



CAMP - GS DYNAMIC KIT

CAMP - GS DYNAMIC KIT:

Part #: 62AM4PEB (1,000 tests), 62AM4PEC (20,000 tests), 62AM4PEJ (100,000 tests)

Version: #10 of April 2024 **Storage temperature:** 2-8°C

For research use only. Not for use in diagnostic procedures.

ASSAY PRINCIPLE

Revvity cAMP - Gs Dynamic assay is a competitive immunoassay intended to measure cyclic AMP (cAMP) accumulation in cells. It enables the direct pharmacological characterization of compounds acting on Gs-coupled receptors in either adherent or suspension cells.

The principle is based on HTRF® technology. Native cAMP produced by cells or unlabeled cAMP (standard curve) compete with d2-labeled cAMP (red acceptor) for binding to monoclonal cAMP Europium Cryptate-labeled antibody (Europium donor). The specific signal is inversely proportional to the concentration of cAMP in the standard or sample (Fig. 1).

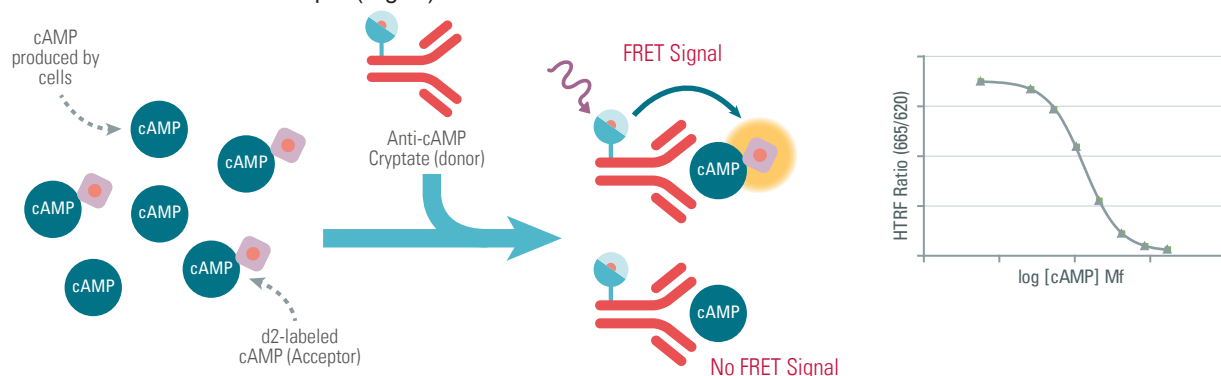


Figure 1: Principle of HTRF cAMP competitive binding assay.

As for all other HTRF assays, the calculation of the fluorescence ratio (665 nm/620 nm) eliminates any possible photophysical interferences and means the assay is unaffected by the experimental medium conditions (e.g. culture medium, serum, biotin, colored compounds, etc.).

MATERIALS & EQUIPMENT

MATERIALS PROVIDED:

NUMBER OF TESTS (CAT. #)	62AM4PEB (1,000 tests)	62AM4PEC (20,000 tests)	62AM4PEJ (100,000 tests)	REF#(when available separately)
Anti-cAMP-Cryptate (Europium donor, lyophilized)	1 vial	1 vial	5 vials	
cAMP - d2 (red acceptor, lyophilized)	1 vial	1 vial	5 vials	
cAMP - cAMP Gs Dynamic & Gi Standard Standard Concentrate (lyophilized)	1 vial	1 vial	1 vial	62AM4CDA
Lysis & Detection Buffer 1*	1 vial (13 mL)	-	-	62CL1FDD
Lysis & Detection Buffer 2*	-	1 vial (200 mL)	5 vials (200 mL ea)	62CL2FDF
Stimulation Buffer 1 (5X)**	1 vial (8 mL)	1 vial (100 mL)	5 vials (100 mL ea)	64SB1FDD (100 mL)

* Lysis & Detection Buffer is used to prepare working solutions of acceptor and donor reagents.

