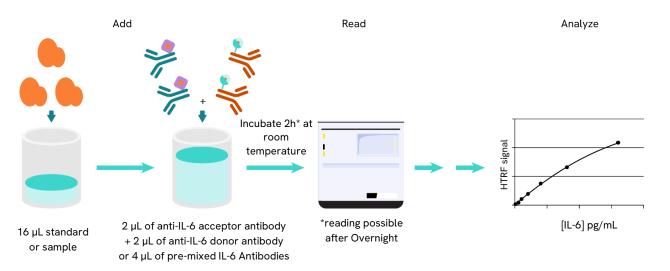


Figure 1: Principle of HTRF IL-6 sandwich assay

PROTOCOL AT A GLANCE



Make sure to use the set-up for Eu Cryptate.

MATERIAL PROVIDED

KIT COMPONENTS		500 TEST	гs*	10,000 TESTS*						
IL-6 Standard Lyophilized		green cap	2 vials	Ī	green cap	2 vials				
IL-6 Eu Cryptate Antibody Frozen 20X	Ī	orange cap	1 vial 50 µL	Ī	red cap	1 vial 1 mL				
IL-6 d2 Antibody Frozen 20X		blue cap	1 vial 50 µL		purple cap	1 vial 1 mL				
Diluent** #5 5X	i	white cap	1 vial 2 mL		white cap	1 vial 10 mL				
Detection Buffer*** #3 Ready-to-use		red cap	2 vials 1.5 mL		red cap	1 vial 50 mL				

^{*} When used as advised, the two available kit sizes will provide sufficient reagents for 500 tests and 10,000 tests respectively in 20 µL final volume. Assay volumes can be adjusted proportionally to run the assay in 96 or 1536 well microplates.

Purchase separately

- HTRF™-Certified Reader. Make sure the setup for Eu Cryptate is used. For a list of HTRF-compatible readers and set-up recommendations, please visit our website.
- Small volume (SV) detection microplates. Use white plate only. For more information about microplate recommendations, please visit our website.

STORAGE AND STABILITY

Kit

- Store the kit at -20°C or below.
- Under proper storage conditions, reagents are stable until the expiry date indicated on the label.
- Diluent and detection buffer are shipped frozen but can be stored at 2-8°C in your premises.

Reagents

Once reconstituted, standard stock solution may be frozen at \leq -60°C, and can be thawed only once. Once thawed, antibody solutions can be frozen once.

To avoid freeze/thaw cycles, it is recommended to dispense remaining stock solutions into disposable plastic vials for storage at \leq -60°C for standard and at \leq -20°C for antibodies.

Volume of standard and antibody aliquots should not be under 10 µL.

Thawed diluent and detection buffer can be stored at 2-8°C on your premises.

REAGENT PREPARATION

Before you begin

- It is very important to prepare reagents in the specified buffers. The use of an incorrect diluent may affect reagent stability and assay results.
- Thaw the frozen reagents at room temperature, allow them to warm up to room temperature for at least 30 mins before use.
- Before use, allow Diluent and Detection buffer to warm up at room temperature and homogenize them with a vortex.
- Antibody solutions must be prepared in individual vials and can be mixed prior to dispensing.
- IL-6 standards (for standard curve) must be prepared in diluent or in the same medium as the samples.

^{**} Medium like cell culture medium can be an alternative to the diluent.

^{***} The Detection buffer is used to prepare working solutions of acceptor and donor reagents.

Take care to prepare stock and working solutions according to the directions for the kit size you have purchased.

