



MANUAL

Technology: HTRF[™] Cytokine

HTRF Human CCL17 Kit

| Part number: | 62HCCL17PEG | 62HCCL17PEH |
|--------------|-------------|--------------|
| Test size | 500 tests | 10,000 tests |

Storage: ≤ -16°C

Version: 01

Date: January 2025

ASSAY PRINCIPLE

This kit is intended for the simple and rapid quantification of human CCL17 in supernatant and offers a fast alternative to ELISA.

The detection principle of this kit is based on HTRF[™] technology (Homogeneous Time-Resolved Fluorescence). As shown in Figure 1, CCL17 is detected in a sandwich assay by using anti-CCL17 antibody labelled with Europium cryptate (donor), and anti-CCL17 antibody labelled with d2 (acceptor).

When the dyes are in close proximity, the excitation of the donor with a light source (laser or flash lamp) triggers a Fluorescence Resonance Energy Transfer (FRET) towards the acceptor, which in turn fluoresces at a specific wavelength (665 nm). Signal intensity is proportional to the number of antigen-antibody complexes formed and therefore to the CCL17 concentration.



Figure 1: Principle of HTRF CCL17 sandwich assay

PROTOCOL AT A GLANCE



Make sure to use the set-up for Eu Cryptate.

MATERIAL PROVIDED

| KIT COMPONENTS | 500 TES ⁻ | rs* | 10,000 TESTS* | | | | |
|--|----------------------|-------------------|---------------|------------|-----------------|--|--|
| CCL17 Standard Lyophilized | green cap | 2 vials | I | green cap | 2 vial | | |
| CCL17 Eu Cryptate Antibody Frozen 20X | orange cap | 1 vial 50 µL | | red cap | 1 vial 1 mL | | |
| CCL17 d2 Antibody Frozen 20X | blue cap | 1 vial 50 µL | | purple cap | 1 vial 1 mL | | |
| Diluent** #5 5X | yellow cap | 1 vial 2 mL | | White cap | 1 vial 10 mL | | |
| Detection Buffer*** #3 Ready-to-use | transparent cap | 2 vials 1.5 mL | | red cap | 1 vial 50 mL | | |

* When used as advised, the two available kit sizes will provide sufficient reagents for 500 tests and 10,000 tests respectively in 20 μL final volume. Assay volumes can be adjusted proportionally to run the assay in 96 or 1536 well microplates.

** Medium like cell culture medium can be an alternative to the diluent.

*** The Detection buffer is used to prepare working solutions of acceptor and donor reagents.

Purchase separately

- HTRF[™]-Certified Reader. Make sure the setup for Eu Cryptate is used. For a list of HTRF-compatible readers and set-up recommendations, please visit our website.
- Small volume (SV) detection microplates. Use white plate only. For more information about microplate recommendations, please visit our website.

STORAGE AND STABILITY

Kit

- Store the kit at -17°C or below.
- Under proper storage conditions, reagents are stable until the expiry date indicated on the label.
- Diluent and detection buffer are shipped frozen but can be stored at 2-8°C in your premises.

Reagents

- If lyophilized, reconstituted reagents, antibodies, and standard stock solutions may be frozen and thawed only once. To avoid freeze/thaw cycles, it is recommended to dispense remaining stock solutions into disposable plastic vials for storage at -16°C or below.
- Volume of Human CCL17 standard aliquots should not be under 10 μ L.

REAGENT PREPARATION

Before you begin

- It is very important to prepare reagents in the specified buffers. The use of an incorrect diluent may affect reagent stability and assay results.
- Thaw the frozen reagents at room temperature, allow them to warm up to room temperature for at least 30 mins before use.
- Before use, allow Diluent and Detection buffer to warm up at room temperature and homogenize them with a vortex.
- Antibody solutions must be prepared in individual vials and can be mixed prior to dispensing.
- CCL17 standards (for standard curve) must be prepared in diluent or in the same medium as the samples.

Take care to prepare stock and working solutions according to the directions for the kit size you have purchased.

To prepare reagent stock solutions



The Detection buffer is ready-to-use.

To prepare antibody working solutions

Each well requires 2 µL of CCL17-Eu Cryptate Antibody and 2 µL of CCL17 d2 Antibody. Prepare the two antibody solutions in separate vials.