** Stimulation Buffer 1 is used to prepare working standard solutions, cells & compounds

Note: Reagent preparation instructions are included below

PURCHASE SEPARATELY:

cAMP ASSAY SPARE REAGENTS		
IBMX ⁽¹⁾ (PDE inhibitor, 500 mM)	62AMXADA (40 µL ⁽³⁾)	62AMXADC (500 µL ⁽³⁾)
Forskolin ⁽²⁾ (Adenylyl cyclase activator, 10 mM)	62AMYADA (40 µL ⁽³⁾)	62AMYADC (800 µL ⁽³⁾)

- Cell Culture Medium (appropriate for cell line used)
- Low volume white microplates⁽⁴⁾
- HTRF[®]-Certified Reader⁽⁵⁾

- (1) IBMX is the most commonly used phosphodiesterase Pan-inhibitor. It guarantees high level of cAMP accumulation in the cells.
- (2) Forskolin activates the adenylyl cyclase enzyme and increases the intracellular level of cAMP. It is used as a positive control for Gs applications (biological models validation, maximal intracellular cAMP modulation determination). It is highly used for Gi coupled receptor study as preactivation step to show cAMP level inhibition upon cell stimulation.
- (3) 100% DMSO Stock solution
- (4) Assay volumes can be adjusted proportionally to run the assay in 96- or 1536-well microplates. For more information about microplate recommendations, please visit our website at: www.revvy.com
- (5) For a list of HTRF-compatible readers and set-up recommendations, visit www.revvy.com

STORAGE AND STABILITY

Store the kit at 2-8°C until the expiration date indicated on the package.

Once reconstituted, stock solutions are stable at 2-8°C for **one week**. Stored at -16°C or below, they are stable for **6 months**. Stock solutions may be frozen and thawed only once. Freeze in aliquots to avoid multiple freeze/thaw cycles.

Working solutions of conjugates (1X) and standards (for standard dose response curve) can be stored at 2-8°C for up to **24 hours**. DO NOT FREEZE.

SPARE REAGENTS

Solutions of IBMX and Forskolin stored at 4°C are stable for **3 months**. Stock solution may be frozen and thawed only once. Freeze in aliquots to avoid multiple freeze/thaw cycles.

Note: Cryptate Antibody working solutions (prepared with frozen stock solutions) should be filtered before use to improve assay reproducibility.

REAGENT PREPARATION**BEFORE YOU BEGIN:**

1. It is very important to prepare reagents in the buffer specified. Use of an incorrect diluent may affect reagent stability and assay results.
2. Allow the lyophilized reagents to warm up to room temperature for at least 30 mins before reconstitution.
3. Working solutions must be used within **24 hours**. Do not freeze.
4. Stimulation buffer supplemented with IBMX must be prepared just before use
5. Forskolin (FSK) working solution must be prepared just before use
6. Conjugate solutions must be prepared in individual vials and dispensed separately for the assay.
7. cAMP standards (for standard curve) must be prepared in the same stimulation buffer as that used for the cell based assay supplemented with IBMX.
8. Stimulation Buffer 1 included in the kit should be replaced by cell culture medium if the compound stimulation time is above 2 h.
9. Take care to prepare stock and working solutions according to the directions below for the kit size you have purchased.
10. Be sure to set up your reader for Eu3+ Cryptate and read the fluorescence emission at two different wavelengths (665nm and 620nm) on a compatible HTRF[®] reader. (more information about HTRF[®] compatible readers and set-up recommendations available at www.revvy.com)

TO PREPARE STOCK SOLUTIONS:

If the kit is not going to be used at once, please consider aliquoting the stock solutions before freezing considering solutions can only be frozen/thawed once.

