

# High-content analysis: advancing your knowledge to help combat infectious disease.

## Using high-content analysis to study infectious disease.

Infectious diseases caused by bacterial, viral or parasitic pathogens are a major burden to global health. The increased globalization of modern society, with travel and trade that facilitates the spread of emerging and re-emerging infectious diseases, and phenomena such as anti-microbial resistance, underscore the importance of the development of new preventative and therapeutic approaches.

High-content analysis plays a significant role in infectious disease research as reviewed by Ang and Pethe (2016), enabling high-throughput functional and phenotypic assays that can be adapted to a wide range of pathogens. Originally developed as a complementary technology to traditional biochemical high-throughput screening (HTS) in drug discovery, today high-content analysis is established in a far broader area of the life science space as an unbiased imaging method to assess cellular function. Applications include genetic siRNA interference or CRISPR screens for identifying host factors involved in host-pathogen interactions, but also compound screens for drug discovery.

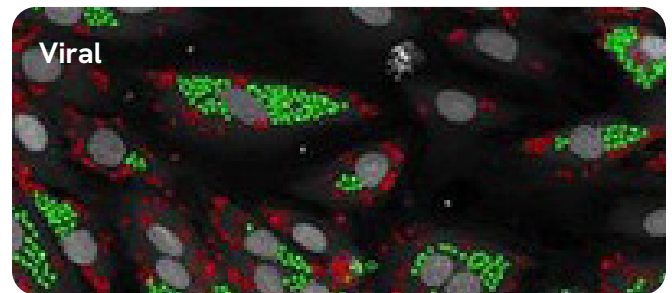


Image from Barrows NJ, Campos RK, Powell ST, Prasanth KR, Schott-Lerner G et al. A screen of FDA-approved drugs for inhibitors of Zikavirus infection. *Cell Host Microbe* 2016;20(2):259-70

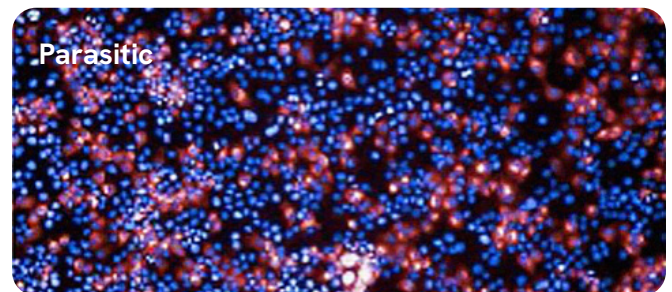
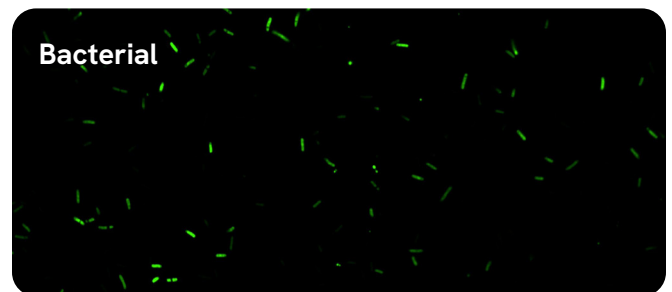


Image from Automated High-Content Assay for Compounds Selectively Toxic to *Trypanosoma cruzi* in a Myoblastic Cell Line. Alonso-Padilla, J. et al. *PLoS Negl Trop Dis*. 2015; Jan, 9(1): e0003493.



Confocal image of *Salmonella* bacteria labelled with GFP acquired on Opera Phenix™ with a 63x water objective NA 1.15. Sample courtesy of D. Goulding, Sanger Institute, UK.

## More meaningful results

Screening can be performed within the context of a host cell, allowing drugs that target host cell or microbial factors to be discovered. High-content screening allows the analysis of complex processes such as host cell entry, intracellular trafficking or cell-to-cell spread using fixed samples or live cell imaging.

