



## HTRF Human IL12/23 p40 Detection kit

**Part # 62HIL12P40PEG & 62HIL12P40PEH**

**Test Size#:** 500 tests (62HIL12P40PEG), 10,000 tests (62HIL12P40PEH)

**Assay volume:** 20  $\mu$ L

**Revision:** #03 of June 2024 **Store at:**  $\leq -16^{\circ}\text{C}$

**This product is intended for research purposes only. It is not intended to be used for therapeutic or diagnostic purposes.**

### ASSAY PRINCIPLE

Revvity' human IL12/23 p40 assay is only intended for the quantitative measurement of IL12/23 p40 in supernatant using HTRF<sup>®</sup> technology. The assay is compatible with human samples only and is highly specific for p40, cross-reacting with all p40-containing cytokines such as IL12, IL23, p40 monomer and p40-p40 (p80) dimer.

IL12/23 p40 is detected in a sandwich assay format using 2 different specific antibodies, one labeled with Europium Cryptate (donor) and the second with d2 (acceptor).

The detection principle is based on HTRF<sup>®</sup> technology. When the labelled antibodies bind to the same antigen, the excitation of the donor with a light source (laser or flash lamp) triggers a Fluorescence Resonance Energy Transfer (FRET) towards the acceptor, which in turn fluoresces at a specific wavelength (665 nm). The two antibodies bind to the IL12/23 p40 present in the sample, thereby generating FRET. Signal intensity is proportional to the number of antigen-antibody complexes formed and therefore to the IL12/23 p40 concentration (Fig. 1).

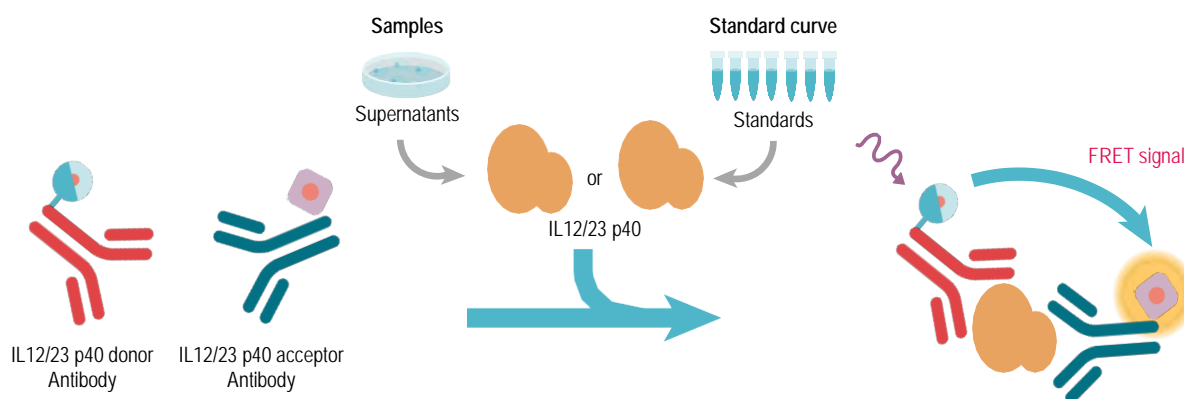
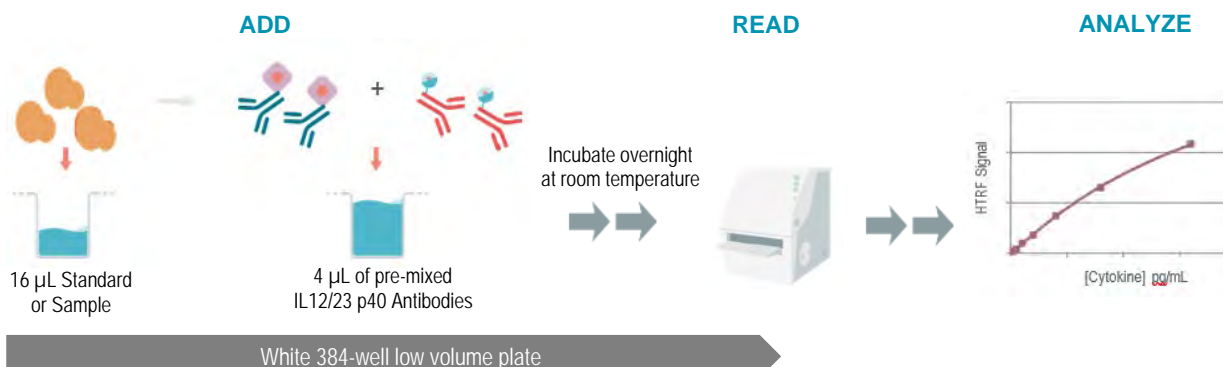