1,000 TESTS KIT	20,000/100,000 TESTS KITS
<p>1. Reconstitute the cAMP Eu Cryptate antibody (donor conjugate) with 1.1 mL distilled water. Mix gently. <i>This is a 5X stock solution which can be stored for one week at 2-8°C or frozen up to 6 months.</i></p> <p>2. Reconstitute the cAMP d2 reagent (acceptor conjugate) with 1.1 mL distilled water. Mix gently. <i>This is a 5X stock solution which can be stored for one week at 2-8°C or frozen up to 6 months.</i></p> <p>3. Reconstitute the cAMP Gs Dynamic & Gi Standard Concentrate using chosen stimulation buffer (e.g., DMEM, MEM, RPMI). See instructions on vial label for reconstitution volume. Mix gently after reconstitution. <i>The Standard Concentrate is stable frozen for 6 months.</i></p>	<p>1. Reconstitute the cAMP Eu Cryptate antibody (donor conjugate) with 5 mL distilled water. Mix gently. <i>This is a 20X stock solution which can be stored for one week at 2-8°C or frozen up to 6 months.</i></p> <p>2. Reconstitute the cAMP d2 reagent (acceptor conjugate) with 5 mL distilled water. Mix gently. <i>This is a 20X stock solution which can be stored for one week at 2-8°C or frozen up to 6 months.</i></p> <p>3. Reconstitute the cAMP Gs Dynamic & Gi Standard Concentrate using chosen stimulation buffer (e.g., DMEM, MEM, RPMI). See instructions on vial label for reconstitution volume. Mix gently after reconstitution. <i>The Standard Concentrate is stable frozen for 6 months.</i></p>

TO PREPARE WORKING SOLUTIONS:

1,000 TESTS KIT	20,000/100,000 TESTS KITS
<p>1. Stimulation Buffer 1: From the 5X stock solution: dilute 1 volume of Stimulation Buffer 1 (5X) in 4 volumes of distilled water</p>	<p>1. Stimulation Buffer 1: From 5X stock solution: dilute 1 volume of Stimulation Buffer 1 (5X) in 4 volumes of distilled water</p>
<p>2. Detection reagents: From the 5X stock solutions of cAMP Eu Cryptate antibody and cAMP d2 reagent (reconstituted reagents) dilute in separate vials, 1 volume of each stock solutions in 4 volumes of Lysis & detection Buffer 1 (e.g., 0.5 mL of reconstituted reagent + 2 mL of Lysis & Detection Buffer).</p>	<p>2. Detection reagents: From the 20X stock solutions of cAMP Eu Cryptate antibody and cAMP d2 reagent (reconstituted reagents) dilute in separate vials, 1 volume of each stock solutions in 19 volumes of Lysis & detection Buffer 2 (e.g., 0.5 mL of reconstituted reagent + 9.5 mL of Lysis & Detection Buffer)</p>

CAMP ASSAY SPARE REAGENTS

3. IBMX

IBMX Stock solution is provided at 500 mM in 100% DMSO. Dilute IBMX stock solution in the stimulation buffer at the desired concentration. An empirical concentration of working solution at 0,5 mM is recommended. (e.g., 40 µL of IBMX stock solution + 39 960 µL of stimulation buffer)

4. Forskolin

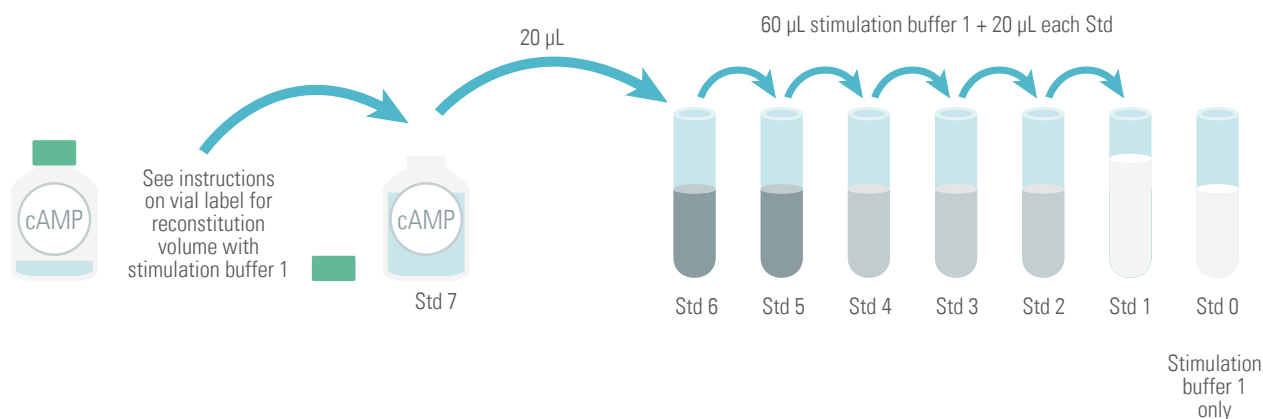
The forskolin stock solution is provided at 10 mM in DMSO.