To prepare reagent stock solutions

10,000 TESTS **500 TESTS** Anti-IL-6 Eu Cryptate antibody Thaw the IL-6 Eu Cryptate antibody. Mix Thaw the IL-6 Eu Cryptate antibody. Mix gently. This 20X stock solution can be frozen and gently. This 20X stock solution can be frozen and stored at -20°C or below. To avoid stored at -20°C or below. To avoid freeze/thaw cycles, it is recommended to freeze/thaw cycles, it is recommended to dispense remaining stock solutions into dispense remaining stock solutions into disposable plastic vials for storage at -20°C or disposable plastic vials for storage at -20°C or below. below. Anti-IL-6 d2 antibody Thaw the IL-6 d2 antibody. Mix gently. This Thaw the IL-6 d2 antibody. Mix gently. This 20X 20X stock solution can be frozen and stored at stock solution can be frozen and stored at -20°C or below. To avoid freeze/thaw cycles, it -20°C or below. To avoid freeze/thaw cycles, is recommended to dispense remaining stock it is recommended to dispense remaining stock solutions into disposable plastic vials for solutions into disposable plastic vials for storage at -20°C or below. storage at -20°C or below. **IL-6 Standard** Reconstitute the IL-6 standard with distilled Reconstitute the IL-6 standard with distilled water. Volume of reconstitution is indicated on water. Volume of reconstitution is indicated on the vial label. The reconstituted standard the vial label. The reconstituted standard solution can be frozen and stored at -60°C or solution can be frozen and stored at -60°C or below below Diluent Dilute 5-fold the 5 X diluent #5 with distilled Dilute 5-fold the 5 X diluent #5 with distilled water: homogenize the 5 X diluent #5 with a water: homogenize the 5 X diluent #5 with a vortex and add 1 volume of stock solution in 4 vortex and add 1 volume of stock solution in 4 volumes of distilled water (e.g., 10 mL of volumes of distilled water (e.g., 1 mL of diluent + 4 mL of distilled water). Mix gently after diluent + 40 mL of distilled water). Mix gently dilution. This 1X diluent can be frozen and after dilution. This 1X diluent can be frozen stored at -20°C or below. and stored at -20°C or below. **Detection buffer**

The Detection buffer is ready-to-use.

To prepare antibody working solutions

Each well requires 2 μ L of IL-6-Eu Cryptate Antibody and 2 μ L of IL-6 d2 Antibody. Prepare the two antibody solutions in separate vials.

500 TESTS 10,000 TESTS IL-6 Eu Cryptate antibody Dilute 20-fold the 20X stock Dilute 20-fold the 20X stock solution solution (thawed reagent) of IL-6 Eu (thawed reagent) of IL-6 Eu Cryptate Cryptate antibody with Detection antibody with Detection buffer #3: 19 vol. 1 vol. 19 vol. buffer #3: add 1 volume of Eu add 1 volume of Eu Cryptate Cryptate antibody stock solution in antibody stock solution in 19 19 volumes of detection buffer (e.g. volumes of detection buffer (e.g.10 10 μL of Eu Cryptate antibody μL of Eu Cryptate antibody stock stock solution + 190 μ L of detection solution + 190 μ L of detection buffer). buffer). IL-6 d2 antibody Dilute 20-fold the 20X stock Dilute 20-fold the 20X stock solution solution (thawed reagent) of IL-6 d2 (thawed reagent) of IL-6 d2 antibody 19 vol. 1 vol. 19 vol. 1 vol. antibody with Detection buffer #3: with Detection buffer #3: add 1 add 1 volume of d2 antibody stock volume of d2 antibody stock solution in 19 volumes of detection solution in 19 volumes of detection buffer (e.g. 10 µL of Eu Cryptate buffer (e.g. 10 µL of Eu Cryptate antibody stock solution + 190 µL of antibody stock solution + 190 µL of detection buffer). detection buffer). **Antibody Mix** It is possible to pre-mix the two It is possible to pre-mix the two ready-to-use antibody solutions just ready-to-use antibody solutions just prior to dispensing the reagents by prior to dispensing the reagents by adding 1 volume of d2 antibody adding 1 volume of d2 antibody solution to 1 volume of Cryptate solution to 1 volume of Cryptate antibody solution (e.g. 1 mL of d2 antibody solution (e.g. 20 mL of d2 antibody + 1 mL of Cryptate antibody + 20 mL of Cryptate antibody). antibody).