## More information from cellular samples

Fully automated imaging and unbiased quantitative image analysis exploits the full potential of microscopy, allowing the characterization of pathogens as well as host cell phenotypes. Infection rates can be determined with high sensitivity, since individual infected cells can be identified. The process of infection can be described with

up to hundreds of readouts per cell allowing the linkage of infection rates to other readouts such as host cell morphology or host cell signalling.

## More questions answered simultaneously

Data sets are more information-rich compared to non-imaging read-outs, allowing a number of conclusions to be drawn from one experiment, such as drug efficacy against a pathogen as well as drug toxicity on host cells.

## More samples analyzed

The automation of imaging tasks allows the analysis of significantly more samples compared to conventional microscopy, enabling genome-wide or kinome-wide siRNA screens or screening of large compound libraries.

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## Viral disease - featured publications

### **Ebola virus modulates transforming growth factor $\beta$ signaling and cellular markers of mesenchyme-like transition in hepatocytes**

*Jason Kindrachuk and colleagues*

Ebola virus (EBOV) is a serious public health concern, causing severe hemorrhagic disease in humans with a mortality rate of more than 78%. The authors used a kinome analysis to investigate the host kinome response over time in human hepatocyte cells infected with EBOV. This showed that transforming growth factor  $\beta$  (TGF- $\beta$ )-mediated pathways are modulated during EBOV infection and replication. Phenotypic screening showed that EBOV infection is inhibited in the presence of TGF- $\beta$  and related pathway kinase inhibitors.

### **A screen of FDA-approved drugs for inhibitors of Zika virus infection**

*Nicholas Barrows and colleagues*

High-content screening and analysis of a panel of 774 FDA-approved drugs identified over 20 molecules that could inhibit Zika virus infection in human cells including cervical, placental, neural stem cells and primary amnion cells - all are potential new therapeutics in the fight against Zika.

### **Identification of proteins bound to dengue viral RNA *in vivo* reveals new host proteins important for virus replication**

*Stacia Phillips and colleagues*

An *in vivo* approach involving UV crosslinking, antisense-mediated affinity purification and mass spectrometry identified host proteins that physically associate with dengue virus RNA. Small interfering RNA-mediated gene silencing, combined with high-content screening demonstrated that over half of these proteins are likely to be involved in regulating dengue virus replication.

