



AlphaPlex-545 (Tb) Human ICAM-1 Detection Kit

Product number: AP282TB-HV/C/F

Research Use Only. Not for use in diagnostic procedures.

Product Information

Application: This kit is designed for the quantitative determination of Intercellular Adhesion Molecule 1 (ICAM-1) in cell culture supernatant and human serum using a homogeneous AlphaPlex 545 assay - Terbium (no wash steps). The assay shows negligible cross-reactivity with other subtypes and species of ICAM-1.

Sensitivity: Lower Detection Limit (LDL): 4.57 pg/mL
Lower Limit of Quantification (LLOQ): 15.36 pg/mL
EC₅₀: 14.53 ng/ml

Dynamic range: Kit designed to detect [ICAM-1] between: 4.57 – 100,000 pg/mL (Figure 1).

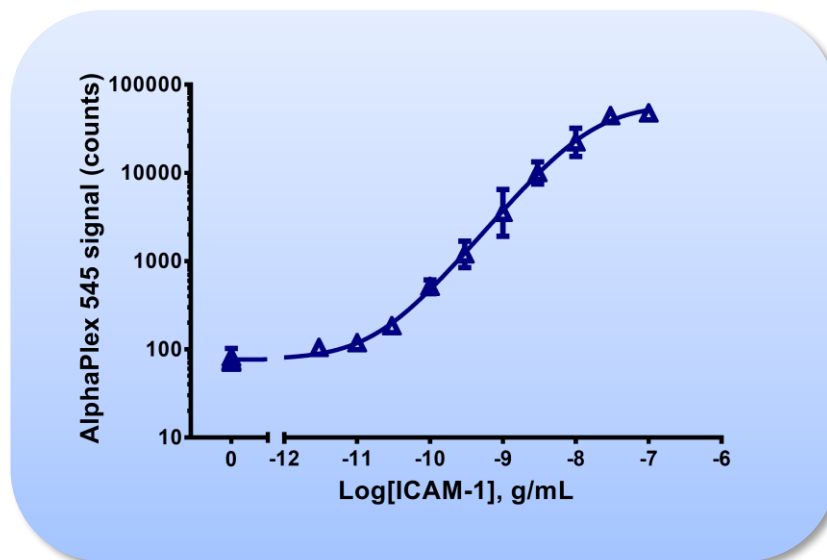


Figure 1. Typical sensitivity curve in AlphaLISA Immunoassay Buffer. The data was generated using a white Optiplate™-384 microplate and the EnVision® Multilabel Plate Reader 2103 with Alpha option.

Storage: Store kit in the dark at +4°C. Once reconstituted, the human ICAM-1 analyte is stable for at least 75 days at -20°C. Limit the number of freeze-thaw cycles.

Stability: This kit is stable for at least 6 months from the manufacturing date when stored in its original packaging and the recommended storage conditions.

Analyte of Interest

Intercellular Adhesion Molecule-1 (ICAM-1) is a dimeric transmembrane glycoprotein with a molecular weight of 80 - 114 kDa. ICAM-1 is a member of the immunoglobulin family and can be expressed on non-hematopoietic cells, such as vascular endothelial cells. It is involved in the transmigration of leukocytes to sites of inflammation. A soluble form of ICAM-1 (sICAM-1) has been identified in serum and cerebrospinal fluid, and it contains the five extracellular Ig-domains of ICAM-1. sICAM-1 is an inflammatory marker that has been associated with several common diseases such as diabetes, heart disease, stroke, and malaria. High levels of sICAM-1 have been found in serum from patients with cardiovascular disease, cancer, HIV-1, and autoimmune diseases. sICAM-1 is able to competitively bind the ligands of membrane-bound ICAM-1, such as LFA-1, Mac-1, and human rhinovirus.