Figure 1. Principle of HTRF IL12/23 p40 sandwich assay

### MANUAL AT A GLANCE



Make sure to use the set-up for Eu<sup>3+</sup> Cryptate. For more information about set-up and compatible HTRF<sup>®</sup> readers, please visit our website at: [www.revvity.com](http://www.revvity.com)

## MATERIALS

KIT COMPONENTS	500 TESTS CAT#62HIL12P40PEG	10,000 TESTS CAT#62HIL12P40PEH
IL 12/23 p40 Standard Lyophilized	2 vials	2 vials
IL 12/23 p40 Eu Cryptate Antibody Frozen 20X	1 vial 50 $\mu$ L	1 vial 1 mL
IL 12/23 p40 d2 Antibody Frozen 20X	1 vial 50 $\mu$ L	1 vial 1 mL
Diluent* #5 5X	1 vial 2 mL	1 vial 10 mL
Detection Buffer** #3 Ready-to-use	2 vials 1.5 mL	1 vial 50 mL

\* To prepare working standard solutions, culture medium can be an alternative the diluent.

\*\* The Detection Buffer is used to prepare working solutions of acceptor and donor reagents.

### For reading, an HTRF®- certified reader is needed

For a list of HTRF-compatible readers and set-up recommendations, please visit [www.revivity.com](http://www.revivity.com)

### Purchase separately

96-well or 384-well small volume (SV) detection microplates - For more information about microplate recommendations, please visit our website at: [www.revivity.com](http://www.revivity.com)

## STORAGE AND STABILITY

### KIT:

- Store the kit at  $\leq -16^{\circ}\text{C}$ . Under appropriate storage conditions, reagents are stable until the expiry date

### REAGENTS:

- Once reconstituted, standard stock solution may be frozen, and can be thawed only once.
- Once thawed, antibody solutions can be frozen once.
- To avoid freeze/thaw cycles, it is recommended to dispense remaining stock solutions into disposable plastic vials for storage at  $\leq -60^{\circ}\text{C}$ .
- Volume of standard and antibody aliquots should not be under 10  $\mu\text{L}$ .
- Thawed diluent and detection buffer can be stored at  $2-8^{\circ}\text{C}$  on your premises.






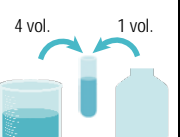
## REAGENT PREPARATION

### BEFORE YOU BEGIN

- It is very important to prepare reagents in the specified buffers. The use of an incorrect diluent may affect reagent stability and assay results.
- Thaw the frozen reagents at room temperature.
- Before use, allow all kit's reagents to warm up at room temperature then
  - homogenise buffer and diluent with a vortex
  - centrifuge (NEVER vortex) the antibodies to gather all liquid at the bottom of the vial
- It is recommended to filter buffers before use.
- Antibody solutions must be prepared in individual vials and can be mixed prior to dispensing.

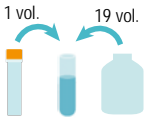
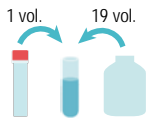
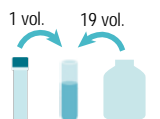
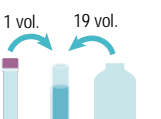
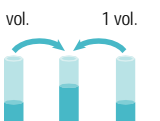
**TAKE CARE TO PREPARE STOCK AND WORKING SOLUTIONS ACCORDING TO THE DIRECTIONS FOR THE KIT SIZE YOU HAVE PURCHASED.**