To prepare working standards solutions

- Each well requires 16 µL of standard.
- Dilute the standard stock solution serially with diluent #5 (1X)
- If culture medium is used to dilute the standard, we recommend to supplement it with serum (2 to 10%) or BSA (0.2 to 1%) in order to avoid IL-6 sticking to assay plates.
- In order to check for a potential interference effect from your own assay buffer when using the assay for the first time, we highly recommend the parallel preparation of a standard curve in your own supplemented cell culture medium and in diluent #5 (1X).
- In order to counteract any standard sticking, we recommend changing tips between each dilution.

A recommended standard dilution procedure is listed and illustrated below:

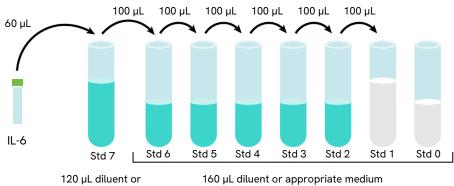
Dilute the standard stock solution 3-fold with diluent; this yields the Standard Max solution (12 000 pg/mL)

Dilute the standard stock solution 3-fold with diluent #5 (1X) to prepare high standard (Std 7): e.g. take $60 \mu L$ of standard stock solution and add it to $120 \mu L$ of diluent #5 (1X). Mix gently.

Use the high standard (Std 7) to prepare the standard curve using serial dilutions as follows:

- Dispense 160 μL of diluent #5 (1X) in each vial from Std 6 to Std 0.
- Add 100 μ L of standard to 160 μ L of diluent #5 (1X), mix gently and repeat the serial dilution to make standard solutions: std6, std5, std4, std3, std2, std1.

This will create 7 standards for the analyte. Std 0 (Negative control) is diluent #5 (1X) or appropriate culture medium alone



120 µL diluent or appropriate medium

STANDARD	NDARD SERIAL DILUTIONS						
Standard Stock solution	Reconstituted lyophilizate	36 ng/mL					
Standard 7	60 μL Standard stock Solution + 120 μL diluent 1X	12 000 pg/mL					
Standard 6	Standard 6 100 µL standard 7 + 160 µL diluent 1X						
Standard 5	100 μL standard 6 + 160 μL diluent 1X	1775.1 pg/mL					
Standard 4	100 μL standard 5 + 160 μL diluent 1X	682.7 pg/mL					
Standard 3	100 μL standard 4 + 160 μL diluent 1X	262.6 pg/mL					
Standard 2	100 μL standard 3 + 160 μL diluent 1X	101 pg/mL					
Standard 1	100 μL standard 2 + 160 μL diluent 1X	38.8 pg/mL					
Standard 0	160 μL diluent 1X	0					

To prepare samples

- Each well requires 16 µL of sample.
- Just after their collection, put the samples at 4°C and test them immediately. For later use, samples should be dispensed into disposable plastic vials and stored at -60°C or below. Avoid multiple freeze/thaw cycles.
- Samples with a concentration above the highest standard (Std 7) must be diluted diluent in your appropriate sample medium.
- To obtain additional information or support, please contact the HTRF technical support team.

ASSAY PROTOCOL

		STANDARD (STD 0 - STD 7)	SAMPLES							
Step 1		Dispense 16 µL of each IL-6 standard (Std 0 - Std 7) into each standard well	Dispense 16 µL of each sample into each sample well							
Step 2		Add 2 μL of IL-6 d2 antibody working solution to all wells								
Step 3		Add 2 µL of IL-6 Eu Cryptate antibo	ody working solution to all wells.							
Step 4	0	Seal the plate and incubate from 2 hours at RT *Reading possible after Overnight								
Step 5		Remove the plate sealer and read	on an HTRF™ compatible reader							