Description of the AlphaPlex 545 Assay

AlphaPlex 545 technology allows for the detection of molecules of interest in buffer, cell culture media, serum and plasma in a highly sensitive, quantitative, reproducible and user-friendly mode. In an AlphaPlex 545 assay, a biotinylated Anti-Analyte Antibody binds to the Streptavidin-coated Alpha Donor beads, while another Anti-Analyte Antibody is conjugated to AlphaPlex 545 Acceptor beads. In the presence of the analyte, the beads come into close proximity. The excitation of the Donor beads provokes the release of singlet oxygen molecules that triggers a cascade of energy transfers in the Acceptor beads, resulting in a sharp peak of light emission at 545 nm (Figure 2). Combining this assay with an AlphaLISA or AlphaPlex 645 - based kit will allow the quantification of 2 (or more) analytes in the same well.

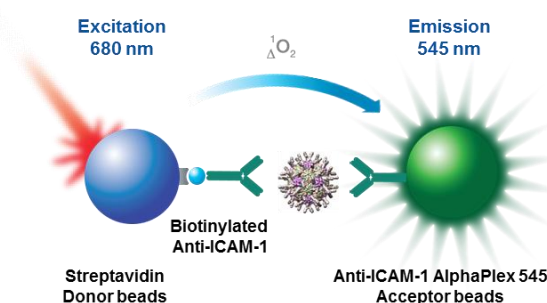


Figure 2. AlphaPlex 545 assay principle.

Precautions

- The AlphaScreen® Donor beads are light-sensitive. All the other assay reagents can be used under normal light conditions. All Alpha assays using the Donor beads should be performed under subdued laboratory lighting (< 100 lux). Green filters (LEE 090 filters (preferred) or Roscolux filters #389 from Rosco) can be applied to light fixtures.
- All blood components and biological materials should be handled as potentially hazardous. The analyte included in this kit is from a source.
- Some analytes are present in saliva. Take precautionary measures to avoid contamination of the reagent solutions.
- The Biotinylated Anti-Analyte Antibody contains sodium azide. Contact with skin or inhalation should be avoided.

Kit Content: Reagents and Materials

| Kit components | AP282TB-HV (100 assay points ^{***}) | AP282TB-C (500 assay points ^{***}) | AP282TB-F (5000 assay points ^{***}) |
|---|---|--|--|
| AlphaPlex 545 Anti-ICAM-1 Acceptor beads stored in PBS, 0.05% Kathon, pH 7.2 | 20 µL @ 5 mg/mL (1 brown tube, <u>white</u> cap) | 50 µL @ 5 mg/mL (1 brown tube, <u>white</u> cap) | 500 µL @ 5 mg/mL (1 brown tube, <u>white</u> cap) |
| Streptavidin (SA)-coated Donor beads stored in 25 mM HEPES, 100 mM NaCl, 0.05% Kathon, pH 7.4 | 40 µL @ 5 mg/mL (1 brown tube, <u>black</u> cap) | 200 µL @ 5 mg/mL (1 brown tube, <u>black</u> cap) | 2 mL @ 5 mg/mL (1 brown tube, <u>black</u> cap) |
| Biotinylated Anti-ICAM-1 Antibody stored in PBS, 0.1% Tween-20, 0.05% NaN ₃ , pH 7.4 | 20 µL @ 500 nM (1 tube, <u>black</u> cap) | 50 µL @ 500 nM (1 tube, <u>black</u> cap) | 500 µL @ 500 nM (1 tube, <u>black</u> cap) |
| Human ICAM-1 peptide | 1 µg, lyophilized * (1 tube, <u>clear</u> cap) | 1 µg, lyophilized * (1 tube, <u>clear</u> cap) | 1 µg, lyophilized * (1 tube, <u>clear</u> cap) |
| AlphaLISA Immunoassay Buffer (10X) ** | 2 mL, 1 small bottle | 10 mL, 1 small bottle | 100 mL, 1 large bottle |

* Please note that one ICAM-1 analyte vial contains an amount of ICAM-1 sufficient for performing 10 standard curves. Additional vials can be ordered separately (cat # AL282S).

** Extra buffer can be ordered separately (cat # AL000C: 10 mL, cat # AL000F: 100 mL).

*** The number of assay points is based on an assay volume of 100 µL in 96-well plates (AP282TB-HV) or 50 µL in 96- or 384-well assay plates using the kit components at the recommended concentrations.