## Additional publications featuring high-content technologies in the study of viral disease

- Jiang B. et al. [Pharmacological Modulators of Epithelial Immunity Uncovered by Synthetic Genetic Tracing of SARS-CoV-2 Infection Responses](#). *Science Advances*. 2023.
- Ang ML and Pethe K. [Contribution of high-content imaging technologies to the development of anti-infective drugs](#). *Cytometry A*. 2016; Aug; 89(8):755-60.
- Barrows NJ. et al. [A Screen of FDA-Approved Drugs for Inhibitors of Zika Virus Infection](#). *Cell Host & Microbe*. 2016; 20(2):259-70.
- Cai Y. et al. [Simian hemorrhagic fever virus cell entry is dependent on CD163 and uses a clathrin-mediated endocytosis-like pathway](#). *Journal of Virology*. 2015; 89(1):844-856.
- Chang SY. et al. [A natural component from \*Euphorbia humifusa\* Willd displays novel, broad-spectrum anti-influenza activity by blocking nuclear export of viral ribonucleoprotein](#). *Biochemical and biophysical research communications*. 2016; 471(2):282-9.
- Chen H. et al. [BET-Inhibitors Disrupt Rad21-Dependent Conformational Control of KSHV Latency](#). *PLOS Pathogens*. 2017;13(1):e1006100.
- Cortjens B. et al. [Neutrophil Extracellular Traps Cause Airway Obstruction During Respiratory Syncytial Virus Disease](#). *The Journal of Pathology*. 2016; 238(3):401-411.
- Ekins S. et al. [Machine Learning Models Identify Molecules Active Against the Ebola Virus \*in vitro\*](#). *F1000 Research*. 2016; 4:1091.
- Kim H-Y. et al. [Benzothiazepinecarboxamides: Novel Hepatitis C Virus Inhibitors that Interfere with Viral Entry and the Generation of Infectious Virions](#). *Antiviral Research*. 2016; 129:39-46.
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- Ko, M. et al. [Screening of FDA-approved drugs using a MERS-CoV clinical isolate from South Korea identifies potential therapeutic options for COVID-19](#). *bioRxiv preprint*. 2019
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- McMullan LK. et al. [Characterisation of Infectious Ebola Virus from the Ongoing Outbreak to Guide Response Activities in the Democratic Republic of the Congo: A Phylogenetic and \*in Vitro\* Analysis](#). *The Lancet Infectious Diseases*. 2019; 19 (9): 1023-32.
- McMullan LK. et al. [The Lipid Moiety of Brincidofovir is Required for \*in vitro\* Antiviral Activity Against Ebola Virus](#). *Antiviral Research*. 2016; 125:71-78.
- Park JH. et al. [Identification of Novel Membrane-associated Prostaglandin E Synthase-1 \(mPGES-1\) Inhibitors with Anti-influenza Activities \*in vitro\*](#). *Biochemical and Biophysical Research Communications*. 2016; 469(4):848-855.
- Phillips SL. et al. [Identification of Proteins Bound to Dengue Viral RNA \*in vivo\* Reveals New Host Proteins Important for Virus Replication](#). *mBio*. 2016; 7(1):e01865-15.
- Postnikova EN. et al. [Scalable, Semi-Automated Fluorescence Reduction Neutralization Assay for Qualitative Assessment of Ebola Virus-Neutralizing Antibodies in Human Clinical Samples](#). *PLOS ONE*. 2019; 14 (8): e0221407.
- Radoshitzky SR. et al. [siRNA Screen Identifies Trafficking Host Factors that Modulate Alphavirus Infection](#). *PLoS Pathogens*. 2016; 12:e1005466.
- Wang C. et al. [The Serine/Threonine Kinase AP2-Associated Kinase 1 Plays an Important Role in Rabies Virus Entry](#). *Viruses*. 2019; 12 (1): 45.
- Wang G. et al. [The G Protein-Coupled Receptor FFAR2 Promotes Internalization during Influenza A Virus Entry](#). *Journal of Virology*. 2019; 94 (2).

## Bacterial disease - featured publications

### Testing chemical & genetic modulators in *Mycobacterium tuberculosis* infected cells using phenotypic assays

Vincent Delorme and colleagues

A better understanding of the intracellular stage of *M. tuberculosis* pathogenesis may aid development of new agents in the battle against tuberculosis infection. High-content screening was used to investigate the role of host genes in intracellular replication, identify small molecule inhibitors within the host cell, and examine the role of bacterial genes in intracellular trafficking.

### A dual microscopy-based assay to assess *Listeria monocytogenes* cellular entry and vacuolar escape

J.J. Quereda, J. Pizarro-Cerda and colleagues

Listeriosis-causing *L. monocytogenes* invades mammalian cells and escapes from the vacuole, enabling proliferation in the host cell cytoplasm. A two-step method involving high-content screening was developed to investigate cell invasion and vacuolar rupture in a single experiment, providing a powerful tool which could be used for identifying the factors involved in these processes in bacteria and other pathogens.

### B cell selection and therapeutic antibody characterization using the operetta high-content imaging system

Infection with the intestinal bacterium *Clostridium difficile* is the most common cause of healthcare-associated diarrhoea that can develop in patients after hospitalization and treatment with antibiotics. *C. difficile* is resistant to a wide range of antibiotics and, for this reason, new treatments for severe cases are desperately needed. *C. difficile* infection is mediated by the production of toxins by the bacterium, and treatment with toxin-binding agents, such as antibodies is a promising approach to reducing or inhibiting the clinical manifestations. This case study describes the workflow used by researchers at AIMM to generate antibodies against *C. difficile* ToxB.