**TO PREPARE DILUENT, STANDARD & ANTIBODY STOCK SOLUTIONS:**

500 TESTS		10,000 TESTS	
<b>IL12/23 p40 Eu Cryptate antibody</b>			
Thaw the IL12/23 p40 Eu Cryptate antibody. Centrifuge. This 20X stock solution can be frozen and stored at $\leq -60^{\circ}\text{C}$ .			Thaw the IL12/23 p40 Eu Cryptate antibody. Centrifuge. This 20X stock solution can be frozen and stored at $\leq -60^{\circ}\text{C}$ .
<b>IL12/23 p40 d2 antibody</b>			
Thaw the IL12/23 p40 d2 antibody. Centrifuge. This 20X stock solution can be frozen and stored at $\leq -60^{\circ}\text{C}$ .			Thaw the IL12/23 p40 d2 antibody. Centrifuge. This 20X stock solution can be frozen and stored at $\leq -60^{\circ}\text{C}$ .
<b>IL12/23 p40 Standard</b>			
Reconstitute the IL12/23 p40 standard with distilled water. Volume of reconstitution is indicated on the vial label. The reconstituted standard solution can be frozen and stored at $-60^{\circ}\text{C}$ or below.			Reconstitute the IL12/23 p40 standard with distilled water. Volume of reconstitution is indicated on the vial label. The reconstituted standard solution can be frozen and stored at $-60^{\circ}\text{C}$ or below.
<b>Diluent</b>			
Dilute 5-fold the 5X diluent #5 with distilled water: Homogenize the 5X diluent #5 with a vortex and add 1 volume of stock solution in 4 volumes of distilled water e.g. 1 mL of diluent + 4 mL of distilled water Mix gently after dilution.			Dilute 5-fold the 5X diluent #5 with distilled water: Homogenize the 5X diluent #5 with a vortex and add 1 volume of stock solution in 4 volumes of distilled water e.g. 10 mL of diluent + 40 mL of distilled water Mix gently after dilution

**TO PREPARE WORKING ANTIBODY SOLUTIONS:**

Each well requires 4  $\mu\text{L}$  of pre-mixed IL12/23 p40 antibodies. Prepare the two antibody solutions in separate vials

500 TESTS		10,000 TESTS	
<b>IL12/23 p40 Eu Cryptate antibody</b>			
Dilute 20-fold the 20 X stock solution (thawed reagent) of IL12/23 p40 Eu Cryptate antibody with detection buffer #3: e.g. 10 $\mu\text{L}$ of thawed Eu Cryptate antibody stock solution + 190 $\mu\text{L}$ of detection buffer.			Dilute 20-fold the 20 X stock solution (thawed reagent) of IL12/23 p40 Eu Cryptate antibody with detection buffer #3: e.g. 10 $\mu\text{L}$ of thawed Eu Cryptate antibody stock solution + 190 $\mu\text{L}$ of detection buffer).
<b>IL12/23 p40 d2 antibody</b>			
Dilute 20-fold the 20 X stock solution (thawed reagent) of IL12/23 p40 d2 antibody with detection buffer #3: e.g. 10 $\mu\text{L}$ of thawed d2 antibody stock solution + 190 $\mu\text{L}$ of detection buffer.			Dilute 20-fold the 20 X stock solution (thawed reagent) of IL12/23 p40 d2 antibody with detection buffer #3: e.g. 10 $\mu\text{L}$ of thawed d2 antibody stock solution + 190 $\mu\text{L}$ of detection buffer.
<b>Antibody mix</b>			
Pre-mix the two ready-to-use antibody solutions just prior to dispensing the reagents: e.g. 200 $\mu\text{L}$ of d2 antibody + 200 $\mu\text{L}$ of Eu Cryptate antibody			Pre-mix the two ready-to-use antibody solutions just prior to dispensing the reagents: e.g. 200 $\mu\text{L}$ of d2 antibody + 200 $\mu\text{L}$ of Eu Cryptate-antibody

**TO PREPARE WORKING STANDARDS SOLUTIONS:**

- Each well requires 16 µL of standard.
- Serially dilute the standard stock solution with diluent #5 or with the cell culture medium used to prepare your samples, supplemented with BSA or 10% FCS.
- **Due to the stability of the IL12/23 p40, it is mandatory to prepare the standard curve just before the assay.**
- In order to check for a potential interference effect from your own assay buffer when using the assay for the first time, we highly recommend the parallel preparation of a standard curve in your own supplemented cell culture medium and in diluent.
- In order to counteract any standard sticking, we recommend changing tips between each dilution.