Sodium azide should **not** be added to the stock reagents. High concentrations of sodium azide (> 0.001 % final in the assay) might decrease the AlphaPlex 545 signal. Note that sodium azide from the Biotinylated Antibody stock solution will not interfere with the AlphaPlex 545 signal (0.0001% final in the assay).

Specific additional required reagents and materials:

The following materials are recommended:

| Item | Suggested source | Catalog # |
|----------------------------------|------------------|-----------|
| TopSeal™-A Adhesive Sealing Film | Revvity Inc. | 6050195 |
| EnVision®-Alpha Reader | Revvity Inc. | - |

Recommendations

General recommendations:

- The volume indicated on each tube is guaranteed for single pipetting. Multiple pipetting of the reagents may reduce the theoretical amount left in the tube. To minimize loss when pipetting beads, it is preferable not to pre-wet the tip.
- Centrifuge all tubes (including lyophilized analyte) before use to improve recovery of content (2000g, 10-15 sec). Re-suspend all reagents by vortexing before use.
- Use Milli-Q® grade H₂O (18 MΩ•cm) to dilute 10X AlphaLISA Immunoassay Buffer.
- When diluting the standard or samples, change tips between each standard or sample dilution. When loading reagents in the assay microplate, change tips between each standard or sample addition and after each set of reagents.
- When reagents are added to the microplate, make sure the liquids are at the bottom of the well.
- Small volumes may be prone to evaporation. It is recommended to cover microplates with TopSeal-A Adhesive Sealing Films to reduce evaporation during incubation. Microplates can be read with the TopSeal-A Film.
- AlphaPlex 545 signal is detected using an EnVision Multilabel Reader equipped with the Alpha option using the following settings: Total Measurement Time: 550 ms, Laser 680 nm Excitation Time: 180 ms, Mirror: 640as (Barcode# 444), Emission Filter: Wavelength 570nm, bandwidth: 100nm, Transmittance 75%, (Barcode# 244).
- The standard curves shown in this technical data sheet are provided for information only. A standard curve must be generated for each experiment. The standard curve should be performed in AlphaLISA Immunoassay buffer.

Assay Procedure

IMPORTANT: PLEASE READ THE RECOMMENDATIONS BELOW BEFORE USE

- The manual described below is an **example** for generating one standard curve in a 50 µL final assay volume (48 wells, triplicate determinations) and 452 samples. The manuals also include testing samples in 384 well plates. If different amounts of samples are tested, the volumes of all reagents must be adjusted accordingly, as shown in the table below. **** These calculations do not include excess reagents to account for losses during transfer of solutions or dead volumes.*
- The standard dilution manual is provided for information only. As needed, the number of replicates or the range of concentrations covered can be modified.
- Use of four background points in triplicate (12 wells) is recommended when LDL/LLOQ is calculated. One background point in triplicate (3 wells) can be used when LDL/LLOQ is not calculated.

| | | Volume | | | | | |
|-------------|------------------|--------|--------|---------------------|-----------------|----------------|---|
| Format | # of data points | Final | Sample | AlphaPlex 545 beads | Biotin Antibody | SA-Donor beads | Plate recommendation |
| AP282TB- HV | 100 | 100 µL | 10 µL | 20 µL | 20 µL | 50 µL | White OptiPlate-96 (cat # 6005290) White ½ AreaPlate-96 (cat # 6005560) |
| AP282TB-C | 250 | 100 µL | 10 µL | 20 µL | 20 µL | 50 µL | White OptiPlate-96 (cat # 6005290) White ½ AreaPlate-96 (cat # 6005560) |
| | 500 | 50 µL | 5 µL | 10 µL | 10 µL | 25 µL | White ½ AreaPlate-96 (cat # 6005560) White OptiPlate-384 (cat # 6007290) Light gray AlphaPlate™-384 (cat # 6005350) |
| | 1 250 | 20 µL | 2 µL | 4 µL | 4 µL | 10 µL | Light gray AlphaPlate-384 (cat # 6005350) ProxiPlate™-384 Plus (cat # 6008280) White OptiPlate-384 (cat # 6007290) |
| | 2 500 | 10 µL | 1 µL | 2 µL | 2 µL | 5 µL | Light gray AlphaPlate-1536 (cat # 6004350) |
| AP282TB-F | 5 000 | 50 µL | 5 µL | 10 µL | 10 µL | 25 µL | White ½ AreaPlate-96 (cat # 6005560) White OptiPlate-384 (cat # 6007290) Light gray AlphaPlate-384 (cat # 6005350) |
| | 12 500 | 20 µL | 2 µL | 4 µL | 4 µL | 10 µL | Light gray AlphaPlate-384 (cat # 6005350) ProxiPlate-384 Plus (cat # 6008280) White OptiPlate-384 (cat # 6007290) |
| | 25 000 | 10 µL | 1 µL | 2 µL | 2 µL | 5 µL | Light gray AlphaPlate-1536 (cat # 6004350) |

