

Research use only. Not for use in diagnostic procedures. You are authorized to utilize these frozen cell preparations one time only. Any attempt to transfer, re-use, or propagate these cells is expressly unauthorized and a violation of the product terms and conditions of sale.

cAMPZen®

Human Prostanoid EP₂ Receptor, Frozen Cells

Product No.: ES-562-CF

Lot No.: 3156334

Material Provided

Cells: 1 x 1 mL frozen aliquot

Format: ~2.5 x 106 cells / mL in EMEM, 10% FBS, with 10 % DMSO

Product Information

Cellular Background: HEK-293

Frozen cells info: Frozen recombinant, CHO-K1 cells expressing the human

Prostanoid CRTH2 (DP2) receptor.

DNA Sequence: Identical to coding sequence of GenBank AY275471.

Corresponding Protein Sequence: Identical to Swissprot S51312

Storage Conditions: Store in liquid nitrogen (vapor phase) immediately upon receipt, or

maximum 15 days at -80°C. cAMPZen® is designed for single use

only. Do not refreeze.

Quality Control

 EC_{50} for a reference agonist is determined using a LANCE[®] Ultra cAMP assay. Mycoplasma test is performed using MycoAlert[®] Mycoplasma detection kit. We certify that these results meet our quality release criteria.

Prostaglandin E_2 - (EC₅₀): 0.092 nM

Mycoplasma: This cell line tested negative for Mycoplasma.



Recommended Thawing Conditions and Handling of Frozen Cells

- Carefully follow instructions below to obtain the expected results. Most Frozen cells are intended to be
 assayed immediately upon thawing. Exceptionally, where specified, some frozen cell products require an
 overnight incubation in Cell Medium to enable them to perform optimally.
- The recommended media catalogue number and supplier reference information are listed in this Product Technical Data Sheet (last page). Media composition is specifically defined for each cell type and receptor. The use of incorrect media or component substitutions can lead to altered product performance. Additionally, the instructions for the preparation of ligands must be carefully followed to avoid ligand precipitation, degradation or adsorption. Inappropriate preparation may result in a non-representative pharmacology.
- The complete thawing procedure must not exceed 30 min. Cell viability below 90% upon thawing may indicate that the Frozen cells were affected by incorrect thawing procedure and may yield to lower performance. Ensure the cells are not clumped and are evenly distributed in the assay plates. <u>Gently</u> pipet up and down if cells are clumped before dispensing the cells. Frozen cells <u>cannot</u> be re-frozen.

Assay Medium (for immediate thaw and use): LANCE kit Assay Buffer (see below)

Cell Medium (for overnight incubation prior to use): EMEM, Ham's F-12, 10% FBS

Thawing Cells:

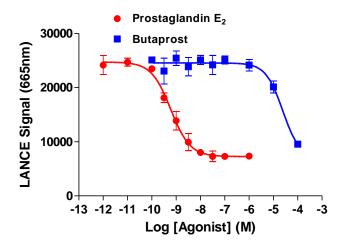
- Using appropriate personal protective equipment, rapidly place the frozen aliquot in a 37°C water bath (do not submerge) until its content is thawed completely. Immediately remove from water bath, spray aliquot with 70% ethanol and wipe excess. Under aseptic conditions using a sterile pipette, gently resuspend the cells in the cryovials and transfer content to a sterile centrifuge tube containing 10 mL of the Assay or Cell Medium prewarmed to 37°C, and centrifuge (150 x g, 5 min.). Do not exceed the recommended centrifugal force. Discard supernatant using a sterile pipette. Gently resuspend cell pellet in 5 mL of appropriate pre-warmed medium by gently pipetting up and down to break up any clumps. For immediate use, dilute cells to recommended cell density in Assay Medium.
- For an overnight incubation step, plate the cells in Cell Medium in a T25 cm² culture flask. Incubate overnight at 37°C in a humidified atmosphere with 5% CO₂. To harvest cells, under aseptic conditions, remove media, rinse with 1.5 mL of calcium and magnesium-free PBS, add 1.5 mL Versene or calcium and magnesium-free PBS/0.5 mM EDTA, and incubate at room temperature until cells detach (do not exceed 5-10 minutes). Add 3 mL of Assay Medium, collect the cells, centrifuge (150 x g, 5 min) and resuspend in Assay Medium to the recommended cell density.