Working solution preparation : it is recommended to prepare 2 sequential dilutions to avoid Forskolin precipitation . First, an intermediate dilution of 1/5 in 100% DMSO (e.g., 40 µL of stock solution + 160 µL of DMSO) , followed by a second dilution from this predilution in the stimulation buffer at the desired concentration.

STANDARD CURVE

Run a standard dose response curve to determine the **linear dynamic range** of the assay with your cell culture medium (Stimulation Buffer 1) in your laboratory. This will also verify that your assay is generating the expected S/B and IC50. In particular, the IC10 and IC90 will be useful in experiments to optimize the cell density of the cell lines you will stimulate with compounds.

TO PREPARE WORKING STANDARDS:



STANDARD	SERIAL DILUTIONS	cAMP WORKING SOLUTION nM	cAMP FINAL CONCENTRATION nM
Std7	Reconstituted cAMP Standard	2848*	712
Std6	20 µL Std7 + 60 µL Stimulation Buffer 1	712	178
Std5	20 µL Std6 + 60 µL Stimulation Buffer 1	178	44.5
Std4	20 µL Std5 + 60 µL Stimulation Buffer 1	44.5	11.1
Std3	20 µL Std4 + 60 µL Stimulation Buffer 1	11.1	2.78
Std2	20 µL Std3 + 60 µL Stimulation Buffer 1	2.78	0.69
Std1	20 µL Std2 + 60 µL Stimulation Buffer 1	0.69	0.17
Std0 (positive control)	60 µL Stimulation Buffer 1	0	0

*The actual concentration of cAMP after reconstituting the provided vial is indicated on the label of the maximum standard

STANDARD CURVE ASSAY MANUAL

	STANDARD (STD7-STD0)	NEGATIVE CONTROL
Step 1	Dispense 5 µL of each cAMP standard (Std7-Std0) into each standard well, in triplicate.	Dispense 5 µL Stimulation Buffer 1 into negative control wells, in triplicate.
Step 2	Add 5 µL of Stimulation Buffer 1 into each standard well.	Add 5 µL of Stimulation Buffer 1 into negative control wells.
Step 3	Add 5 µL of cAMP d2 reagent working solution into each standard well.	Add 5 µL of Lysis & Detection Buffer into negative control wells.
Step 4	Add 5 µL of cAMP Eu Cryptate antibody working solution into each standard well.	Add 5 µL of cAMP Eu Cryptate antibody working solution into negative control wells.
Step 5	Seal the plate and incubate 1 hour at room temperature.	Seal the plate and incubate 1 hour at room temperature.
Step 6	Remove the plate seal and read on an HTRF compatible reader.	Remove the plate seal and read on an HTRF compatible reader.

Suggested plate map:

	1	2	3
A	5 μ L Std 0 5 μ L Stimulation. Buffer 1 5 μ L cAMP d2 reagent 5 μ L cAMP Eu Cryptate antibody	Repeat Well A1	Repeat Well A1
B	5 μ L Std 1 5 μ L Stimulation. Buffer 1 5 μ L cAMP d2 reagent 5 μ L cAMP Eu Cryptate antibody	Repeat Well B1	Repeat Well B1
C	5 μ L Std 2 5 μ L Stimulation. Buffer 1 5 μ L cAMP d2 reagent 5 μ L cAMP Eu Cryptate antibody	Repeat Well C1	Repeat Well C1
D	5 μ L Std 3 5 μ L Stimulation. Buffer 1 5 μ L cAMP d2 reagent 5 μ L cAMP Eu Cryptate antibody	Repeat Well D1	Repeat Well D1
E	5 μ L Std 4 5 μ L Stimulation. Buffer 1 5 μ L cAMP d2 reagent 5 μ L cAMP Eu Cryptate antibody	Repeat Well E1	Repeat Well E1
F	5 μ L Std 5 5 μ L Stimulation. Buffer 1 5 μ L cAMP d2 reagent 5 μ L cAMP Eu Cryptate antibody	Repeat Well F1	Repeat Well F1
G	5 μ L Std 6 5 μ L Stimulation. Buffer 1 5 μ L cAMP d2 reagent 5 μ L cAMP Eu Cryptate antibody	Repeat Well G1	Repeat Well G1
H	5 μ L Std 7 5 μ L Stimulation. Buffer 1 5 μ L cAMP d2 reagent 5 μ L cAMP Eu Cryptate antibody	Repeat Well H1	Repeat Well H1
I	5 μ L Stimulation. Buffer 1 5 μ L Stimulation. Buffer 1 5 μ L Lysis & Detection Buffer 5 μ L cAMP Eu Cryptate antibody	Repeat Well I1	Repeat Well I1