Manual for ICAM-1 AlphaPlex 545 Assay

3 Step Manual – Dilution of standards in 1X AlphaLISA Immunoassay Buffer. The manual described below is for one standard curve (48 wells). *If a different amount of samples are tested, the volumes of all reagents have to be adjusted accordingly.*

Steps for Preparing Reagents

- 1) Preparation of 1X AlphaLISA Immunoassay Buffer:
Add 5 mL of 10X AlphaLISA Immunoassay Buffer to 45 mL H₂O.
- 2) Preparation of ICAM-1 analyte standard dilutions:
 - a. Reconstitute 1 ug lyophilized ICAM-1 with 100 µl of water by gently vortexing, avoiding pipetting solution up and down to avoid bubbles.
 - b. Store unused analyte at -20°C.
 - c. Prepare standard dilutions as follows in 1X AlphaLISA Immunoassay Buffer (change tip between each standard dilution):

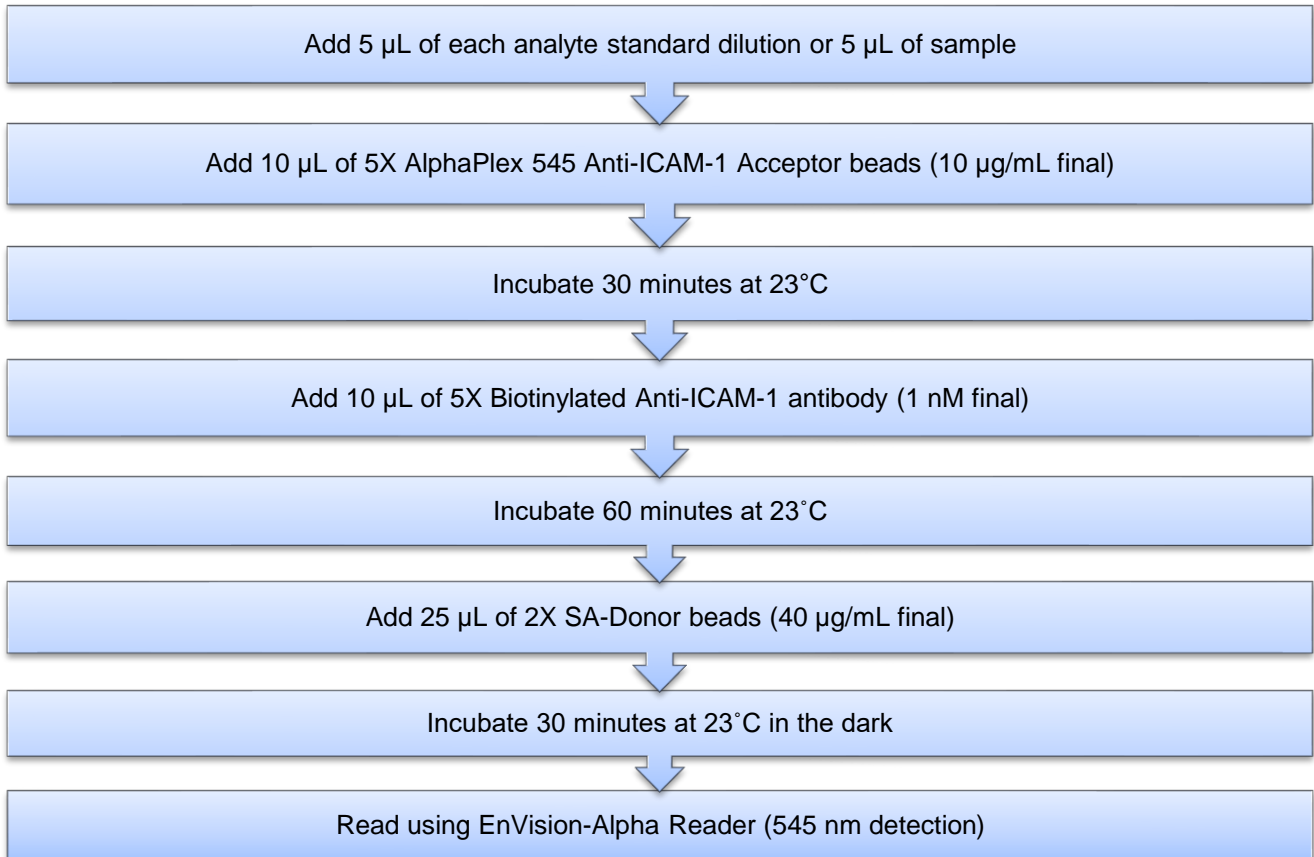
| Tube | Vol. of ICAM-1(µL) | Vol. of diluent (µL) * | [ICAM-1] in standard curve | |
|-------------------|-------------------------------|------------------------|----------------------------|-----------------|
| | | | (g/mL in 5 µL) | (pg/mL in 5 µL) |
| A | 10 µL of reconstituted ICAM-1 | 90 | 1.00E-06 | 1000000 |
| B | 60 µL of tube A | 140 | 3.00E-07 | 300000 |
| C | 60 µL of tube B | 120 | 1.00E-07 | 100000 |
| D | 60 µL of tube C | 140 | 3.00E-08 | 30000 |
| E | 60 µL of tube D | 120 | 1.00E-08 | 10000 |
| F | 60 µL of tube E | 140 | 3.00E-09 | 3000 |
| G | 60 µL of tube F | 120 | 1.00E-09 | 1000 |
| H | 60 µL of tube G | 140 | 3.00E-10 | 300 |
| I | 60 µL of tube H | 120 | 1.00E-10 | 100 |
| J | 60 µL of tube I | 140 | 3.00E-11 | 30 |
| K | 60 µL of tube J | 120 | 1.00E-11 | 10 |
| L | 60 µL of tube K | 140 | 3.00E-12 | 3 |