A recommended standard dilution procedure is listed and illustrated below:

1. Reconstitute the standard vial with the volume indicated on the vial label using distilled water.
2. Prepare the following dilutions:

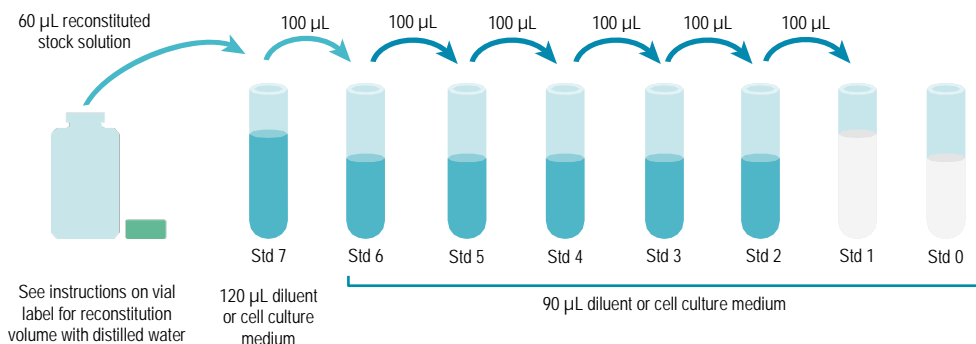
- Dilute the reconstituted standard stock solution 3-fold with diluent or with cell culture medium.

In practice: take 60 µL of stock solution and add it to 120 µL of diluent or cell culture medium. Mix gently. This yields the high standard (Std 7: 3000 pg/mL) for the top of the curve.

- Use the high standard (Std 7) to prepare the standard curve using serial dilutions as follows:

- Dispense 90 µL of diluent or cell culture medium into each vial from Std 6 to Std 0
- Add 100 µL of standard to 90 µL of diluent or cell culture medium, mix gently and repeat the serial dilution to make standard solutions: std6, std5, std4, std3, std2, std1

This will create 7 standards for the analyte. Std 0 (Negative control) is diluent or appropriate culture medium alone.







STANDARD	SERIAL DILUTIONS	WORKING SOLUTIONS
Standard Stock solution	Reconstitute the vial following the indications given on the vial label	30000 pg/mL
Standard 7	60 µL reconstituted standard stock solution + 120 µL diluent	10000 pg/mL
Standard 6	100 µL Standard 7 + 90 µL diluent	5263 pg/mL
Standard 5	100 µL Standard 6 + 90 µL diluent	2770 pg/mL
Standard 4	100 µL Standard 5 + 90 µL diluent	1458 pg/mL
Standard 3	100 µL Standard 4 + 90 µL diluent	767 pg/mL
Standard 2	100 µL Standard 3 + 90 µL diluent	404 pg/mL
Standard 1	100 µL Standard 2 + 90 µL diluent	213 pg/mL
Standard 0	90 µL diluent	0

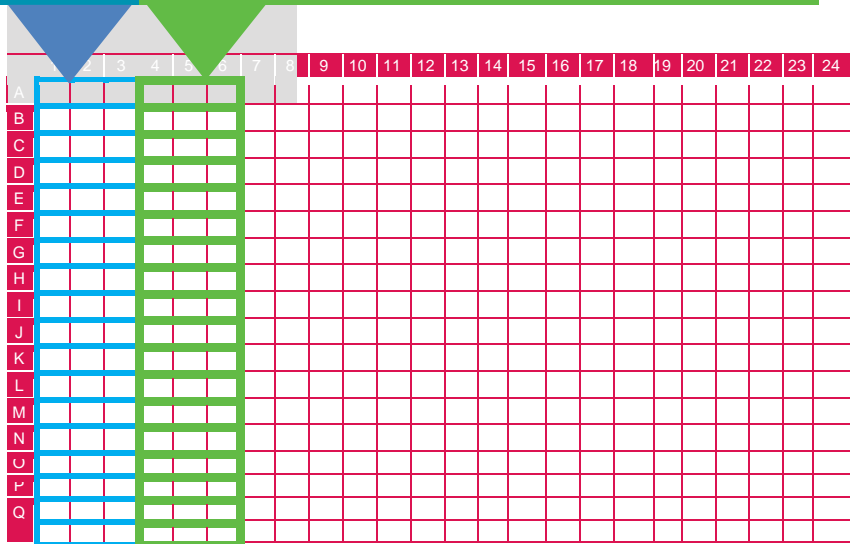
**TO PREPARE SAMPLES**

- Each well requires 16 µL of sample.
- Just after their collection, store the samples at 4°C and test them immediately. For later use, samples should be dispensed into disposable plastic vials and stored at ≤-60°C. Avoid multiple freeze/thaw cycles.
- All samples with a concentration above the highest standard (Std 7) must be diluted in diluent #5 or in your cell culture medium.