	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23	24
A																								
B																								
C																								
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M																								
N																								
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	1	2	3
Standard curve			
Negative control (e.g. no cAMP d2 reagent)			

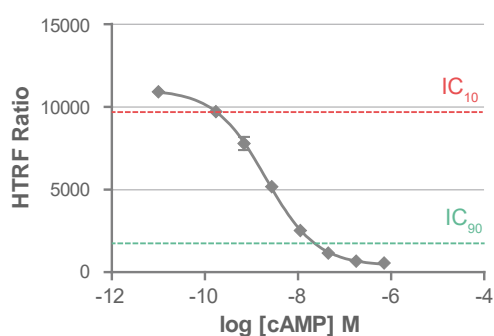
DATA REDUCTION & INTERPRETATION

1. Calculate the ratio of the acceptor and donor emission signals for each individual well.

$$\text{Ratio} = \frac{\text{Signal 665 nm}}{\text{Signal 620 nm}} \times 10^4$$

2. Plot the HTRF ratio versus the cAMP concentrations.

For more information about data reduction, please visit www.revvy.com



Characteristics of the assay relative to the IC_{50} and the signal over background.

Obtained using the PHERAstar Plus reader (BMG LABTECH).

A Guide to Optimizing Agonists Gs", "A Guide to Optimizing Antagonists Gs

	IC_{50} (nM)	S/B
Incubation 1 hour at RT	5.6	15

CELL-BASED ASSAY

BEFORE RUNNING A CELL-BASED ASSAY:








1. Various cell parameters must be optimized that depend on the type of compound to be screened (agonist or antagonist) and on the particular G protein coupled to the GPCR (G_i or G_s). These parameters include cell density, IBMX* concentration, agonist concentration (for antagonist mode assay), and stimulation time.
2. Cell density must be also optimized to ensure that cAMP levels of unstimulated and stimulated cells fall within the linear dynamic range of the assay (IC10 to IC90). If results fall outside the assay's linear range, the data will be inaccurate.
3. Compounds must be diluted in Stimulation Buffer 1 with the optimized IBMX concentration (recommended 0.5mM final) . Alternatively, for stimulation time above 2 hours, compounds could be diluted in cell culture medium supplemented with IBMX.
4. IBMX may have its own effect on cAMP accumulation. We recommend checking this effect by running experimental wells with and without IBMX in the stimulation buffer.
5. Set up your reader for Eu3+ Cryptate and read the fluorescence emission at two different wavelengths (665nm and 620nm) on a compatible HTRF® reader. (more information about HTRF® compatible readers and set-up recommendations available at www.revvity.com)

*A phosphodiesterase inhibitor must be added to the culture medium to prevent cAMP degradation and promote its accumulation.

Forskolin may be used as an optional control for your cell based assay.

For complete guidelines on fully optimizing your cAMP Gs assay, please refer to our Technical Guides: "A Guide to Optimizing Agonists Gs", "A Guide to Optimizing Antagonists Gs".