| M ** (background) | 0 | 100 | 0 | 0 |
| N ** (background) | 0 | 100 | 0 | 0 |
| O ** (background) | 0 | 100 | 0 | 0 |
| P ** (background) | 0 | 100 | 0 | 0 |

* Dilute standards in diluent (e.g. 1X AlphaLISA Immunoassay Buffer).
At low concentrations of analyte, a significant amount of analyte can bind to the vial. Therefore, load the analyte standard dilutions in the assay microplate within 60 minutes of preparation.

** Four background points in triplicate (12 wells) are used when LDL is calculated. If LDL does not need to be calculated, one background point in triplicate can be used (3 wells).

- 3) Preparation of 5X Anti-ICAM-1 AlphaPlex 545 Acceptor beads (50 µg/mL):
 - a. Prepare just before use.
 - b. Add 50 µL of 5 mg/mL AlphaPlex 545 Anti-ICAM-1 Acceptor beads to 4950 µl of 1X AlphaLISA Immunoassay Buffer.

- 4) Preparation of 5X biotinylated Anti ICAM-1 Antibody (5 nM):
 - a. Prepare just before use.
 - b. Add 50 μL of 500nM biotinylated Anti ICAM-1 Antibody to 4950 μL of 1X AlphaLISA Immunoassay Buffer.
- 5) Preparation of 2X Streptavidin (SA) Donor beads (80 $\mu\text{g}/\text{mL}$):
 - a. Prepare just before use.
 - b. Keep the beads under subdued laboratory lighting.
 - c. Add 200 μL of 5 mg/mL SA-Donor beads to 12 300 μL of 1X AlphaLISA Immunoassay Buffer.
- 6) In a white Optiplate (384 wells):



Read Settings: AlphaPlex 545 signal is detected using an EnVision Multilabel Reader equipped with the Alpha option using the following settings: Total Measurement Time: 550 ms, Laser 680 nm Excitation Time: 180 ms, Mirror: 640as (Barcode# 444), Emission Filter: Wavelength 570nm, bandwidth: 100nm, Transmittance 75%, (Barcode# 244).