| 500 TESTS | | | 10,000 TESTS |
|--|----------------|----------------|--|
| | CCL17 Eu Cry | ptate antibody | |
| Dilute 20-fold the 20X stock solution (thawed reagent) of CCL17 Eu Cryptate antibody with Detection buffer #3: add 1 volume of Eu Cryptate antibody stock solution in 19 volumes of detection buffer (e.g. 10 µL of Eu Cryptate antibody stock solution + 190 µL of detection buffer). | 1 vol. 19 vol. | 1 vol. 19 vol. | Dilute 20-fold the 20X stock solution (thawed reagent) of CCL17 Eu Cryptate antibody with Detection buffer #3: add 1 volume of Eu Cryptate antibody stock solution in 19 volumes of detection buffer (e.g. 10 µL of Eu Cryptate antibody stock solution + 190 µL of detection buffer). |
| | CCL17 d2 | 2 antibody | |
| Dilute 20-fold the 20X stock solution (thawed reagent) of CCL17 d2 antibody with Detection buffer #3: add 1 volume of d2 antibody stock solution in 19 volumes of detection buffer (e.g. 10 µL of Eu Cryptate antibody stock solution + 190 µL of detection buffer). | 1 vol. 19 vol. | 1 vol. 19 vol. | Dilute 20-fold the 20X stock solution (thawed reagent) of CCL17 d2 antibody with Detection buffer #3: add 1 volume of d2 antibody stock solution in 19 volumes of detection buffer (e.g. 10 µL of Eu Cryptate antibody stock solution + 190 µL of detection buffer). |
| | Antibo | ody Mix | |
| It is possible to pre-mix the two ready-to-use antibody solutions just prior to dispensing the reagents by adding 1 volume of d2 antibody solution to 1 volume of Cryptate antibody solution (e.g. 1 mL of d2 antibody + 1 mL of Cryptate antibody). | 1 vol. | 1 vol. | It is possible to pre-mix the two ready-to-use antibody solutions just prior to dispensing the reagents by adding 1 volume of d2 antibody solution to 1 volume of Cryptate antibody solution (e.g. 20 mL of d2 antibody + 20 mL of Cryptate antibody). |

To prepare working standards solutions

- Each well requires 16 µL of standard.
- Dilute the standard stock solution serially with diluent #5 (1X)
- If culture medium is used to dilute the standard, we recommend to supplement it with serum (2 to 10%) or BSA (0.2 to 1%) in order to avoid CCL17 sticking to assay plates.
- In order to check for a potential interference effect from your own assay buffer when using the assay for the first time, we highly recommend the parallel preparation of a standard curve in your own supplemented cell culture medium and in diluent #5 (1X).
- In order to counteract any standard sticking, we recommend changing tips between each dilution.

A recommended standard dilution procedure is listed and illustrated below:

1. Reconstitute the standard vial with the volume indicated on the vial label using distilled water (DO NOT vortex). Wait for at least 15 min post-reconsistution before preparing the standard dilutions.

2. Prepare the following dilutions:

Dilute the standard stock solution 3-fold with diluent; this yields the Standard Max solution (5,500 pg/mL) Dilute the standard stock solution 3-fold with diluent #5 (1X) to prepare high standard (Std 7): e.g. take 60 μ L of standard stock solution and add it to 120 μ L of diluent #5 (1X). Mix gently.

Use the high standard (Std 7) to prepare the standard curve using serial dilutions as follows:

- Dispense 110 μ L of diluent #5 (1X) in each vial from Std 6 to Std 0.
- Add 100 µL of standard to 110 µL of diluent #5 (1X), mix gently and repeat the serial dilution to make standard solutions: std6, std5, std4, std3, std2, std1.

This will create 7 standards for the analyte. Std 0 (Negative control) is diluent #5 (1X) or appropriate culture medium alone



¹²⁰ µL diluent or appropriate medium

130 µL diluent or appropriate medium

| STANDARD | SERIAL DILUTIONS | WORKING SOLUTIONS |
|-------------------------|---|-------------------|
| Standard Stock solution | Reconstituted lyophilisate | 16.5 ng/mL |
| Standard 7 | 60 μL Standard stock Solution + 120 μL diluent 1X | 5 500 pg/mL |
| Standard 6 | 100 μL standard 7 + 130 μL diluent 1X | 2391.3 pg/mL |
| Standard 5 | 100 μL standard 6 + 130 μL diluent 1X | 1039.7 pg/mL |
| Standard 4 | 100 μL standard 5 + 130 μL diluent 1X | 452 pg/mL |
| Standard 3 | 100 μL standard 4 + 130 μL diluent 1X | 196.5 pg/mL |
| Standard 2 | 100 μL standard 3 + 130 μL diluent 1X | 85.5 pg/mL |
| Standard 1 | 100 μL standard 2 + 130 μL diluent 1X | 37.2 pg/mL |
| Standard 0 | 130 µL diluent 1X | 0 |

To prepare samples

- Each well requires 16 µL of sample.
- Just after their collection, put the samples at 4°C and test them immediately. For later use, samples should be dispensed into disposable plastic vials and stored at -16°C or below. Avoid multiple freeze/thaw cycles.
- Samples with a concentration above the highest standard (Std 7) must be diluted diluent in your appropriate sample medium.
- To obtain additional information or support, please contact the HTRF technical support team.