Recommended Cell Density per Assay Point (LANCE®): 2500 cells/well

- Do not dilute the cells below the recommended cell density. Cell density per assay point will depend on the
 kit used to determine cAMP concentration. As a general rule, 4 to 5 times less cells are used when working
 with the cAMP LANCE® Ultra compared to when working with the cAMP LANCE® kit. The optimal cell density
 when using other cAMP kits needs to be determined.
- Ligand(s) and cells must be well mixed. When running a cAMP assay, centrifuging the plate (150 x g, 30 sec.) after addition of cells and ligands will ensure adequate mixing. If this step is omitted, the cells may not respond to the ligands as expected because insufficient contact with the ligand was made.



Typical Product Data



| Agonist | EC ₅₀ (M) | | |
|------------------------------|-------------------------|--|--|
| Prostaglandin E ₂ | 6.0 x 10 ⁻¹⁰ | | |
| Butaprost | 2.4 x 10 ⁻⁵ | | |

Figure 1: Agonist Response in LANCE® cAMP assay

An agonist dose-response experiment was performed in 384-well format using 5 000 cells/well. Cell stimulation was performed for 30 min at room temperature. Reader: EnVision $^{\circ}$. Data from a representative experiment are shown. The Z'-factor was calculated for Prostaglandin E₂ with at least 16 background and 16 maximal signal points (Z'= 0.70).



LANCE® Ultra cAMP Assay Procedure

Stimulation Buffer: HBSS, 5 mM HEPES, 0.1 % Protease-free BSA, 0.5 mM IBMX, pH 7.4.

Cells/well: For compounds not tested herein we recommend titrating the cells for optimal

performance, i.e. 500-3 000 cells per assay point.

cAMP measurements can be performed with the LANCE® *Ultra* cAMP 384 Kit (Revvity # TRF0262), according to the manufacturer instructions. Briefly:

Protocols for a 384-well white Optiplate (total assay volume of 20 µL):

| cAMP Standard curve | G _s Agonist | G _s Antagonist | G _i Forskolin titration | G _i Agonist | G _i Antagonist |
|---|-------------------------|---------------------------|---------------------------------------|-------------------------|---------------------------|
| 5 μL cAMP Standard | 5 µL cell suspension | 5 µL cell suspension | 5 µL cell suspension | 5 µL cell suspension | 5 μL cell suspension |
| 5 μL Stimulation Buffer | 5 μL Agonist | 2.5 µL Antagonist | 5 µL Forskolin | 2.5 µL Agonist | 2.5 µL Antagonist |
| _ | _ | - 2.5 µL Agonist | | 2.5 μL | 2.5 µL |
| | 2.9 μι | 2.0 µL Agoriist | | Forskolin | Forskolin/Agonist |
| Incubate 30 min at room temperature (optional step for cAMP Standard curve) | | | | | |
| 5 μL 4X Eu-cAMP Tracer Working Solution | | | | | |
| 5 μL 4X U <i>Light</i> -anti-cAMP Working Solution | | | | | |
| Incubate 1 h at room temperature | | | | | |
| Read on an EnVision® instrument. Remove microplate seal prior to reading | | | | | |

- 1. Thawed cells prepared as described above are resuspended in stimulation buffer at the desired concentration of cells/mL.
- 2. Prepare the 4X Tracer Working Solution by making a 1/50 dilution of the Eu-cAMP stock solution in the cAMP Detection Buffer.
- 3. Prepare an ULight-anti-cAMP Intermediate Solution by making a 1/10 dilution of the ULight-anti-cAMP stock solution in cAMP Detection Buffer. Prepare the 4X ULight-anti-cAMP Working Solution by making a 1/30 dilution of the ULight-anti-cAMP intermediate solution in the cAMP Detection Buffer.

Notes:

For 96- and 1536-well formats, adjust proportionally the volume of each assay component in order to maintain the volume ratios for the 384-well format. Do not modify the Eu-cAMP and/or the ULight-anti-cAMP concentrations.