Data Analysis

- Calculate the average count value for the background wells.
- Generate a standard curve by plotting the AlphaPlex 545 counts versus the concentration of analyte. A log scale can be used for either or both axes. No additional data transformation is required.
- Analyze data according to a nonlinear regression using the 4-parameter logistic equation (sigmoidal dose-response curve with variable slope) and a $1/Y^2$ data weighting (the values at maximal concentrations of analyte after the hook point should be removed for correct analysis).
- The LDL is calculated by interpolating the average background counts (12 wells without analyte) + 3 x standard deviation value (average background counts + (3xSD)) on the standard curve.

- The LLOQ as measured here is calculated by interpolating the average background counts (12 wells without analyte) + 10 x standard deviation value (average background counts + (10xSD)) on the standard curve. Alternatively, the true LLOQ can be determined by spiking known concentrations of analyte in the matrix and measuring the percent recovery, and then determining the minimal amount of spiked analyte that can be quantified within a given limit (usually +/- 20% or 30% of the real concentration).
- Read from the standard curve the concentration of analyte contained in the samples.
- If samples have been diluted, the concentration read from the standard curve must be multiplied by the dilution factor.

Assay Performance Characteristics

AlphaPlex 545 assay performance described below was determined using the 3 step manual.

Assay Sensitivity:

The LDL and LLOQ were calculated as described above. The values correspond to the lowest concentration of analyte that can be detected in a volume of 5 μ L using the recommended assay conditions.

| LDL (pg/mL) | LLOQ (pg/mL) | Buffer/Cell culture media | # of experiments |
|-------------|--------------|------------------------------|------------------|
| 4.57 | 15.36 | AlphaLISA Immunoassay Buffer | 10 |
| 4.85 | 16.35 | DMEM+ 10% FBS | 6 |
| 10.96 | 37.71 | RPMI+ 10% FBS | 6 |

* Note that LDL/ LLOQ can be decreased (i.e. sensitivity increased) by increasing the volume of analyte in the assay (e.g. use 10 μ L of analyte in a final assay volume of 50 μ L).

** Only the analytes were prepared in Cell Culture media. All of other components were prepared in Immunoassay Buffer.

Assay Precision:

The following assay precision data were calculated from the three independent assays using two different kit lots. In each lot, the analytes were prepared in AlphaLISA Immunoassay Buffer (IAB), DMEM medium, or RPMI medium with 10% FBS. Each assay consisted of one standard curve comprising 12 data points in triplicate and 12 background wells containing no analyte. The assays were performed in a 384-well format using AlphaLISA Immunoassay Buffer.

- Intra-assay precision:

The intra-assay precision was determined using 3 independent experiments for a total of 16 independent determinations in triplicate. CV% were calculated for each individual experiment then averaged. Shown is the average intra-experimental CV%.

| ICAM-1 | IAB | DMEM | RPMI |
|--------|-----|------|------|
| CV% | 8 | 8 | 8 |

- Inter-assay precision:

The inter-assay precision was determined using the data across 3 independent experiments with 16 measurements in triplicate. CV% was calculated by comparing the same measurement in each experiment. The CV% for all 16 measurements were then averaged. Shown is the inter-experimental CV%.

| ICAM-1 | IAB | DMEM | RPMI |
|--------|-----|------|------|
| CV% | 13 | 12 | 12 |

- Spike Recovery:

Three known concentrations of ICAM-1 were spiked into AlphaLISA Immunoassay Buffer (IAB), DMEM medium, RPMI medium, with 10% FBS. All samples, including non-spiked Immunoassay Buffers were measured in the assay. The average recovery was reported from 4 independent experiments each with 3 measurements in triplicate and compared to an IAB standard.

| Spiked ICAM-1 (ng/mL) | % Recovery | | |
|-----------------------|------------|------|------|
| | IAB | DMEM | RPMI |
| 3 | 90 | | 103 |
| 0.3 | 98 | | 110 |
| 0.03 | 110 | | 80 |

Human Serum Experiments

Human Serum (HS) was purchased and AlphaLISA Immunoassay Buffer (IAB) was used as the diluent. ICAM-1 was detected in the normal Human Serum (data not shown). ICAM-1 is expected to be present at detectable levels in HS from normal healthy subjects.