### Additional publications featuring high-content technologies in the study of bacterial disease

- Aylan B. et al. [ATG7 and ATG14 Restrict Cytosolic and Phagosomal Mycobacterium Tuberculosis Replication in Human Macrophages](#). *Nat Microbiol.* 2023, 8 (5), 803-818.
- Abdelmohsen U, Cheng C, Reimer A, Kozjak-Pavlovic V, Ibrahim A, Rudel T. et al. [Antichlamydial Sterol from the Red Sea Sponge \*Callyspongia aff. implexa\*](#). *Planta Medica.* 2015;81(05):382-387.
- Ang ML and Pethe K. [Contribution of high-content imaging technologies to the development of anti-infective drugs](#). *Cytometry A.* 2016; Aug; 89(8):755-60.
- Blättner S, Das S, Paprotka K, Eilers U, Krischke M, Kretschmer D. et al. [Staphylococcus aureus Exploits a Non-ribosomal Cyclic Dipeptide to Modulate Survival within Epithelial Cells and Phagocytes](#). *PLoS Pathogens.* 2016;12(9):e1005857.
- Christophe T. et al. [High Content Screening Identifies Decaprenyl-phosphoribose 2' epimerase as a Target for Intracellular Antimycobacterial Inhibitors](#). *PLoS Pathogens.* 2009; 5(10):e1000645.
- Clare RH. et al. [Development and Validation of a Whole-Cell Screen: A Route to Macrolide Drugs against Onchocerciasis and Lymphatic Filariasis](#). *J Biomol Screen.* 2015; 20(1):64-9.
- Delorme V, Song O-R, Baulard A and Brodin P. [Testing Chemical and Genetic Modulators in Mycobacterium tuberculosis Infected Cells Using Phenotypic Assays](#). *Methods in Molecular Biology.* 2015; 1285:387-411.
- Mellouk N. et al. [Shigella Subverts the Host Recycling Compartment to Rupture its Vacuole](#). *Cell Host & Microbe.* 2014; 16(4):517-30.
- Knight M, Braverman J, Asfaha K, Gronert K, Stanley S. [Lipid droplet formation in Mycobacterium tuberculosis infected macrophages requires IFN- \$\gamma\$ /HIF-1 \$\alpha\$  signaling and supports host defense](#). *PLOS Pathogens.* 2018;14(1):e1006874.

- Pooja S, Pushpanathan M, Gunasekaran P, Rajendhran J. [Endocytosis-Mediated Invasion and Pathogenicity of \*Streptococcus agalactiae\* in Rat Cardiomyocyte \(H9C2\)](#). *PLoS ONE*. 2015;10(10):e0139733.
- Quereda JJ. et al. [A Dual Microscopy-based Assay to Assess \*Listeria monocytogenes\* Cellular Entry and Vacuolar Escape](#). *Applied and Environmental Microbiology*. 2016; 82(1):211-217.
- Queval CJ. et al. [STAT3 Represses Nitric Oxide Synthesis in Human Macrophages upon \*Mycobacterium tuberculosis\* Infection](#). *Scientific Reports*. 2016; 6:29297.
- Ranjani J, Pushpanathan M, Mahesh A, Niraimathi M, Gunasekaran P, Rajendhran J. [Pseudomonas aeruginosa PAO1 Induces Distinct Cell Death Mechanisms in H9C2 cells and its Differentiated Form](#). *Journal of Basic Microbiology*. 2015;55(10):1191-1202.

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## Parasitic disease - featured publications

### High-throughput assay and discovery of small molecules that interrupt malaria transmission

David M Plouffe and colleagues

The Saponin-lysis Sexual Stage Assay (SaLSSA) is a high-throughput, cost-effective assay for identifying small molecules with malaria transmission-blocking activity. A total of 13,983 compounds were analyzed, which included some with consistent low nanomolar transmission-blocking activity. The assay provides a tool for the discovery and development of transmission-blocking drugs.