**ASSAY MANUAL**

		STANDARD (STD 0 - STD 7)	SAMPLES
Step 1		Dispense 16 µL of each IL12/23 p40 standard (Std 0 - Std 7) into each standard well.	Dispense 16 µL of each sample into each sample well.
Step 2		Dispense 4 µL of pre-mixed IL12/23 p40 antibodies working solution into all wells.	
Step 3		Seal the plate and incubate overnight at room temperature.	
Step 4		Remove the plate sealer and read on an HTRF® compatible reader.	

	1	2	3	4	5	6
A	16 µL Std 0 4 µL pre-mixed IL12/23 p40 antibodies	Repeat Well A1	Repeat Well A1	16 µL sample 1 4 µL pre-mixed IL12/23 p40 antibodies	Repeat Well A4	Repeat Well A4
B	16 µL Std 1 4 µL pre-mixed IL12/23 p40 antibodies	Repeat Well B1	Repeat Well B1	16 µL sample 2 4 µL pre-mixed IL12/23 p40 antibodies	Repeat Well B4	Repeat Well B4
C	16 µL Std 2 4 µL pre-mixed IL12/23 p40 antibodies	Repeat Well C1	Repeat Well C1	16 µL sample 3 4 µL pre-mixed IL12/23 p40 antibodies	Repeat Well C4	Repeat Well C4
D	16 µL Std 3 4 µL pre-mixed IL12/23 p40 antibodies	Repeat Well D1	Repeat Well D1	16 µL sample ... 4 µL pre-mixed IL12/23 p40 antibodies	Repeat Well D4	Repeat Well D4
E	16 µL Std 4 4 µL pre-mixed IL12/23 p40 antibodies	Repeat Well E1	Repeat Well E1	16 µL sample ... 4 µL pre-mixed IL12/23 p40 antibodies	Repeat Well E4	Repeat Well E4
F	16 µL Std 5 4 µL pre-mixed IL12/23 p40 antibodies	Repeat Well F1	Repeat Well F1	16 µL sample ... 4 µL pre-mixed IL12/23 p40 antibodies	Repeat Well F4	Repeat Well F4
G	16 µL Std 6 4 µL pre-mixed IL12/23 p40 antibodies	Repeat Well G1	Repeat Well G1	16 µL sample ... 4 µL pre-mixed IL12/23 p40 antibodies	Repeat Well G4	Repeat Well G4
H	16 µL Std 7 4 µL pre-mixed IL12/23 p40 antibodies	Repeat Well H1	Repeat Well H1	16 µL sample ... 4 µL pre-mixed IL12/23 p40 antibodies	Repeat Well H4	Repeat Well H4



## DATA REDUCTION & INTERPRETATION

1. Calculate the ratio of the acceptor and donor emission signals for each individual well.

$$\text{Ratio} = \frac{\text{Signal 665 nm}}{\text{Signal 620 nm}} \times 10^4$$

2. Calculate the delta ratio of the acceptor and donor emission signals for each individual well. The Standard 0 (Negative control) plays the role of an internal assay control.

$$\text{delta Ratio} = \text{Ratio Standard or sample} - \text{Ratio Standard 0}$$

3. Calculate the % CVs. The mean and standard deviation can then be worked out from ratio replicates.

$$\text{CV(\%)} = \frac{\text{Standard deviation}}{\text{Mean Ratio}} \times 100$$

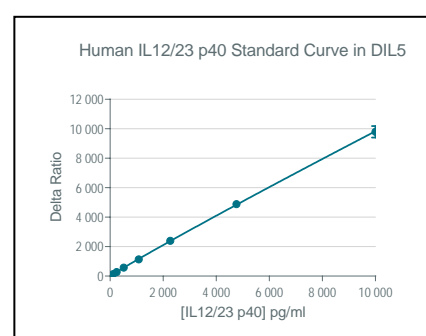
For more information about data reduction, please visit [www.revity.com](http://www.revity.com)

## RESULTS

This data must not be substituted for the data obtained in the laboratory, and should be considered only as an example. Results may vary from one HTRF® compatible reader to another.