CELL-BASED ASSAY MANUAL

		NEGATIVE CONTROL*	NON-STIMULATED CELLS**	STIMULATED CELLS
Step 1		Dispense 5 µL of cells to all wells.	Dispense 5 µL cells to all wells.	Dispense 5 µL cells to all wells.
Step 2		Add 5 µL Stimulation Buffer 1.	Add 5 µL Stimulation Buffer 1.	Add 5 µL of test compounds (2X).
Step 3		Seal the plate and incubate for appropriate time at 37°C.	Seal the plate and incubate for appropriate time at 37°C.	Seal the plate and incubate for appropriate time at 37°C.
Step 4		Add 5 µL Lysis & Detection Buffer.	Add 5 µL of cAMP-d2 reagent reagent working solution.	Add 5 µL of cAMP-d2 reagent working solution.
Step 5		Add 5 µL of CAMP Eu-Cryptate antibody working solution.	Add 5 µL of CAMP Eu-Cryptate antibody working solution.	Add 5 µL of CAMP Eu-Cryptate antibody working solution.
Step 6		Seal the plate and incubate for appropriate time at RT.	Seal the plate and incubate for appropriate time at RT.	Seal the plate and incubate for appropriate time at RT.
Step 7		Remove the plate sealer and read on an HTRF compatible reader	Remove the plate sealer and read on an HTRF compatible reader	Remove the plate sealer and read on an HTRF compatible reader

*The Negative Control wells represent the non-specific fluorescence stemming from the Cryptate.

**Data with non-stimulated cells indicate the basal cellular level of cAMP before compound treatment.

DATA REDUCTION & INTERPRETATION

1. Calculate the ratio of the acceptor and donor emission signals for each individual well.

$$\text{Ratio} = \frac{\text{Signal 665 nm}}{\text{Signal 620 nm}} \times 10^4$$

2. Plot the HTRF ratio versus compound concentrations.

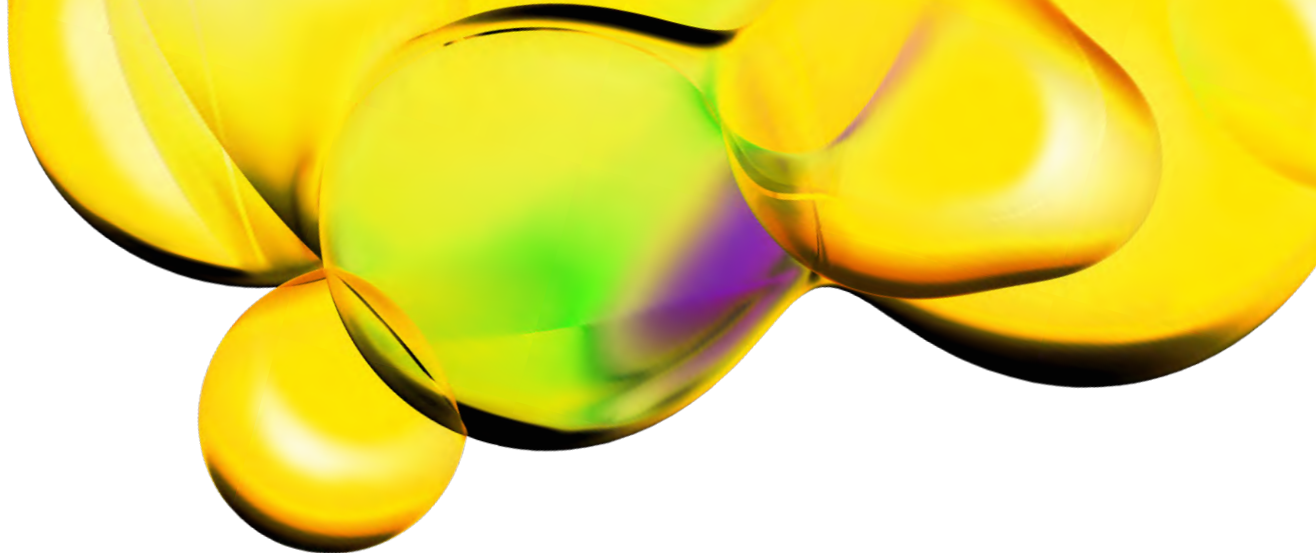
For more information about data reduction, please visit www.revvity.com

ASSAY FLEXIBILITY AND MINIATURIZATION

It is very easy to modify the assay (here described in 384-well, low volume format) to 1536-well (10 μL) or 96 half-well (100 μL) formats. Simply adjust the volume of each assay component proportionally in order to maintain the same reagent concentrations as the 20 μL assay. Assay performance is the same in 96 up to 1536-well plate format.

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