ASSAY PROTOCOL

| | | STANDARD (STD 0 – STD 7) | SAMPLES | | | | | | |
|--------|---|--|--|--|--|--|--|--|--|
| Step 1 | | Dispense 16 µL of each CCL17 standard (Std 0 - Std 7) into each standard well | Dispense 16 µL of each sample into each sample well | | | | | | |
| Step 2 | | Add 2 µL of CCL17-d2 antibody working solution to all wells | | | | | | | |
| Step 3 | | Add 2 µL of CCL17-Eu Cryptate antibody working solution to all wells. | | | | | | | |
| Step 4 | Ġ | Seal the plate and incubate from 2 hours at RT | | | | | | | |
| Step 5 | | Remove the plate sealer and read | on an HTRF™ compatible reader | | | | | | |

| | 1 | 2 | 3 | 4 | 5 | 6 | |
|---|---|----------------------|-------------------|--|----------------|----------------|--|
| A | 16 μL Std 0 (Negative control) 2 μL CCL17-d2 2 μL CCL17-Eu Cryptate | Repeat Well A1 | Repeat Well A1 | <mark>16 μL sample 1</mark> 2 μL CCL17-d2 2 μL CCL17-Eu Cryptate | Repeat Well A4 | Repeat Well A4 | |
| В | 16 μL Std 1 2 μL CCL17-d2 2 μL CCL17-Eu Cryptate | Repeat Well B1 | Repeat Well B1 | <mark>16 μL sample 2</mark> 2 μL CCL17-d2 2 μL CCL17-Eu Cryptate | Repeat Well B4 | Repeat Well B4 | |
| с | 16 μL Std 2 2 μL CCL17-d2 2 μL CCL17-Eu Cryptate | Repeat Well C1 | Repeat Well C1 | <mark>16 μL sample 3</mark> 2 μL CCL17-d2 2 μL CCL17-Eu Cryptate | Repeat Well C4 | Repeat Well C4 | |
| D | <mark>16 μL Std</mark> 3 2 μL CCL17-d2 2 μL CCL17-Eu Cryptate | Repeat Well D1 | Repeat Well D1 | <mark>16 μL sample</mark> 2 μL CCL17-d2 2 μL CCL17-Eu Cryptate | Repeat Well D4 | Repeat Well D4 | |
| E | 16 μL Std 4 2 μL CCL17-d2 2 μL CCL17-Eu Cryptate | Repeat Well E1 | Repeat Well E1 | <mark>16 μL sample</mark> 2 μL CCL17-d2 2 μL CCL17-Eu Cryptate | Repeat Well E4 | Repeat Well E4 | |
| F | 16 μL Std 5 2 μL CCL17-d2 2 μL CCL17-Eu Cryptate | Repeat Well F1 | Repeat Well F1 | <mark>16 μL sample</mark> 2 μL CCL17-d2 2 μL CCL17-Eu Cryptate | Repeat Well F4 | Repeat Well F4 | |
| G | 16 μL Std 6 2 μL CCL17-d2 2 μL CCL17-Eu Cryptate | Repeat Well G1 | Repeat Well G1 | 16 μL sample 2 μL CCL17-d2 2 μL CCL17-Eu Cryptate | Repeat Well G4 | Repeat Well G4 | |
| н | 16 μL Std 7 2 μL CCL17-d2 2 μL CCL17-Eu Cryptate | Repeat Well H1 | Repeat Well H1 | <mark>16 μL sample</mark> 2 μL CCL17-d2 2 μL CCL17-Eu Cryptate | Repeat Well H4 | Repeat Well H4 | |

| | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 | 11 | 12 | 13 | 14 | 15 | 16 | 17 | 18 | 19 | 20 | 21 | 22 | 23 | 24 |
|---|---|---|---|---|---|---|---|---|---|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|
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DATA REDUCTION & INTERPRETATION

1) Calculate the ratio of the acceptor and donor emission signals for each individual well.

Ratio =
$$\frac{\text{Signal } 665 \text{ nm}}{\text{Signal } 620 \text{ nm}} \times 10^4$$

 Calculate the % CVs. The mean and standard deviation can then be worked out from ratio replicates.

CV (%)=
$$\frac{\text{Standard deviation}}{\text{Mean Ratio}}$$
 × 100

For more information about data reduction, please visit our website.

RESULTS

This data must not be substituted for the data obtained in the laboratory, and should be considered only as an example.

Results may vary from one HTRF[™] compatible reader to another.

Standard curve fitting with the 4 Parameter Logistic (4PL with $1/Y^2$) model

| | | Ratio (1) | CV% (2) |
|------------|---------------------|--------------|---------|
| Standard 0 | Negative control | 1389 | 7.5% |
| Standard 7 | 5 500 pg/mL | 24362 | 2.3% |
| Standard 6 | 2391.3 pg/mL | 19800 | 2.8% |
| Standard 5 | 1039.7 pg/mL | 10092 | 2.6% |
| Standard 4 | 452 pg/mL | 5623 | 4.2% |
| Standard 3 | 196.5 pg/mL | 3312 | 1.5% |
| Standard 2 | 85.5 pg/mL | 2452 | 7.0% |
| Standard 1 | 37.2 pg/mL | 1724 | 4.0% |



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