### Additional publications featuring high-content technologies in the study of parasitic disease

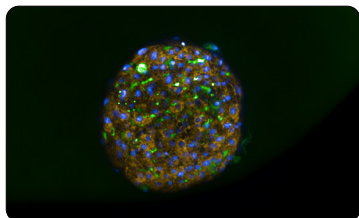
- Rosenthal MR. and Ng CL. [High-Content Imaging as a Tool to Quantify and Characterize Malaria Parasites](#). *Cell Reports Methods*. 2023; 3: 100516.
- Ang ML and Pethe K. [Contribution of high-content imaging technologies to the development of anti-infective drugs](#). *Cytometry A*. 2016; Aug; 89(8):755-60.
- Alonso-Padilla J. et al. [Automated High-Content Assay for Compounds Selectively Toxic to \*Trypanosoma cruzi\* in a Myoblastic Cell Line](#). *PLoS Neglected Tropical Diseases*. 2015; 9(1):e0003493.
- Clare R, Cook D, Johnston K, Ford L, Ward S, Taylor M. [Development and Validation of a High-Throughput Anti-Wolbachia Whole-Cell Screen A Route to Macroparasitic Drugs against Onchocerciasis and Lymphatic Filariasis](#). *Journal of Biomolecular Screening*. 2015;20(1):64-69.
- De Rycker M. et al. [Identification of Trypanocidal Activity for Known Clinical Compounds Using a New \*Trypanosoma cruzi\* Hit-Discovery Screening Cascade](#). *PLoS Neglected Tropical Diseases*. 2016; 10(4):e0004584.
- Duez J. et al. [Splenic Retention of \*Plasmodium falciparum\* Gametocytes to Block the Transmission of Malaria](#). *Antimicrobial Agents and Chemotherapy*. 2015; 59(7):4206-4214.
- Fletcher S, Lucantoni L, Sykes M, Jones A, Holleran J, Saliba K et al. [Biological characterization of chemically diverse compounds targeting the \*Plasmodium falciparum\* coenzyme A synthesis pathway](#). *Parasites & Vectors*. 2016;9(1).
- Hart C, Munro T, Andrews K, Ryan J, Riches A, Skinner-Adams T. [A Novel \*in vitro\* Image-Based Assay Identifies New Drug Leads for Giardiasis](#). *International Journal for Parasitology: Drugs and Drug Resistance*. 2017;7(1):83-89.
- Khare S. et al. [Anti-Trypanosomal Treatment with Benznidazole is Superior to Posaconazole Regimens in Mouse Models of Chagas Disease](#). *Antimicrobial Agents and Chemotherapy*. 2015; 59(10):6385-6394.
- Khare S. et al. [Utilizing Chemical Genomics to Identify Cytochrome b as a Novel Drug Target for Chagas Disease](#). *PLoS Pathogen*. 2015; 11(7):e1005058.
- Lucantoni L. et al. [A Simple and Predictive Phenotypic High Content Imaging Assay for \*Plasmodium falciparum\* Mature Gametocytes to Identify Malaria Transmission Blocking Compounds](#). *Scientific Reports*. 2015; 5:16414.
- Plouffe DM. et al. [High-Throughput Assay and Discovery of Small Molecules that Interrupt Malaria Transmission](#). *Cell Host and Microbe*. 2016; 19(1):114-126.



- Roth A, Maher SP, Conway AJ, Ubalee R, Chaumeau V, Andolina C et al. [A comprehensive model for assessment of liver stage therapies targeting Plasmodium vivax and Plasmodium falciparum](#) *Nature Communications* 2018; 9:(1837)
- Siquella X, Nebl T, Thompson J, Whitehead L, Malpede B, Salinas N. et al. [Plasmodium Falciparum Ligand Binding to Erythrocytes Induce Alterations in Deformability Essential for Invasion.](#) *eLife*. 2017; 6:e21083.
- Sykes ML and Avery VM. [Development and Application of a Sensitive, Phenotypic, High-throughput Image-based Assay to Identify Compound Activity Against Trypanosoma cruzi Amastigotes.](#) *International Journal for Parasitology. Drugs and Drug Resistance.* 2015; 5(3):215-28.
- Tegazzini D. et al. [A Replicative in vitro Assay for Drug Discovery Against Leishmania donovani.](#) *Antimicrobial Agents and Chemotherapy.* 2016; 60(6):3524-3532.