LANCE® cAMP Assay Procedure

Precautions and Recommendations:

Do not vigorously vortex solutions containing cAMP antibody.

• When preparing the Detection Mix, always dilute the Eu-SA component first, and then add the Biotin-cAMP component to the Eu-SA solution.

Assay Buffer: HBSS, 5 mM HEPES, 0.1 % Protease-free BSA, 0.5 mM IBMX, pH 7.4.

Cells/well: 5000. For compounds not tested herein we recommend titrating the cells for

optimal performance, i.e. 1000-10 000 cells per assay point.

Antagonist Pre-incubation: Simultaneous addition of antagonists with reference agonist.

Agonist Stimulation: 30 min at room temperature (22°C).

cAMP measurements were performed with the LANCE® cAMP 384 Kit (Revvity # AD0263), according to the manufacturer instructions. Briefly:

1. Compounds (6 µL/well) were dispensed into a 384-well white Optiplate:

| | G_{α_S} and G_{α_i} assay modes | | G _{αs} assay mode | | Gα assay mode | | |
|------------|---|------------------------|----------------------------|------------------------|------------------------------|------------------------------------|--|
| | Basal | Forskolin | Agonist Assay | Antagonist Assay | Agonist Assay | Antagonist Assay | |
| Buffer | 6 µL | - | - | - | - | - | |
| Antagonist | - | - | - | 3 µL of 4x final conc. | - | 3 μL of 4x final conc. | |
| Agonist | - | - | 6 μL of 2x final conc. | 3 µL of 4x final conc. | 6 μL of 2x final conc. in 2x | 3 µL of 4x final conc. in 4x final | |
| Forskolin | - | 6 μL of 2x final conc. | - | - | final FK conc. | FK conc. | |

- 2. Thawed cells prepared as exposed above were resuspended in assay buffer at the concentration of 4.2×10^5 cells/mL.
- 3. The Alexa Fluor® 647-anti cAMP antibody was added 1/100 (vol/vol) to the cells suspension.
- 4. $6 \mu L/well$ of cell and antibody suspension (2 500 cells/well) were dispensed on top of the compounds prepared in the 384 well Optiplate.
- 5. After incubation for 30 min at room temperature the reaction was stopped by addition of 12 μL of Detection
- 6. The plate was incubated for 60 min at room temperature and read with an EnVision®.

Note: Assays can also be miniaturized into 1536-well format.



Materials and Instrumentation

The following tables provide the references of compounds and reagents used or recommended for the characterization of the human Prostanoid EP₂ Frozen cells, as well as some advice on how to use these compounds:

| Name | Provider | Cat no | Working Stock Solution |
|------------------------------|----------|--------|-------------------------|
| Prostaglandin E ₂ | Cayman | 14010 | 10 mM in DMSO |
| Butaprost | Cayman | 13741 | 25 mM (already diluted) |

Table 2. References of cell culture media and assay buffers.

Note: The table below lists generic media and additives typically used for Revvity Frozen cells. For product specific media and additives, please refer to the "Recommended Thawing Conditions and Handling of Frozen Cells" section.

| Name | Provider | Cat no |
|---|---------------------|------------|
| HAM's F-12 | Hyclone | SH30026.02 |
| DMEM | Hyclone | SH30022.02 |
| Advanced DMEM/F12 (serotonin receptors) | Invitrogen | 12634-010 |
| EMEM | BioWitthaker | 06-174G |
| EX-CELL DHFR ⁻ media (DHFR deficient cell lines) | Sigma | C8862 |
| FBS | Wisent | 80150 |
| FBS dialyzed | Wisent | 80950 |
| Calcium and magnesium-free PBS | GIBCO | 11010 |
| Standard HBSS (with CaCl ₂ and MgCl ₂) | GIBCO | 14025 |
| HEPES | MP Biomedicals, LLC | 101926 |
| BSA, Protease-free | Sigma | A-3059 |
| Trypsin-EDTA | Hyclone | SH30236.02 |
| Sodium Pyruvate | GIBCO | 11360 |
| L-Glutamine | GIBCO | 25030 |
| NEAA (non-essential amino acids) | GIBCO | 11140 |
| IBMX | Sigma | I-5879 |
| Forskolin | Sigma | F6886 |

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