- Dilutional Linearity:

Dilutional linearity was determined by serial dilutions of Human Serum diluted 8-fold in IAB then supplemented with 50 ng/mL of ICAM-1.

| Dilution Factor | % Recovery |
|-----------------|------------|
| 1 | |
| 2 | |
| 4 | |
| 8 | |
| 16 | |
| 32 | |

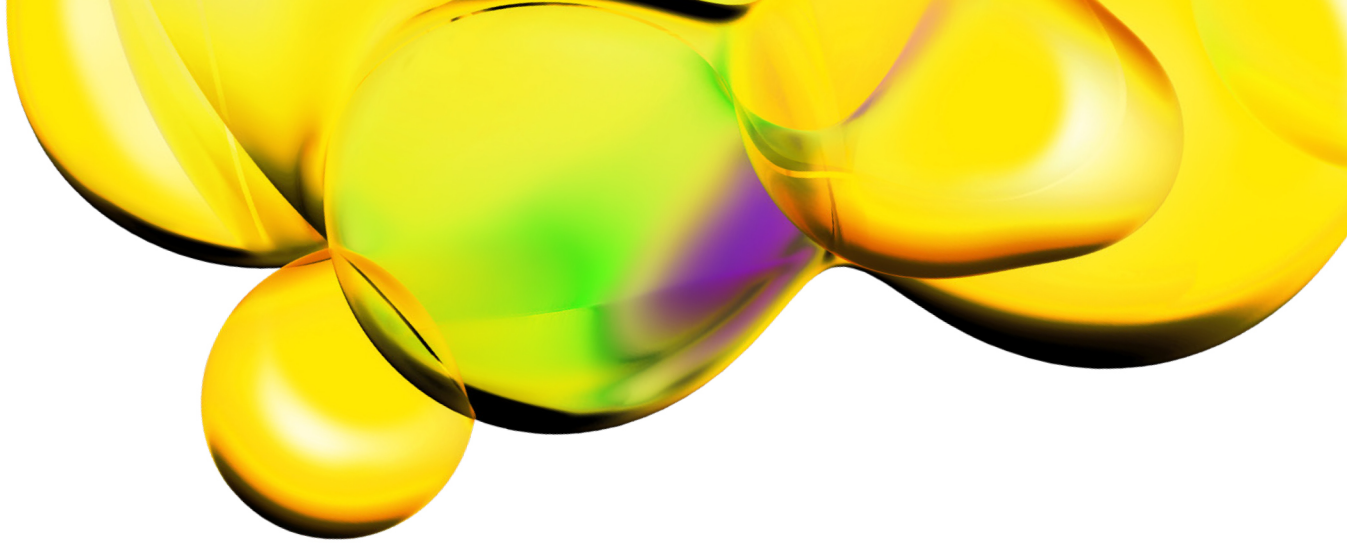
Specificity:

Cross-reactivity of the AlphaLISA ICAM-1 Kit was tested using the following proteins at 0.1 µg/mL in AlphaLISA Immunoassay Buffer

| Protein | % Cross Reactivity |
|--------------|--------------------|
| Mouse ICAM-1 | 0 |
| Rat ICAM-1 | 0 |
| Human ICAM-3 | 0 |
| Human ICAM-5 | 0 |

The possible interference from human Integrin $\alpha M\beta 2$ (Mac-1) and Integrin $\alpha L\beta 2$ (LFA-1) was investigated. The human ICAM-1 was kept at a constant concentration (EC50 value of the standard curve). The binding proteins were titrated into the assay. No interference was observed up to 10 µg/mL, which is the maximum concentration tested.

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