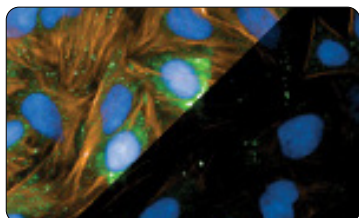
## High-content analysis and screening technologies to study infectious disease

Revvity has more than a decade of experience in developing confocal high-content screening systems. The Opera Phenix™ Plus and Operetta CLS™ systems have been fully designed in-house, which allowed control of all design aspects and technologies. Besides combining market leading automated microscopy and image analysis performance, special attention was paid to the requirements of live cell experiments from light management and environmental control, to the ease of cleaning and minimizing of risk of internal contamination of the system.



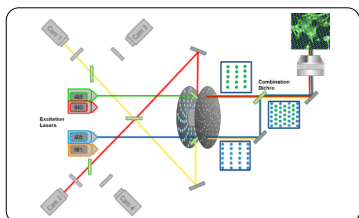
### Confocal imaging

Spinning disk confocality is a fast and gentle technique to reject light from out of focus planes, allowing live cell imaging with minimal photobleaching and phototoxicity. This option gives full flexibility to optimize image quality, depending on the size and morphology of intracellular and extracellular pathogen samples.



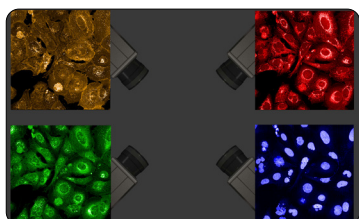
### Water immersion objectives

Proprietary automated water-immersion objectives with very high numerical aperture deliver and capture more photons allowing imaging of weakly stained samples and provide a higher resolution in XYZ than conventional objectives. This enables good image quality from tiny objects like pathogens inside cells.



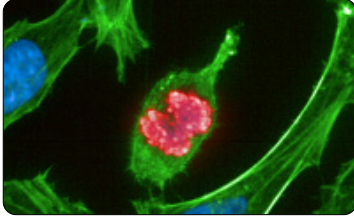
### Synchrony optics

The proprietary Synchrony Optics™ technology reduces spectral crosstalk during simultaneous imaging on multicamera Opera Phenix Plus systems. This increases speed up to 4-fold, allowing genome-wide siRNA screens or large compound screens to be done in reasonable time without compromising on sensitivity.



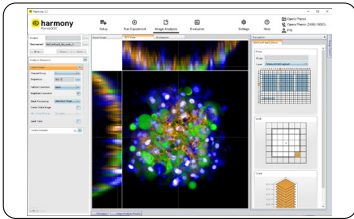
### sCMOS cameras

The large format sCMOS cameras – one in the Operetta CLS and up to four in the Opera Phenix Plus – deliver low noise, wide dynamic range and high resolution – perfect for capturing large numbers of cells, e.g. to detect infection rates with high sensitivity.



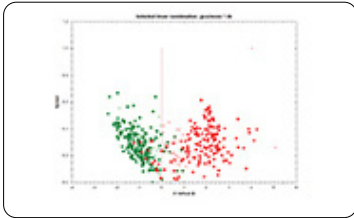
## Intelligent image acquisition

Intelligent acquisition technology – PreciScan™ – employs a pre-scan at low magnification to identify objects of interest and a re-scan of these objects at higher magnification, limiting the acquisition to just part of the wells or particular wells of interest. This saves time and hard drive space when analyzing rare events like low multiplicities of infection (MOI).



## Harmony software building blocks

Revvity's Harmony® software is known for its easy-to-use building blocks, which allow step-by-step image analysis, making it easy for even novice users to generate results, without prior image analysis knowledge. The Operetta CLS and the Opera Phenix Plus are therefore ideally suited for multi-user facilities as new scientists can quickly become familiar with the system and software.



## Machine learning

Machine learning techniques, either supervised or unsupervised, enable the user to distinguish phenotypes using classifiers based on feature combinations rather than on a single parameter or allow image segmentation using a learn-by-example approach. While other systems may require an image analysis expert to create an algorithm, machine learning makes it easy to do it on your own.



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