	1	2	3	4	5	6
Α	16 μL Std 0 (Negative control) 2 μL IL-6-d2 2 μL IL-6-Eu Cryptate	Repeat Well A1	Repeat Well A1	16 μL sample 1 2 μL IL-6-d2 2 μL IL-6-Eu Cryptate	Repeat Well A4	Repeat Well A4
В	16 µL Std 1 2 µL IL-6-d2 2 µL IL-6-Eu Cryptate	Repeat Well B1	Repeat Well B1	<mark>16 μL sample 2</mark> 2 μL IL-6-d2 2 μL IL-6-Eu Cryptate	Repeat Well B4	Repeat Well B4
С	16 µL Std 2 2 µL IL-6-d2 2 µL IL-6-Eu Cryptate	Repeat Well C1	Repeat Well C1	<mark>16 μL sample 3</mark> 2 μL IL-6-d2 2 μL IL-6-Eu Cryptate	Repeat Well C4	Repeat Well C4
D	16 µL Std 3 2 µL IL-6-d2 2 µL IL-6-Eu Cryptate	Repeat Well D1	Repeat Well D1	16 μL sample 2 μL IL-6-d2 2 μL IL-6-Eu Cryptate	Repeat Well D4	Repeat Well D4
Е	16 µL Std 4 2 µL IL-6-d2 2 µL IL-6-Eu Cryptate	Repeat Well E1	Repeat Well E1	<mark>16 μL sample</mark> 2 μL IL-6-d2 2 μL IL-6-Eu Cryptate	Repeat Well E4	Repeat Well E4
F	16 µL Std 5 2 µL IL-6-d2 2 µL IL-6-Eu Cryptate	Repeat Well F1	Repeat Well F1	1 <mark>6 μL sample</mark> 2 μL IL-6-d2 2 μL IL-6-Eu Cryptate	Repeat Well F4	Repeat Well F4
G	16 µL Std 6 2 µL IL-6-d2 2 µL IL-6-Eu Cryptate	Repeat Well G1	Repeat Well G1	16 μL sample 2 μL IL-6-d2 2 μL IL-6-Eu Cryptate	Repeat Well G4	Repeat Well G4
Н	16 µL Std 7 2 µL IL-6-d2 2 µL IL-6-Eu Cryptate	Repeat Well H1	Repeat Well H1	1 <mark>6 μL sample</mark> 2 μL IL-ό-d2 2 μL IL-ό-Eu Cryptate	Repeat Well H4	Repeat Well H4

	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23	24
Α																								
В																								
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DATA REDUCTION & INTERPRETATION

1) Calculate the ratio of the acceptor and donor emission signals for each individual well.

Ratio =
$$\frac{\text{Signal 665 nm}}{\text{Signal 620 nm}} \times 10^4$$

2) Calculate the % CVs. The mean and standard deviation can then be worked out from ratio replicates.

$$CV (\%) = \frac{Standard\ deviation}{Mean\ Ratio} \times 100$$

For more information about data reduction, please visit our website.

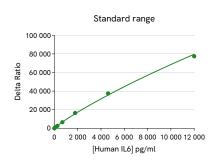
RESULTS

This data must not be substituted for the data obtained in the laboratory, and should be considered only as an example.

Results may vary from one HTRF™ compatible reader to another.

Standard curve fitting with the 4 Parameter Logistic (4PL with 1/Y2) model

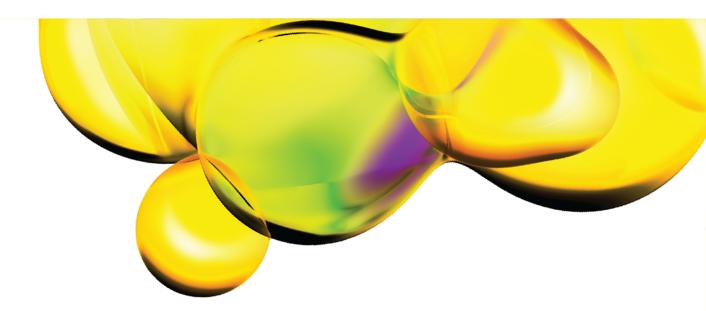
		Ratio (1)	CV% (2)
Standard 0	Negative control	1117	8%
Standard 1	38.8 pg/mL	1777	5,1%
Standard 2	101 pg/mL	2409	4,4%
Standard 3	262.6 pg/mL	3831	0,5%
Standard 4	682.7 pg/mL	7714	3,2%
Standard 5	1775.1 pg/mL	17712	2,1%
Standard 6	4615.4 pg/mL	38652	1,2%
Standard 7	12 000 pg/mL	78740	2,6%



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