Standard curve fitting with the 4 Parameter Logistic (4PL 1/y2) model (For more information about curve fitting please visit [www.revity.com](http://www.revity.com))

		Ratio (1)	delta R (2)	CV% (3)
Standard 0	Negative control	417	0	1%
Standard 1	213 pg/mL	555	138	3%
Standard 2	404 pg/mL	693	276	4%
Standard 3	767 pg/mL	997	579	4%
Standard 4	1458 pg/mL	1557	1139	5%
Standard 5	2770 pg/mL	2813	2396	2%
Standard 6	5263 pg/mL	5301	4883	4%
Standard 7	10000 pg/mL	10207	9790	4%



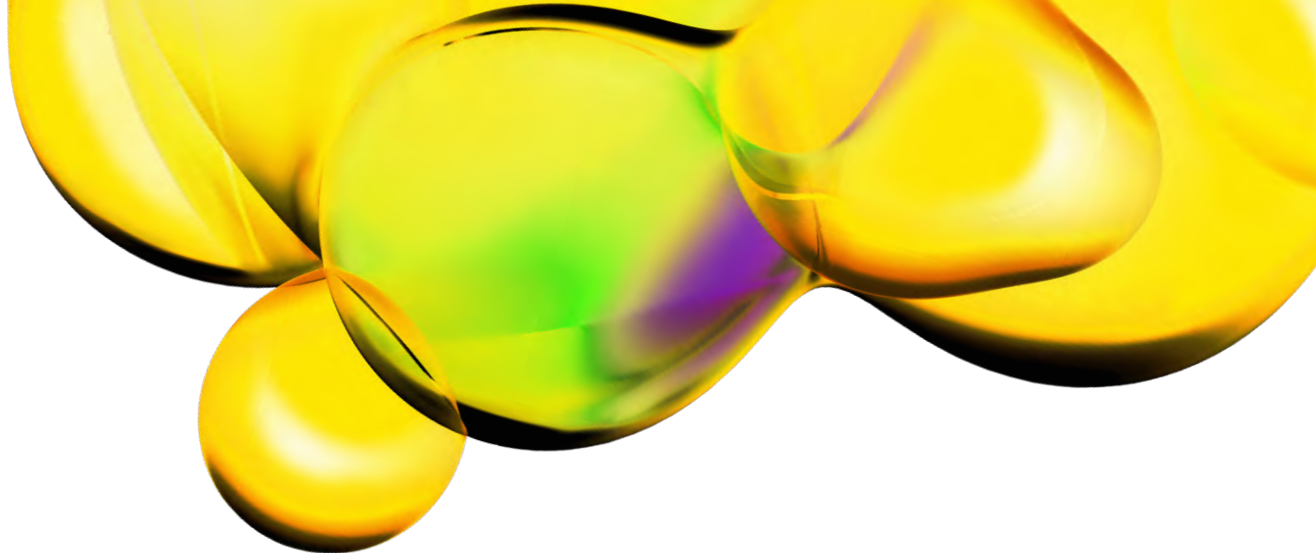
## ANALYTICAL ASSAY PERFORMANCE

	Diluent	DMEM	RPMI
Assay range (pg/mL**)	71 pg/mL to 10000 pg/mL		
Limit of detection (LoD*) = Std 0 mean + 2 SD	15 pg/mL	56 pg/mL	55 pg/mL
Limit of quantification (LoQ*)	71 pg/mL	96 pg/mL	109 pg/mL
Incubation time	overnight at room temperature		

\*\*NIBSC (95/544) value (IU/mL) = 0,01 x HTRF hIL12/23 p40 value (pg/mL). Note that this NIBSC I.S. equivalence is given is for IL12p70 as there is no I.S. for p40.

\*The analytical sensitivity was calculated from data obtained with an HTRF compatible reader after overnight incubation, this may vary from one HTRF compatible reader to another.

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